## THE INTERACTION BETWEEN DIETARY PROTEINS AND RESISTANT STARCH ON LARGE BOWEL HEALTH

A thesis submitted to the University of Adelaide for the degree of Doctor of Philosophy

## **SHUSUKE TODEN**

**B.Sc. (Hons)** 

University of Adelaide, School of Molecular Biomedical Science,

**Discipline of Physiology** 

And

**CSIRO Human Nutrition, Adelaide** 

July 2007

## **TABLE OF CONTENTS**

DECLARATIO	Ni
ACKNOWLED	GEMENTS ii
STATEMENT (	OF AUTHORSHIP iii
ABSTRACT	vii
PUBLICATION	IS ARISING FROM THESIS xiii
PRESENTATIO	DNS xiv
ABBREVIATIO	DNSxv
CHAPTER 1: R	ESEARCH BACKGROUND1
1.1 Diet and	Bowel Health2
1.1.1 E	Bowel Diseases2
1.1.1.1	Inflammatory Bowel Diseases
1.2 Colorec	tal Cancer4
1.2.1 P	Pathogenesis of Colorectal Cancer5
1.2.2	Genomic Instability, DNA Damage and Cancer9
1.2.2.1	DNA Adducts10
1.2.2.2	DNA damage (single and double strand DNA breaks)11
1.2.3 N	Sutrition and Colorectal Cancer12
1.3 Influence	e of Dietary Protein on Large Bowel Health14
1.3.1 F	Red Meat16
1.3.1.1	Heterocyclic amines
1.3.1.2	N-nitrosocompounds
1.3.1.3	Polycyclic Aromatic Hydrocarbons19
1.3.1.4	Heme Iron19

	1.3.2	White Meat
	1.3.3	Casein
	1.3.4	Whey Protein
	1.3.5	Soy Protein
	1.4 Dieta	ry Fibre
	1.5 Resis	stant Starch
	1.5.1	Gut Microbiology
	1.5.2	Fermentation
	1.5.3	SCFA
	1.5.3.	1 Butyrate
	1.6 Color	nic Mucus Barrier35
	1.6.1	Mucins and MUC genes
	1.7 Biom	arkers of Large Bowel Health
	1.7.1	Proliferation and Apoptosis
	1.7.2	Comet Assay
	1.8 HYP	OTHESES AND AIMS41
	1.8.1	General Hypotheses
	1.8.2	Specific Aims and Objective of This Thesis41
C	CHAPTER 2	
R	RESISTANT S	STARCH PREVENTS COLONIC DNA DAMAGE INDUCED BY HIGH DIETARY
С	COOKED REI	D MEAT OR CASEIN IN RATS
C	CHAPTER 3	
D	IFFERENTL	AL EFFECTS OF DIETARY WHEY, CASEIN AND SOY ON COLONIC DNA
D	AMAGE AN	D LARGE BOWEL SCFA IN RATS FED DIETS LOW AND HIGH IN RESISTANT
S	TARCH	
C	CHAPTER 4	

DOSE-DEPENI	DENT REDUCTION OF DIETARY PROTEIN-INDUCED COI	LONOCYTE DNA
DAMAGE BY	RESISTANT STARCH IN RATS CORRELATES MORE HIGHLY	Y WITH CAECAL
BUTYRATE T	HAN WITH OTHER SHORT CHAIN FATTY ACIDS	62
CHAPTER 5		70
HIGH RED M	IEAT DIETS INDUCE GREATER NUMBERS OF COLONIC	DNA DOUBLE-
STRAND BRE	CAKS THAN WHITE MEAT IN RATS: ATTENUATED BY	HIGH AMYLOSE
MAIZE STAR(	СН	70
CHAPTER 6:	CONCLUSIONS	104
6.1 Concl	usions and Implications	105
6.1.1	Differential Effects of Various Protein Types	105
6.1.2	Protective effects of RS	108
6.1.2.1	Dose-Response Effects of RS	110
6.1.2.2	2 Oxidative Damage	110
6.1.3	Mucus Layer Thickness	111
6.1.4	Red Meat and DNA DSB	112
6.2 Future	e Directions	113
6.2.1	Animal and Human Models	113
6.2.2	Mechanisms of Protein Induced Colonic DNA Damage	115
6.2.3	Protective Mechanisms of RS	116
6.2.4	Red Meat and Colorectal Cancer Risk	117
REFERENCE	ES	

### DECLARATION

This thesis contains no material which has been accepted for the award of any other degree or diploma in any university or tertiary institution, and to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give my consent to this thesis, when deposited in the University library, being available for photocopy or loan.

The author acknowledges that copyright of published works contained within this thesis resides with the copyright holders of those works.

#### Shusuke Toden

#### ACKNOWLEDGEMENTS

I would like to start by thanking my primary supervisor Dr. Michael Conlon for guidance, encouragement and support throughout my post-graduate study. You have given me a great degree of freedom and thank you for believing in me. I would like to thank Dr. David Topping for supervising me and also developing my skill as a scientist. Furthermore, more thanks goes to Assoc. Prof. Pat Buckley, Assoc. Prof. Mike Nordstrom, Dr. Tony Bird and Prof. Tim Miles for wise advices and support.

I would also like to thank Phil Thomas for not only being a great mate but also being my role model and helping me progress as a scientist. A special thanks goes to Damien Belobrajdic, Tanya Lewanowitsch, Nathan O'Callaghan, Grant Brinkworth, Jenny McInerney, Balazs Bajka, Catherine Seccafien and Roger King for supports.

My work could not have been completed without the help of Ben Scherer, Debbie Davies, Caroline Cooke, Corinna Bennett, David Courage and Jacqui Rickard for their assistance in animal handling and biochemical assays.

This project would not have been possible without the financial assistance of CSIRO Food Futures National Research Flagship and CSIRO Division of Human Nutrition. The University of Adelaide travelling grants and Discipline of Physiology scholarships must also be acknowledged for their additional research funding.

Finally I would like to thank my family and friends for their support, in particular my grand mother for encouragements and motivations.

## STATEMENT OF AUTHORSHIP

#### **Publication 1:**

#### RESISTANT STARCH PREVENTS COLONIC DNA DAMAGE INDUCED BY HIGH DIETARY COOKED RED MEAT OR CASEIN IN RATS

#### Toden, S., Bird, A.R., Topping, D.L. & Conlon, M.A.

Cancer Biology and Therapy (2006) Mar;5(3):267-72

#### TODEN, S. (Candidate)

Designed the experiment, performed the analyses of all samples, interpreted the data and wrote the manuscript.

Signed......Date.....

#### BIRD, A.R.

Study design, data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### TOPPING, D.L.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....Date.

#### CONLON, M.A.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....Date.

#### **Publication 2:**

#### DIFFERENTIAL EFFECTS OF DIETARY WHEY, CASEIN AND SOYA ON COLONIC DNA DAMAGE AND LARGE BOWEL SCFA IN RATS FED DIETS LOW AND HIGH IN RESISTANT STARCH

#### Toden, S., Bird, A.R., Topping, D.L. & Conlon, M.A.

British Journal of Nutrition (2007) 97(3):535-543

#### TODEN, S. (Candidate)

Designed the experiment, performed the analyses of all samples, interpreted data and wrote the manuscript.

Signed	Date
~	

#### BIRD, A.R.

Study design, data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### **TOPPING, D.L.**

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### CONLON, M.A.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### **Publication 3:**

#### DOSE-DEPENDENT REDUCTION OF DIETARY PROTEIN-INDUCED COLONOCYTE DNA DAMAGE BY RESISTANT STARCH IN RATS IN MORE HIGHLY CORRELATED WITH LEVELS OF CAECAL BUTYRATE THAN OTHER SHORT CHAIN FATTY ACIDS

#### Toden, S., Bird, A.R., Topping, D.L. & Conlon, M.A.

Cancer Biology and Therapy (2006) Feb;26(2):253-258

#### TODEN, S. (Candidate)

Designed the experiment, performed the analyses of all samples, interpreted the data and wrote the manuscript.

Signed......Date.....

#### BIRD, A.R.

Study design, data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### TOPPING, D.L.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....Date.

#### CONLON, M.A.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....Date.

#### **Publication 4:**

#### HIGH RED MEAT DIETS INDUCED GREATER NUMBERS OF COLONIC DNA DOUBLE-STRAND BREAKS THAN WHITE MEAT IN RATS: ATTENUATION BY HIGH AMYLOSE MAIZE STARCH

#### Toden, S., Bird, A.R., Topping, D.L. & Conlon, M.A.

Carcinogenesis (2007) In Press

#### TODEN, S. (Candidate)

Designed the experiment, performed the analyses of all samples, interpreted the data and wrote the manuscript.

Signed	 Date

#### BIRD, A.R.

Study design, data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### **TOPPING, D.L.**

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### CONLON, M.A.

Supervised development of the work, helped in data interpretation and manuscript evaluation

I give consent for S. Toden to present this paper for examination towards the degree of Doctor of Philosophy

Signed......Date.....

#### ABSTRACT

A review of the literature revealed that diet plays an important role in serious human noninfectious large bowel diseases including cancer and inflammatory bowel diseases. Dietary protein (especially as red and processed meats) has been implicated as a positive risk factor for colorectal cancer while starch which is not digested in the small intestine (resistant starch, RS) appears to be protective. The series of experiments described in this thesis were aimed to determine the effects of dietary proteins and RS on indices of colon health in an animal model, the laboratory rat. Genetic damage is a prerequisite for carcinogenesis and this was assessed by a specific assay (the comet assay) which gives a measure of DNA strand breaks. Loss of mucus barrier function is thought to contribute to inflammatory bowel disease by permitting bacterial translocation and this was measured optically using a microscope micrometer. Other biomarkers were measured as described below. There were four major experiments.

## 1. Effects of dietary red meat and casein on colonic DNA damage and interaction with resistant starch

Previous studies had shown that higher dietary protein (as casein) induced genetic damage in rat colonocytes and that RS (fed as a high amylose maize starch) was protective. This study was aimed at establishing whether a high protein diet fed as cooked red meat had similar effects and whether RS was protective. Rats were fed diets containing either 15 % or 25% casein or 25% barbecued lean red beef, each with or without 48% high amylose maize starch (as a source of RS) for 4 weeks. As expected, high dietary casein caused a 2fold increase in colonic single-strand DNA breaks compared with a low casein diet and reduced the thickness of the colonic mucus layer by 41%. High levels of cooked meat caused 26% more DNA damage than the high casein diet but reduced mucus thickness to a similar degree as casein. Addition of RS to the diet abolished the increase in DNA damage and the loss of colonic mucus thickness induced by either high protein diet. It is thought that RS promotes large bowel health through the SCFA produced by the large bowel bacteria. One acid in particular (butyrate) has been associated particularly with promotion of normal large bowel function and protection against disease. In keeping with this hypothesis, caecal and faecal short chain fatty acid pools (including those of butyrate) were increased by inclusion of RS in the diet. DNA damage is an early step in the initiation of cancer and these findings agree with the population data which suggest that total dietary protein and red meat promote risk of colorectal cancer. However, inclusion of resistant starch in the diet could significantly reduce that risk.

## 2. Differential effects of dietary whey, soy and casein on colonic DNA damage and interaction with resistant starch

The preceding experiments showed that high levels of animal-derived proteins increased colonocyte genetic damage and loss of the mucus barrier in rats. This second experiment was designed to determine whether diets high in different types of dairy protein (casein or whey) or a plant protein isolate (soy) had similar adverse effects on colonic DNA and mucus barrier function and whether inclusion of RS in the diet was protective. Adult male Sprague Dawley rats were fed a diet containing 15 % or 25 % casein, whey or soy protein, each with or without 48 % high amylose maize starch for 4 weeks. In confirmation of the earlier studies, higher levels of dietary casein increased colonocyte DNA damage significantly. However, whey did not increase genetic damage. Colonic DNA damage

was highest for soy when fed at both 15% and 25% protein in the absence of RS. Inclusion of RS in the diet attenuated colonocyte DNA damage due to higher dietary protein in all three groups. The colonic mucus barrier was thinner in rats fed higher dietary protein but the effect was reversed by feeding RS. Caecal total SCFA and butyrate pools were low in rats fed the digestible starch and were higher in rats fed RS. However, there was no relationship between caecal or faecal SCFA and genetic damage or mucus thickness. Caecal and colonic tissue weight and colon length were higher in rats fed RS, consistent with greater SCFA supply. These data confirm that higher dietary protein of animal (casein) or plant (soy) origin increases genetic damage and loss of the mucus barrier indicating that this is an effect of protein and not its source. These findings accord with the epidemiological data which link dietary protein to greater risk of colorectal cancer and inflammatory bowel disease. However, the data show also that dietary proteins differ in their specific actions on genetic damage and mucus thickness. Further, the data from the feeding of whey suggest that not all proteins are equivalent in their capacity to provoke adverse changes in colonic integrity. While the data show that RS raised large bowel and faecal SCFA, they indicate their levels were not related directly to these biomarkers.

# 3. Dose response effects of resistant starch on protein induced colonic DNA damage

The accumulated data linking greater protein intakes to adverse changes in the colon were obtained at dietary levels which were not unreasonable in terms of animal or human consumption. However, the dietary level of RS which were fed were relatively high (48% by weight) so this study was conducted to determine its effectiveness at lower levels of

dietary inclusion. It was also important to ascertain whether there was a dose-response relationship between RS intake and the observed effects. One of the mechanisms proposed for the induction of colorectal cancer by high dietary protein intakes is oxidative damage to DNA. In this experiment, this was done by assaying with endonuclease III. Adult male rats were fed a diet containing 25% casein with 0%, 10%, 20%, 30% or 40% high amylose maize starch for 4 weeks. As in the preceding studies comet tail moment was greatest and the mucus barrier thinnest in rats fed 0% RS. DNA damage was reduced and the mucus barrier thickened in a logarithmic dose-dependent manner by RS. There was no significant difference between dietary groups associated with oxidative DNA damage as measured by endonuclease III. Caecal and faecal short chain fatty acid (SCFA) pools rose with the increased level of dietary RS. DNA damage of colonocytes correlated negatively with caecal SCFA but the strongest correlation was with caecal butyrate, which is consistent with the proposed role of this SCFA in promoting a normal cell phenotype. The data show that RS prevents protein induced colonic DNA damage in a dose-dependent manner. Inclusion of 10% high amylose maize starch was found to be sufficient to oppose colonocyte DNA damage, and to increase caecal and faecal SCFA pools. Intakes of this order are not unreasonable in terms of human consumption of RS.

# 4. Dose response effects of red and white meat on colonic DNA damage and interaction with resistant starch

The accumulated evidence from large prospective human studies links diet to colorectal cancer risk strongly. The evidence from the animal studies described in this thesis that dietary protein induces colonocyte genetic damage supports a role for high protein intakes in increasing risk. Recently, several large epidemiological studies and a meta-analysis of

prospective studies have found that consumption of dietary red or processed meats, but not white (poultry) meat, is associated with increased risk of colorectal cancer. This is consistent with the data from the preceding studies that specific proteins affected colonic integrity differentially. A large prospective European study (European Prospective Investigation into Cancer and Nutrition) has reported that dietary fibre was protective. The findings reported in this thesis that RS opposes the effects of high dietary protein accord with that conclusion. This study aimed to compare the effects of cooked red (beef) or white (chicken) meat on DNA damage and mucus barrier thickness in rats. The study was designed to determine whether the relationship between the intakes of these meats was Double-strand DNA breaks are thought to relate more closely to dose-dependent. carcinogenesis than single-strand breaks so both were measured. Adult male Sprague-Dawley rats were fed a diet containing 15%, 25% or 35% cooked beef or cooked chicken each with or without 20% high amylose maize starch for four weeks. Both red and white meat increased colonic DNA damage dose-dependently. However, both single and double strand breaks were significantly greater when the rats were fed the red meat diets compared to those fed the white meat. Colonocyte DNA damage was reduced by the consumption of RS while large bowel SCFA were increased. The findings of this study are consistent with the epidemiological data which show that red meat consumption is associated with greater risk of colorectal cancer but that white meat is not.

#### Summary

The data reported in this thesis support the findings of prospective population studies that high dietary protein, red meat in particular, appears to be harmful to the health of the large bowel. However, the data demonstrate also that different protein types have differential effects on the integrity of the colonocyte DNA. Furthermore, the addition of RS to the diet protects against protein-induced colonic DNA damage and maintenance of the colonic mucus barrier, apparently through increased SCFA production by colonic fermentation. The results of these experiments indicate a strong potential for RS to be effective in maintenance of large bowel integrity in the face of high dietary protein.

### **PUBLICATIONS ARISING FROM THESIS**

**Toden, S.**, Bird, A.R., Topping, D.L. & Conlon, M.A. High Red Meat Diets Induce Greater Numbers of Colonic DNA Double-Strand Breaks than White Meat In Rats: Attenuation by High Amylose Maize Starch. Carcinogenesis 2007 *In Press* 

**Toden, S.**, Bird, A.R., Topping, D.L. & Conlon, M.A. Dose-dependent reduction of dietary protein-induced colonocyte DNA damage by resistant starch in rats is more highly correlated with levels of caecal butyrate than other short chain fatty acids. Cancer Biology and Therapy Feb; 26(2):253-258, 2007

**Toden, S.**, Bird, A.R., Topping, D.L. & Conlon, M.A. Differential Effects of Dietary Whey, Casein and Soy on Colonic DNA Damage in Rats. British Journal of Nutrition 97(3):535-543, 2007

**Toden, S.**, Bird, A.R., Topping, D.L. & Conlon, M.A. Resistant starch prevents colonic DNA damage induced by high dietary cooked red meat or casein in rats. Cancer Biology and Therapy Mar;5(3):267-72, 2006

**Toden, S.**, Bird, A. R., Topping, D. L., and Conlon, M. A. Differential effects of dietary whey and casein on colonic DNA damage. Aust. J. Dairy Technol. 60, 146-148 2005

### PRESENTATIONS

#### **Conference Abstracts**

Conlon, M.A., **Toden, S.**, Bird, A.R., Topping, D.L. Resistant starch and colonic mucosal integrity. Asia Pacific Journal of Clinical Nutrition **15** *Suppl:* S45, 2007

**Toden, S.**, Bird, A. R., Topping, D. L., and Conlon, M. A. Resistant starch protects against colonic DNA damage induced by dietary whey, soy and casein in rats Dietary Fibre 2006 Multifunctional Complex of Components 160-161, 2006

Topping, D.L., Bird, A.R., Regina, A., **Toden, S.**, Rahman, S., Keogh, J., Conlon, M.A., Morell, M.K. New cereal foods for improved human health, American Association of Cereal Chemsists Symposium, San Francisco, USA, 2006

Topping, D.L., Bird, A.R., Clarke, J., Conlon, M.A., Keogh ,J., Li, J.L., Morell, M., Regina, A., Rahman, S., **Toden, S.** Engineering Starches for Foods to Improve Human Health, Asia Pacific Congress of Chemical Engineers, Kuala Lumpur, Malaysia, 2006

**Toden, S.**, Bird, A. R., Topping, D. L., and Conlon, M. A. Resistant starch attenuates colonic DNA damage induced whey, soy and casein in rats. Asia Pac J Clin Nutr, **14** *Suppl:* 2005

Topping, D. L., **Toden, S.**, Bird, A. R. and Conlon, M. A. Resistant starch and health, University of Illinois, Chicago IL, USA, 2005

Topping, D.L, Bird, A.R., **Toden, S.**, Conlon M.A., Noakes M., Morell M., Mann G., and Li, Z.L. Interaction between dietary proteins and resistant starch Making Fiber Irresistible: Resistant Starch is a Natural Seminar, Chicago IL, USA, 2005

Topping, D.L., Bird, A.R., **Toden, S**., Conlon M.A., Noakes M., King R., Morell M., Mann G., and Li, Z.L. Resistant Starch: Strategies to Increase Intakes for Health Benefit. National Starch Food Innovation, Health Professionals Symposia, Sydney and Melbourne, 2005

Topping, D., Bird, A.R., **Toden, S.**, Conlon M.A., Noakes M., King R., Morell M., Mann G., and Li, Z.L. Resistant starch as a contributor to the health benefits of whole grains. Whole Grains Summit, Minneapolis, MN, USA, 2005

## ABBREVIATIONS

ACF	aberrant crypt foci
APC	adenomatous polyposis coli
AIN	American Institute of Nutrition
ANOVA	analysis of variance
BaP	benzo(a)pyrene
cm	centi-meter
CRC	colorectal cancer
°C	degrees, Celsius
DNA	deoxyribonucleic acid
DMEM	Dulbecco's Modified Eagle's Medium
DMH	dimethylhydrazine
DTT	dithiothreitol
DSB	double-strand break
FAP	familiar polyposis
g	grams
GSH	glutathione
GST	glutathione S transferase
h	hours
HBSS	Hanks' balanced salt solution
HCA	Heterocyclic amines
HAMS	High amylose maize starch,
HDAC	histone deacetylase

IBD	Inflammatory bowel disease
М	molar
μg	micrograms
μl	milligrams
mm	millimetres
min	minutes
NOC	N-nitrosocompounds
NSP	non-starch polysaccharides
PBS	phosphate buffered saline
РАН	Poly aromatic hydrocarbons
рН	potential of hydrogen
PhIP	2-Amino-1-methyl-6-phenylimidazo[4,5-b]pyridine
RS	Resistant starch,
SCFA	short chain fatty acid
SEM	standard error of the mean
SSB	single-strand break
TCF	transcription factor
TBS	tris buffered saline
wk	weeks

## **CHAPTER 1: RESEARCH BACKGROUND**

The main goal of the work described in this thesis is to determine the effects of high protein diets and RS on large bowel health. This was achieved by comparing the differential effects of several types of dietary protein on colonic DNA damage as an indicator of potential colorectal cancer risk in rats. The potential protective effects of RS on colorectal cancer risk were examined through changes in genetic damage. The effects of the two dietary treatments (RS and dietary protein) on colonic barrier function were examined through determining changes in mucus barrier thickness. The following research background is intended to give the reader an understanding of the current state of knowledge on the actions of different types of protein on the risk of large bowel diseases. Furthermore, this review details information relating to the protective effects of dietary fibre, especially RS, and its potential protective mechanisms against large bowel diseases.

#### 1.1 Diet and Bowel Health

It is well established that there is a very strong link between diet and large bowel health. The principal function of the large bowel is to salvage water from digesta passing from the small intestine. While the absorption of nutrients is largely completed when digesta reaches the terminal ileum, there is a potential for significant dehydration without this recovery (1-3). The period of exposure of the colon wall to digesta is relatively long while this process is completed, increasing the risk of potential damage by carcinogens and toxins.

#### **1.1.1 Bowel Diseases**

There are several non-infectious large bowel diseases which have significant effects on human health. These are inflammatory bowel diseases (IBD; Crohn's disease (CD) and ulcerative colitis (UC)) and colorectal cancer (CRC). Although IBD and CRC are different diseases, it is known that patients with prolonged IBD are at increased risk for developing colon cancer (4-7). UC patients have at least 3 times higher colon cancer risk than in the general population and about 5 % of people with UC develop colon cancer (5, 6). CD patients, with at least 30% of the colon involved in disease, may have an increased risk of colorectal dysplasia and cancer (6). The risk of cancer increases with the severity of the damage to the colon and the duration of the disease. Although it is difficult to associate specific dietary components to IBD risk, studies indicate that CD is increasing in countries where a more Western diet (high in refined foods) is displacing more "traditional" diets (high in unrefined food and dietary fibre) (8).

#### **1.1.1.1 Inflammatory Bowel Diseases**

IBD are more common in Europe and North America where they affect both males and females equally (9). The disease affects people of all ages, but is more common in those aged 15 to 40 years. However, many recent reports have highlighted the rising prevalence of IBD in Asia (10-12). It is estimated that 61,000 Australians have IBD; approximately 28,000 have CD and 33,000 have UC (13). IBD is a group of inflammatory conditions of the large intestine, and in some cases the small intestine. IBD is thought to result from inappropriate and ongoing activation of the mucosal immune system driven by the presence of normal luminal flora and this response is facilitated by defects in both epithelial barrier functions and the mucosal immune system (14). Minor forms of IBD also include: collagenous colitis, lymphocytic colitis, ischaemic colitis, diversion colitis, Behcet's syndrome, infective colitis and indeterminate colitis.

The major difference between UC and CD is that, the former only affects the inner lining of the bowel wall, whereas in CD the inflammation is of the full thickness of the bowel wall and can affect the whole gastrointestinal tract. The most common symptoms of UC include: abdominal pain, blood, mucus or pus in the stool, diarrhoea, fatigue and tiredness, weight loss and loss of appetite (15). Although the cause of UC is still unknown, causative factors suggested include: genetic predisposition, infectious agents, defects in immune system and environmental factors.

As noted, CD involves any part of gastrointestinal tract, but most frequently involves the distal small bowel and colon. Inflammation can vary greatly from a small ulcer over a lymphoid follicle to an extensive chromic inflammation. This inflammation is transmural, and can result in strictures, micro-perforations, and fistulae. CD is often associated with autoimmune disorders outside the bowel, such as aphthous stomatitis and rheumatoid arthritis (16).

#### 1.2 Colorectal Cancer

Western countries including Europe, America and Australia have the highest incidence of colon cancer and account for nearly two-thirds of the approximately one million cases that occur worldwide each year (17). However, many Asian countries including, Japan, China and South Korea have experienced a two to four fold increase in the incidence of CRC in recent decades (18, 19). CRC is the second most common cause of cancer-related death in both men and women in western countries (20). In Australia,

CRC was the second leading type of cancer death in 2004, second only to skin cancer (21, 22).

Genetic factors are thought to be important in CRC development in individuals (23). Among first-degree relatives of colon cancer patients, the lifetime risk of developing CRC is 18%, a 3-fold increase over the general population (21). However, 80% of CRC occur sporadically in patients with no family history of CRC, indicating environment has a large bearing on development of CRC. Diet is considered to be the main environmental factor, with approximately 70% of CRC attributed to an inappropriate diet (23, 24). DNA damage is a prerequisite for cancer induction and it is established that specific food ingredients or their metabolic by-products are genotoxic (25-27). Other environmental factors include viral infections, chemical carcinogens, and radiation. Viral infections and radiation exposure are rare causes of CRC, while dietary carcinogens are much more important because of their direct exposure to the gut.

#### 1.2.1 Pathogenesis of Colorectal Cancer

Cancer is a complex multi-step process and thought to be caused by an accumulation of multiple genetic mutations resulting in a transformed phenotype (28). Genomic instabilities caused through mutation or lack of DNA damage repair can lead to cancer formation (29, 30). There are two major possible processes that may be responsible for inducing genetic mutations in colonocytes: 1) the direct action of a mutagen causing genetic damage or 2) increased proliferation causing an increased risk of errors in DNA repair. A cell accumulating these genetic mutations will likely to undergo full malignant transformation (31). This concept, known as the "multi-step model of carcinogenesis" is

described in detail below (Fig. 1.1).

NOTE: This figure is included on page 6 of the print copy of the thesis held in the University of Adelaide Library.

Figure 1.1 Multiple steps of colorectal carcinogenesis. Significant genes involved in gene regulation in carcinogenesis include adenomatous polyposis coli (APC), K-ras and p53 (Modified from Sharma *et al.* 2001)

Initiation, the first stage of colorectal carcinogenesis, is often associated with mutation in the adenomatous polyposis coli (APC)/ $\beta$ -catenin pathway or mismatch repair genes. Both genes play a pivotal role in the regulation of mucosal proliferation and are classic tumour suppressor genes. Mutations in the APC gene on chromosome 5q21 locus are found in 60 to 80% of sporadic CRC and adenomas (32). In sporadic CRC, mutation in

**Research Background** 

APC initiate the majority of the tumours (33). Mutations in the APC gene are responsible for the disease familiar polyposis (FAP), where patients develop numerous benign tumours of the colon. Some of these tumours will progress to cancer if not removed surgically. APC is known to regulate  $\beta$ -catenin-TCF or Wnt signalling. The APC protein interacts in a complex with  $\beta$ -catenin, glycogen synthase kinase 3 $\beta$  and axin to regulate the levels of  $\beta$ catenin by targeting  $\beta$ -catenin for degradation by the ubiquitination-proteosome pathway (34). In the absence of functional APC,  $\beta$ -catenin levels rise, enabling it to form an active complex with the transcriptional factor TCF-4 (35). The  $\beta$ -catenin-TCF-4 transcription complex targets c-myc (36), which activates the transcription of the cyclin-dependent kinase, cdk 4 and consequently increases cell proliferation (37).

A number of different agents have been found to cause genetic damage to colonocytes. Some of the known agents include dietary contaminants, chemical mutagens, pathogenic bacteria and viruses. Mutations may not necessarily lead to the development of cancer as apoptosis, programmed cell death, can remove cells with genetic mutations (38). In addition, there are many non-genotoxic agents that are associated with increased risk of CRC. These agents include hormones, drugs, physical or mechanical trauma and other chronic irritants (39). Increased cell proliferation is a common histological change induced by these agents which can potentially lead to carcinogenesis.

The K-ras oncogene is thought to be involved in transition from intermediate adenomas to carcinomas in sporadic CRC (33). The K-ras gene product, located on the inner plasma membrane, is involved in the transduction of mitogenic signals. The ras protein is activated transiently as a response to extra-cellular signals that stimulates cell surface receptors (40).

The final stage of carcinogenesis is progression. It involves additional growth of

the adenoma and invasion of the basement membrane. Loss of p53, a tumour suppression gene, is the major genetic change associated with progression of tumour growth to carcinoma. The p53 gene is known to arrest cell cycle progression, increase apoptosis and allow the DNA to be repaired. In cancer cells with mutated p53, cell proliferation is no longer controllable, resulting in inefficient DNA repair and the emergence of genetically unstable cells (41).

CRC can be viewed as a process of polyp formation. Although benign polyps do not invade nearby tissue or spread to other part of the large intestine, they can become malignant over time. The figure below shows the development of CRC through a series of histological stages from adenocarcinoma to carcinoma (42) (**Fig. 1.2**).

NOTE: This figure is included on page 8 of the print copy of the thesis held in the University of Adelaide Library.

Figure 1.2 Multi-stage model of rat colon cancer. (Based on Nakagama et al 2005)

**Research Background** 

#### 1.2.2 Genomic Instability, DNA Damage and Cancer

Recent studies have suggested that the link between DNA damage and early stage cancer is strong (43-46). DNA damage occurs all the time and the majority of time this damage is repaired by DNA repair mechanisms. Therefore, DNA repair is an essential process for cell survival because it protects the genome from potentially harmful mutations. However, accumulation of this damage can lead to activation of proto-oncogenes and inactivation of tumour suppressor genes, which may lead ultimately to cancer formation. Tumourigenesis driven by genetic damage is fuelled by DNA damage and errors made by the DNA machinery (47).

Genomic instability can take two main forms; microsatellite instability and chromosomal instability (48). Microsatellites are stretches of DNA in which a short motif is repeated several times. The most common microsatellite in the humans is a dinucleotide repeat of CA which occurs frequently across the genome (49). Microsatellite instability occurs when a germline microsatellites allele has gained or lost repeated units and thus undergone a somatic change in length (49). Chromosomal instability, or gross rearrangement of chromosomes, occurs when the cells show aneuploidy, a chromosomal state in which abnormal numbers of chromosome sets exist within the nucleus with chromosomal breaks and other defects (50). Furthermore, point mutations, changes in a single base pair, can affect a range of genes with strong contributory links to CRC. As noted previously, these include the oncogene K-ras, and tumour suppressor gene p53. Point mutations can arise through various mechanisms. Spontaneous mutation may occur due to the instability of purine and pyrimidine bases or defects in mismatch repair (51). Common chromosomal aberrations include the loss or gain of whole chromosomes or

chromosome fragments and the amplification of chromosome segments (52). Loss of large regions of chromosomes can lead to the inactivation of tumour suppressor genes, whereas amplification of chromosomal regions might promote tumourigenesis by activation of proto-oncogenes (53).

#### 1.2.2.1 DNA Adducts

DNA adduct formation is another mechanism of mutagenesis. A DNA adduct is the covalent linking of an abnormal radical to DNA with the potential for cancer formation (main carcinogens include: N-nitrosamines, aflatoxins, aromatic amines, and polycyclic aromatic hydrocarbons) (54, 55). The misreplication of DNA adducts can lead to malignancy. Alkylating or oxidative agents cause the majority of documented adducts. Recently, there have been speculations that these adducts are linked to greater CRC risk O<sup>6</sup>-methylguanine DNA adducts are formed between nitrosated glycine (56, 57). derivatives and DNA (56, 58). Glycine, a common dietary amino acid, opens the possibility that its nitroso products could be major alkalylating agents in the human gastrointestinal tract (59, 60). Furthermore, recent in vitro studies have demonstrated that formation of O<sup>6</sup>-methylguanine DNA adducts lead to mutations in p53 similar to those observed in human gastrointestinal tract tumours (61). A study in human volunteers has shown that dietary red meat consumption led to significantly higher O<sup>6</sup>-methylguanine DNA adducts than white meat in colonocytes (57). In addition, a study in rodents treated with a carcinogen (AOM) showed that a combination of fish oil and pectin (a highly fermentable fibre) reduced the amount of O<sup>6</sup>-methylguanine DNA adducts (62). Collectively, these data suggest that adduct formation is a potential mechanism for

carcinogenesis which can be modulated by diet to increase or decrease risk.

#### **1.2.2.2 DNA damage (single and double-strand DNA breaks)**

Chromosome breaks are a documented measure of DNA damage with single-strand DNA breaks (SSB) being one of those measured most frequently. SSB arise constantly from DNA base damage and must be repaired to maintain genomic stability. If the repair process is hindered, this can lead to double-strand breaks during DNA replication and can result in chromosome instability and cell death. DNA double-strand breaks (DSB) are considered to be one of the most dangerous forms of cellular genomic damage. DSB differ from other types of DNA lesion in that both strands of the double helix are damaged which prevent the use of the complementary DNA strands as a template for repair. The accumulation of DNA damage can lead to cancer or serious impairment of cellular functions (**Fig. 1.3**). DSB result from exogenous agents such as ionising radiation and certain chemotherapeutic drugs, endogenously generated reactive oxygen species, mechanical stress on the chromosome and specialized recombination reactions (52, 63).

To counteract DNA damage, all organisms possess DNA repair mechanisms that correct specific types of lesions. Extensive damage can lead to cell death. Cells can enter 'replicative death', a state of irreversible growth arrest, or trigger apoptosis (52). Other protective mechanisms include cell-cycle check points and DNA-damage repair. An epidemiologic study showed consistent and positive associations between suboptimal DNA repair capacity and cancer occurrence (64).

NOTE: This figure is included on page 12 of the print copy of the thesis held in the University of Adelaide Library.

Figure 1.3 Causes, cellular responses and consequences of DNA double-strand breaks (based on van Gent et al. 2001)

#### **1.2.3 Nutrition and Colorectal Cancer**

The relationship between specific dietary components and cancer development and

**Research Background** 

progression has been of great interest for decades. The complexity of diets and the fact that dietary components may modify one or many steps in the cancer process makes simple recommendations difficult (65, 66).

Epidemiological studies indicated that a diet rich in fruits and vegetables is associated with lower incidence of various forms of cancer (67, 68). Many components of fruits and vegetables may be responsible for their protective effect; such as micronutrients, phytochemicals and fibre. Deficiency in micronutrients including folic acid, Vitamin B12, B6, C, E, iron, niacin and zinc have been shown to mimic radiation by causing DNA damage (69-72). Major phytochemicals found in fruits and vegetables are terpenoids (including carotenoids), phenolics, nitrogen containing alkaloids and sulphur compounds (73). Many carotenoids are antioxidants, which can help to protect the body against damage caused by oxygen free radicals and phenolic compounds are known to provide protection via manipulation of phase I and II enzymes (73).

Although there is no internationally accepted definition of dietary fibre, its composition and actions may be described as:

Dietary fibre means that fraction of the edible part of plants or their extracts or synthetic analogues that -(a) are resistant to digestion and absorption in the small intestine, usually with complete or partial fermentation in the large intestine; and (b) promote one of the following beneficial effects -(i) laxation (ii) reduction in blood cholesterol (iii) modulation of blood glucose and, (c) includes polysaccharides, oligosaccharides (degree of polymerisation >2) and lignins. (74)

This definition is important as it shows its range and complexity. Although there have been mixed views on the protective effects of fibre on CRC risk (51, 75-80), recent studies indicate that there is a strong inverse link between fibre consumption and CRC risk (78,

81, 82). A large European prospective study, involving over 500,000 subjects, has shown an inverse relationship between dietary fibre intake and the incidence of large bowel cancer (81). Moreover, a majority of correlative studies in humans showed either a strong or moderate protective effect of dietary fibre or fibre-rich foods on CRC (83). Another recent epidemiological study from Japan, utilising data from over 40,000 subjects, found a decreasing trend in risk of CRC with increasing intake of total dietary fibre (82).

In contrast, epidemiological studies indicate that a diet high in fat and dietary protein may increase risk (84, 85). Present data have linked a high dietary intake of  $\omega$ polyunsaturated fatty acids (PUFA) such as linoleic acid to increase risk of CRC (86). Moreover, human and animal studies have demonstrated a positive association between CRC risk and diets high in red or processed meat (84, 87-91). The evidence from both human and animal experiments regarding the influence of dietary protein on cancer risk, both quantitatively and qualitatively, suggest the quality of protein had no influence on colon cancer risk, while increasing concentrations of protein in the diet did significantly increase risk (92). Another rodent study using carcinogens to induce tumours showed that protein type influenced CRC risk (93).

This thesis focused on two major nutritional components on the large bowel health: dietary protein and fibre.

#### 1.3 Influence of Dietary Protein on Large Bowel Health

In recent years, the increase in obesity has become an enormous concern for the communities around the world. This has lead increasing numbers of people to adopt high protein/low carbohydrate diets for weight control purposes. Although there is strong

evidence that this type of dietary method helps weight loss, a greatly increased consumption of protein may raise concern for large bowel health.

In general, a high fat diet is considered to be harmful for bowel health (94, 95). However, the effects of dietary protein on bowel health are controversial (96). One report suggested that high protein diets (24% of energy intake) could offer significant reductions in risk factors for heart disease as well as enabling weight reduction (97). It is known that increased dietary protein intake enables more undigested protein from the small intestine to reach the large intestine. This had led to the development of the concept of resistant protein by analogy with RS (98). A recent rodent study, however, demonstrated that resistant protein promoted its fermentation which enhanced tumourigenesis (99).

A meta-analysis of prospective population studies identified that protein *per se* was an independent risk factor for CRC (100). Another study suggested that high protein consumption may be harmful for bowel health due to the increased flow of nitrogen to the gut (101). Approximately 2 g of nitrogen enters the large bowel daily mainly in the form of protein, peptides and amino acids, some of which is from endogenous sources. Nitrogen flow from the ileum can be increased by greater protein intake, heat treatment of dietary proteins (to reduce digestibility) and the physical form of foods (101, 102). The main product of bacterial metabolism of nitrogenous residues is ammonia. Ammonia has been suggested to promote tumorigenesis by stimulating cell proliferation (103), which favours the growth of malignant cells (104). In any event, where carbohydrate is rapidly depleted and becomes unavailable, the carbohydrate-to-nitrogen content of the colon decreases and fermentation becomes 'more and more' proteolytic. This results in an increase in ammonia production (105). High dietary protein consumption would aggravate the situation by contributing to the production of amines and ammonia (106). In addition, diets high in

meat (such as those recommended for weight loss) generally provide relatively little starch and NSP. This can also lead to increase in ammonia concentrations partly by an increase in available nitrogen and limited fermentation which lowers the bacterial demand for growth (107). Therefore, undigested dietary protein may also contribute to the increase of ammonia in the colon. It has been estimated that with a typical Western diet, up to 12 g of protein per day can escape digestion (108).

There are many different sources of dietary protein and most have different amino acid compositions. It follows that, if the amino acid profile is a factor in CRC risk, protein sources may differ. Furthermore, the methods of preparation (including cooking) of these protein sources must also be considered. One study demonstrated that different cooking methods have a significant influence on large bowel health (109). Generally, it is thought that broiling may be especially deleterious through the formation of heterocyclic amines (ie potential tumourigenic agents). The following section of this thesis explores the differences between various protein sources and possible mechanisms in which these protein sources may influence large bowel health.

#### 1.3.1 Red Meat

Red meat refers to mammalian meat that appears red before cooking (although the actual colour intensity may vary). This includes beef, veal, lamb, pork and buffalo meat. In contrast, animal proteins derived from birds, fish, crustaceans and reptiles are not red meat, although some do appear reddish in colour. It is the presence of myoglobin, which is an intramuscular oxygen carrier, that gives red meat its characteristic colour.

The effect of red meat on large bowel health has been established rather more

clearly than for other dietary protein sources (87). Red meat has been proposed as a factor likely to increase the risk of CRC through several mechanisms (110). High meat consumption is known to increase levels of amino compounds and nitrosamines. It also alters the large bowel flora and changes enzyme patterns (111, 112). These factors may be either cytotoxic or genotoxic themselves or increase the formation of genotoxic compounds in the gut lumen (113). A rat study demonstrated an increase of 33-59% in the incidence of mature rats with dimethylhydrazine-induced intestinal tumours when barbecued beef was substituted for whey protein concentrate against a high fat diet background (96). Significantly increased urinary phenol and *p*-cresol concentrations were also observed with the 'beef fed' rats relative to 'whey protein fed' rats. Phenol and pcresol have been identified as toxic/mutagenic metabolites associated with increased risk of cancer (107, 114-116). Some studies suggest the association of red meat with the risk of colon cancer may be due to its fat content (117, 118). However, other hypotheses suggest that the current Western diet which is relatively high in meat augments rates of colon cancer by increasing the faecal concentration of endogenous nitrosamines (119), carcinogenic tryptophan metabolites (120), or carcinogens resulting from the cooking of meat (121). Cooking of the meat is thought to be critical to large intestinal health as it was shown in a study that pan-broiling of beef increased mutagenic activity compared to microwave irradiation (122).

Although recent epidemiological studies suggest that there are strong correlations between red meat consumption and CRC risk (89-91), the proposed mechanisms by which red meat causes this damage remain unproven. The following section explores four major mechanisms by which red meat may increase CRC risk.

17

#### **1.3.1.1** Heterocyclic amines

Heterocyclic amines (HCA) are carcinogenic compounds created by the pyrolysis of creatine, amino acids and monosaccharides which can occur during the cooking process of any protein with the appropriate amino acid composition. *In vitro* experimentation has shown that HCA caused DNA mutations (123). Studies in animal models demonstrated that HCA induced tumours at various sites including the colon (124). In addition, a rodent study has shown that APC mutations were induced when PhIP (2-Amino-1-methyl-6-phenylimidazo[4,5-b]pyridine) (the most abundant HCA generated in cooking) was added to the diet (125). Hence, HCA are suggested to be involved in the initiation process of carcinogenesis, as APC gene mutation occurs in 85% of all sporadic and hereditary colorectal tumours in humans (126).

#### 1.3.1.2 N-nitrosocompounds

N-nitrosocompounds (NOC) are among the most toxic chemical carcinogens (127, 128). A small amount in the human body could induce a significant increase in CRC risk. NOC can be produced endogenously in the colon by a two step process. Amino acids are converted into amines and amides by bacteria in the colon, which then undergo N-nitrosation to form NOC (60). The amino acids contained in red meat, but not white meat, are thought to be important in the formation of NOC as red meat concentration in the diet is directly related to the concentration of NOC in the faeces (129, 130). Furthermore, with high red meat consumption, the concentration of NOC are of the same order as the concentration of tobacco-specific NOC in cigarette smoke (129). In addition, a clinical

trial suggested that it is heme iron, not protein residues or inorganic iron, that stimulates endogenous NOC production (131).

#### 1.3.1.3 Polycyclic Aromatic Hydrocarbons

Polycyclic aromatic hydrocarbons (PAH) are formed by incomplete combustion of coal, oil, petrol, tobacco, meats or other organic materials. Exposure to PAH occurs commonly from consumption of pyrolysed foods or inhaled cigarette smoke (132). Although there are over 100 types of polycyclic aromatic hydrocarbons (PAH), extensive study is conducted only for benzo(a)pyrene (BaP) (133). A previous rodent study has shown that when BaP is given to rodents as part of their diet, tumours developed in the forestomach, oesophagus and tongue (134). However, since quantification of individual PAH is difficult and intake of BaP can occur from tobacco, pollution and other burnt foods, it is inappropriate to assume that dietary meat consumption contributes to the majority of PAH intake. Therefore, the contribution of PAH from the diet to CRC risk remains unclear.

#### 1.3.1.4 Heme Iron

Babbs (1990) (135) suggested that high amounts of unabsorbed faecal iron, resulting from excessive dietary iron, may catalyse oxygen radical production. It is thought that unabsorbed iron increases faecal mutagenicity, activating carcinogens or tumour promoters within the large intestinal lumen. However, there is mixed evidence

regarding the association between high dietary iron concentration and colon cancer risk (136). Sesink *et al.* (1999) (137) investigated the different effects of dietary iron sources on risk markers of colon cancer in a rat model. They showed that heme iron increased cell proliferation, lipid peroxidation and faecal water cytotoxicity in comparison to ferric citrate, protoporphyrin IX and bilirubin. Moreover, carcinogen induced cancer studies in rodents showed that heme iron intake increased aberrant crypt foci (ACF) size and also increased tumour counts (138, 139).

#### 1.3.2 White Meat

White meat refers to any light-coloured meat, such as fish, seafood, and particularly chicken. Examples of white meat proteins include proteins derived from birds, fish, reptiles, crustaceans or bivalves. However, meat that turns white when cooked, like pork, are not considered white meat. Epidemiological studies suggest that the effect of white meat consumption on CRC risk is low (118, 140). However, a 6-year prospective study suggested that both red meat and white meat contribute to increases in CRC risk (141). In a clinical study examining white meat consumption and NOC concentrations, subjects consuming 420-600 g per day white meat had significant reduction in concentration of NOC compared to subjects consuming 420-600 g per day red meat, but no significant difference compared to subjects on a 60 g per day red meat diet (130). Although there are conflicting views on white meat consumption and CRC risk, white meat is still regarded as a low risk food component with regard to CRC.

20

#### 1.3.3 Casein

Casein is a dairy protein which makes up about 75-80% of all milk protein. It is heat stable and often used to nutritionally fortify foods and as a dietary supplement, due to its high protein content and quality. Casein and caseinates are used as extenders and tenderizers in imitation sausages, soups and stews. They are also included in coffee whiteners, sauces, ice cream, salad dressing, formulated meat, bakery glazes, and whipped toppings. In addition, casein is the recommended protein source in the standard American Institute of Nutrition (AIN) rodent diets. Although no extensive epidemiological studies have been conducted on casein and CRC, animal studies indicate that high consumption of casein may be linked with CRC (142, 143). Several studies have shown that cooked casein promotes colon cancer in rats and mice (142, 144, 145). Furthermore, one of these studies indicated that cooked casein caused mucosal abrasion and suggested that this may be the reason for the colon cancer promotion (142). In addition, a previous rodent study showed that there was an increase in colonic DNA damage when dietary composition of casein was increased (143).

#### **1.3.4 Whey Protein**

The main dietary source of whey protein comes from cows' milk, which contains 0.8 g of whey protein per 100 ml, and ricotta cheese which contains 10 g per 100 g. Recently whey protein has been added to increasing numbers of dietary products due to its manufacturing properties such as its gelling strength and antioxidant properties. The

tastelessness of whey protein encourages manufacturers to add it into their product since it does not influence the flavour of the final product. Whey is commonly included as an ingredient in bakery goods, bread, ice cream, frozen desserts, non-fat powder milk and cream fillings.

Whey contains a high proportion of water and can be isolated by ultra-filtration, spray drying and evaporation. Non-protein components of whey can be removed by ion-exchange or micro-filtration, which removes the majority of the fat, lactose and denatured proteins. 70-80 % of the total whey protein is primarily composed of two small globular proteins,  $\beta$ -lactoglobulin and  $\alpha$ -lactalbumin. Other minor components of whey protein include lactoferrin, immunoglobulins, glycomacropeptide, bovine serum album and phosopholipo-proteins. There are limited numbers of published dietary interventions studies investigating whey protein. However, increased use of whey protein has increased attention on its effect on factors such as cancer. To date two studies have examined the effect of whey protein on urogenital, pancreas, liver, and breast cancers (146, 147). Bounous *et al.* (2000) (147) indicated that dietary supplementation of whey has antitumour effects in urogenital cancer. However, a phase I-II clinic study on pancreas, liver and breast cancer did not result in any clear conclusion (146). Both studies suggest possible anti-carcinogenic effects of whey protein.

Findings from animal studies have led to a number of speculations on how whey protein may inhibit the carcinogenesis process. The most likely mechanism is the stimulation of cellular glutathione concentrations. The tri-peptide glutathione (GHS) plays an important role in enzyme activity, and metabolic and cell cycle related functions in all cells (148). GHS and glutathione transferase are major components involved in the metabolism of xenobiotics (149). A primary process to increase the GHS is through

22

increased delivery of cysteine to a cell. Whey protein is the richest protein source for this amino acid and effective supply of cysteine may increase GHS concentrations in cells (150).

#### 1.3.5 Soy Protein

As one might expect, soy protein comes from soybeans, a legume from the pea family. Soy plants store protein in their seeds for development of the embryo. These and other legume seed storage proteins belong to the globulin family of such proteins called leguminins and vicilins. Soybeans also contain other proteins such as prolamin, trypsin inhibitors, hemagglutinins and cystein protease. Soybeans are processed into three types of protein products; soy flour, soy concentrate and soy isolate. Soy protein concentrate is soybean without water soluble carbohydrates and contains approximately 70% soy protein. Soy protein concentrate retains most of the fibre of the original soybean. Soy isolate is the most refined form of soy protein and consists of approximately 90% protein and 10% other components. It is derived from defatted soy meal, which has the majority of fats and carbohydrates removed. Soy flour is made by grinding soybeans into a fine powder. Soy protein in various forms is used in a variety of foods such as imitation meats, soups, salad dressings, cheeses, frozen desserts, pasta products and cereals.

The majority of epidemiological studies examining soy consumption and cancer have focused on breast and prostate cancer, both of which are hormone sensitive. However, previous studies have suggested that CRC may also be hormone sensitive (151, 152). A clinical trial showed that CRC risk is reduced in women by the use of post menopausal hormone therapy (152). Soybeans and most soy-based products contain

23

phytochemicals; in particular, isoflavones, which are structurally similar to endogenous estrogen and bind to estrogen receptors (153). These isoflavones and other phytochemicals have been shown to inhibit cancer cell growth and induce apoptosis (154). However, several rodent studies demonstrated that consumption of soy protein induced damage to the colonic epithelium and stimulated proliferation of colonocytes when compared with a casein diet (155, 156).

Furthermore, results from a meta-analysis of population studies indicated that the relationship of soy consumption and cancer risk may depend on whether or not the soy food was fermented or not (157). For example, the risk of stomach cancer was significantly lowered when high amounts of unfermented soy foods (soy milk, tofu and soybeans) were consumed. However, the risk increased with diets containing high amounts of fermented soy foods (miso, fermented soybeans, and soy paste). Similar results were obtained in a more recent meta-analysis by Spector *et al.* (2003) who suggested that an inverse relationship between unfermented soy consumption and rectal cancer onset but not fermented soy products (158).

There are contradictory findings on the risk of CRC and consumption of dietary soy (154-156, 159). Moreover, lowering of plasma cholesterol by soy in comparison with casein indicates that there are different nutritional properties of soy protein compared to other protein types. Therefore, further investigation is needed to elucidate the effects on soy on the large bowel health.

#### 1.4 Dietary Fibre

Fibre is a heterogeneous group of (largely) plant-derived material. As indicated

previously, these are principally carbohydrates and share the characteristic of being indigestible by human small intestinal enzymes. Also as noted, there is no internationally accepted definition of fibre, dietary fibre is generally considered as complex carbohydrates that reach the colon (160, 161). These complex carbohydrates include a range of non-starch polysaccharides (cellulose, hemicellulose, pectin, gums, and mucilages) and starches which resist digestion (74). Dietary fibre is subclassified into two types, soluble and insoluble. The soluble fibres, such as those found in fruits and oats, tend to be highly fermented in the proximal colon including the caecum (162, 163). The insoluble fibres, such as wheat bran, are more slowly fermented and have a greater impact on fermentation events in the distal colon and are detected more readily in faeces (162, 163).

It would be expected that dietary fibre consumption is likely to lead to a reduction of the risk of CRC. However, the extent and mechanism of protection by fibre has not been fully characterised. Several theories have been proposed as shown in Table 1.1. Originally, the protective mechanism of dietary fibre was thought to be due to dilution and binding of toxins and carcinogens in the intestine through its physical presence (164). Increased faecal bulking from fermentation reduces intestinal transit time. This leads to the reduction in mucosal exposure to potential carcinogens or tumour promoters (165, The basic fermentative reaction in the human colon consists of hydrolysis of 166). polysaccharides, oligosaccharides, and disaccharides to their constituent sugars which are then fermented resulting in an increased biomass (167). Fermentation yields metabolisable energy for microbial growth and maintenance (168). Instability of the colonic fermentation from a diet high in fat and protein but low in fibre is considered to cause an irregularity in the mass of digesta in the colon, which induces fluctuations in colonic fermentation and may increase the possibility of the production of cytotxic agents (and loss

25

on protective ones) leading to greater risk of colorectal carcinogenesis (106).

More recently, there has been increased focus on the prebiotic action of fibre on large bowel health. Until now, the prebiotic functions of fibre and their correlation to CRC have not been adequately explored. Fibre exerts marked changes to the luminal environment and colonic microflora (74) and it has been shown that dietary fibre increases the total mass of bacteria but leads to decreases in secondary bile acid production (169).

Large bowel carbohydrate fermentation lowers digesta pH through direct acidification and the fixing of nitrogen in bacterial mass (74). It is proposed that this reduces the potential tumour promoter activity of secondary bile acids. The enzymatic activity of 7 $\alpha$ -dehydroxylase, which coverts primary bile acids to secondary bile acids changes with reduced pH (170). In addition, a human population study has shown that subjects with the lowest faecal pH have the lowest rates of colon cancer (171). Fibre significantly increases concentration of SCFA and this partially contributes to the lowering of pH. The effect of SCFA, butyrate in particular, is discussed in more detail later in this thesis.

Another possible protective mechanism of fibre is the prevention of CRC via modulation of lifestyle factors (172). It was suggested that the lifestyle factors that cause insulin resistance and hyperinsulinaemia, stimulating the growth of CRC cells. There is some *in vitro* evidence that insulin and insulin-like growth factors (IGF) may influence CRC risk and the risk for developing CRC increases by 40% in diabetes (173). This therefore suggests that further investigation is required.

Population studies have suggested that intake of fibre is linked to reduction of CRC risk (174-176). This led to the initial assumption that non-starch polysaccharides (NSP), major components of dietary fibre, have a significant protective contribution against CRC.

26

However, a recent study showed that NSP may not be ae important a contributor to protection as RS (100).

 Table 1.1: Possible Mechanisms for the protective action of dietary fibre on colorectal oncogenesis (based on Young et al. 2005)

NOTE: This table is included on page 27 of the print copy of the thesis held in the University of Adelaide Library.

## 1.5 Resistant Starch

Starch is a substantial component of most human diets. Historically, starch has been thought to be fully digested to glucose in the small intestine as very little starch is normally found in faeces (74). However, this is now known to be incorrect. RS is an undigested starch which reaches the large bowel and is fermented by colonic microflora, thereby contributing to the optimal function of the large bowel through the production of SCFA (168, 177). RS functions similarly to fibre and many nutritionists are of the optimon that it

should be classified as a component of dietary fibre. RS is found in intact (unprocessed) wholegrain cereals and some seeds and nuts, for example oats, rye, wheat, barley, semolina, corn, linseed and sesame seeds. Processed starchy foods such as some breakfast cereals, white bread, rice and pasta also can contain RS. Recently, some processed foods including breads and cereals have been supplemented with RS in the form of a specific high amylose starch (Hi-Maize<sup>TM</sup>), which is derived from corn. Furthermore, foods such as legumes, for example lentilis, baked beans and unripe fruit (especially bananas), contain RS. In addition, cooked cold rice (such as sushi rice) cold pasta salad and cold boiled potato salad are high in RS.

RS is classified into four groups (74, 178) (**Table 1.2**). RS<sub>1</sub> includes starch that is trapped within whole plant cells and food matrices where there is a physical barrier to amylolysis (conversion of starch to sugar by enzyme and acids). In raw starch granules, starch is tightly packed in a radial pattern and is relatively dehydrated. RS<sub>2</sub> comprises of those granules from certain plants that are gelatinised poorly and hydrolysed slowly by  $\alpha$ amylases. RS<sub>3</sub> represents retrograded starch, where the starch granule is partially or completely hydrated through cooking in water and is allowed to stand. This leads to reassociation of the polymer chains leading to RS formation. Examples of RS<sub>3</sub> starches include cooked and cooled rice or potato. RS<sub>4</sub> comprises the chemically modified starches obtained by chemical treatments, like di-starch phosphate ester, to improve the functional characteristics of starch.

Table 1.2. Classification of types of resistant starch (RS), food sources, and factors affecting their resistance to digestion in the colon

Type of			<b>Resistance Minimized</b>
RS	Description	Food sources	by
$RS_1$	Physically protected	Whole or partly milled grains, seeds and legumes	Milling, chewing

RS <sub>2</sub>	Un-gelatinised resistant granules with type B crystallinity, slowly hydrolysed by α-amylase	Raw potatoes, green bananas, some legumes, high amylose corn Cooked and cooled	Food processing and cooking
RS <sub>3</sub>	Retrograded starch	potatoes, bread, cornflakes, food products with repeated moist heat treatment	Processing conditions
$RS_4$	Chemically modified starches due to cross-linking with chemical reagents	Foods in which modified starch have been used (for example, breads, cakes)	Less susceptible to digestibility <i>in vitro</i>

Starch is made up of glucose molecules linked together to form amylose and amylopectins. Amylose has a linear structure and can form tightly packed granules, which are insoluble and difficult to digest. However, amylopectin has a branched structure and thus can not form tightly packed granules and is, therefore, easier to digest. Most plants contains about 20-25% amylose. However, some plants, such as peas, have 60% amylose and certain species of maize, such as Hi-Maize<sup>TM</sup>, can have up to 80% amylose.

Previously it had been assumed that non-starch polysaccharides (NSP), a major fibre component, were the principle fermentative substrates. A population study showed that individuals generally consume less than 20 g of NSP per day (179). These values are well below the 60-80 g substrate per day required to sustain the 10<sup>13</sup>-10<sup>14</sup> organisms found in the normal human large bowel. It was speculated that this gap was most likely filled by RS (179). A clinical study conducted in South Africa demonstrated that RS consumption may provide more protection against CRC risk than NSP (180). The incidence of CRC and other large bowel diseases in the South African black population is much lower than in the white population (181). The dietary fibre consumption of the urban black population tested was lower than in the white population, but starch intake of the urban black population was significantly higher (182). It was also demonstrated that a typical diet eaten by the black population showed increased SCFA and reduced stool pH compared to a

typical diet consumed by white populations (180). Therefore, these studies suggest that the black population diet is better than the white with regard to CRC risk. This was supported by an epidemiological study which showed that RS is more protective against CRC risk than NSP (100).

The experimental evidence supports protective effects of RS against large bowel diseases. A randomized crossover study has shown that daily consumption of RS (approximately 40 per day) significantly increased the daily excretion of faecal nitrogen and decreased faecal concentrations of both ammonia and phenols (166). Ammonia is thought to promote tumourigenesis by stimulating cell proliferation and favouring the growth of malignant cells, while phenols (*p*-cresol and phenol) by-products from the metabolism of aromatic amino acids, are known for the promotion of skin, bladder and bowel cancers (103, 114-116). Epidemiological studies support the protective role for starch (as RS) in CRC (84, 168). However, the animal experiment data with RS in chemically-induced carcinogenesis are somewhat inconclusive (183).

### 1.5.1 Gut Microbiology

The intestinal ecosystem is characterised by dynamic and reciprocal interactions between host and its microflora. However, despite the fact that the importance of gut microflora for human health is being increasingly recognised, the underlying mechanisms of these interactions are very complex. Nonetheless, evidence obtained through animals raised in germ-free conditions have provided important information about the physiological effect of the microbial community in the gut (184). Main functions of gut flora include fermentation of non-digestible dietary residues and endogenous mucus, salvaging of energy as SCFA, control of epithelial cell proliferation and differentiation, development and homeostasis of the immune system and protection against pathogens (185).

The interaction between dietary factors occurs in the lumen of the large bowel (186). It is possible that dietary carcinogenic effects could be mediated by changes in the metabolic activity and composition of gut microflora. Intestinal bacteria potentially play a part in the initiation of colon cancer through the production of carcinogens, co-carcinogens, or procarcinogens (187). Bacteria, such as bacteroides and the *clostridium* genera increase the incidence and growth rate of colonic tumours induced in rodents (188, 189). In contrast, *lactobacillus* and *bifidobacteria* prevent tumourigenesis (190, 191). Although the evidence is inconclusive, colonic microflora seem to be a critical environmental factor in the modulation of CRC risk.

RS is known to change the composition of the intestinal microflora by stimulating the growth of various bacteria including *bifidobacteria*, *lactobacilli*, *eubacteria* and *streptococci* (192). Lower large intestinal pH results from higher SCFA formation and this affects bacterial populations. The enhanced counts of *lactobacilli* inhibit the growth of pathogenic bacteria such as certain *E-coli* strains, *clostridium* difficile, or sulphur/sulphate reducing anaerobic bacteria (193).

#### **1.5.2** Fermentation

Fermentation is an anaerobic redox process, in which the oxidation of the substrate is coupled to the reduction of another substrate or an intermediate derived from the oxidation, with the difference in redox potential of the substrate and the end product providing energy for ATP synthesis (194). The rate, site and extent of dietary fibre/RS fermentation in the gut is dependent on a number of factors including solubility, chemical structure, availability of other more readily fermentable substrates and the composition of the colonic microflora. Insoluble fibre is resistant to colonic microflora fermentation and contributes to faecal bulk.

Fermentation has a number of important consequences in large bowel physiology and possible protection against CRC. Increases in bacterial cell mass, caused by fermentation, results in an increase in faecal weight. With the increase in faecal weight, transit time is reduced and the large bowel contents are diluted. This reduces the time putative carcinogens are in contact with large bowel mucosa. This effect was originally suggested by Burkitt in 1969 to explain the protective effect of dietary fibre against CRC and has stood the test of time (195).

Increased fermentation is also known to generate an acidic environment with a reduction in pH via SCFA production. Several population studies have shown that subjects with the lowest faecal pH have the lowest grades of colon cancer (171). There are several proposals to suggest how a reduction in colonic pH provides an anti-carcinogenic environment. The acidic environment is known to reduce the potential tumour promoter activity of secondary bile acids. Furthermore, low pH inhibits the activity of the bacterial enzyme,  $7\alpha$ -dehydroxylase, which produces secondary bile acids from primary bile acids (170). Therefore, dietary fibre through acidification of colonic contents is considered protective against colon cancer.

### 1.5.3 SCFA

Over the past few decades considerable investigation has been undertaken in order to understand the physiological role of SCFA. Previously, they were considered as a major factor in the aetiology of carbohydrate induced diarrhoea. However, the rapid absorption of SCFA and the metabolism by the intestine led to a more general speculation that SCFA are beneficial (196).

Dietary and endogenous residues that reach the colon may be metabolised by anaerobic bacteria to produce SCFA and other substrates such as lactate, succinate, ethanol, hydrogen, methane and carbon dioxide. The predominant substrates that contribute to the SCFA production are polysaccharides, oligosaccharides, protein, sugars and mucus (51). The principal SCFA that result from carbohydrate fermentation are acetate, propionate and butyrate. Other SCFA produced in smaller quantities from colonic fermentation are isobutyrate, valerate and isovalerate.

SCFA contribute to large bowel function and prevent pathology through their luminal actions and metabolism by colonocytes (74). Daily production of SCFA in man is approximately 100-200 mM. The majority of this is absorbed by the colon via diffusion or anion exchange (196). Furthermore, absorbed SCFA are transported via the portal vein to the liver, and the fraction not absorbed is distributed to other body organs and tissues. Although SCFA provide valuable information on large bowel health, quantification of the colonic SCFA remains difficult. SCFA in colonic contents have been determined in colostomy patients and post-mortem samples, but these approaches are impractical for large-scale dietary studies. Faecal measurements are useful in determining the changes in excretion, but not necessarily in production because faecal SCFA can be influenced by rate of transit time (74). Therefore, the majority of experimental data have been obtained from animal models, mainly rats and pigs.

33

In general, clinical studies have shown that RS increases faecal SCFA concentrations and excretion (197-199). Furthermore, animal studies have shown that feeding of RS raises large bowel total SCFA (200-203), highlighting the strong association between RS consumption and SCFA production in the large bowel. In addition, it was found that consumption of diets high in RS changed large bowel SCFA profiles, and in particular increases in butyrate and propionate (74).

#### 1.5.3.1 Butyrate

Butyrate has been considered as the most important SCFA. It is thought to play a role in maintaining a normal colonocyte population. Fermentation of RS is known to yield a relatively high proportion of butyrate. Butyrate is considered to be an important by-product of RS-metabolising bacteria within the colon (74).

Butyrate has been suggested to decrease the growth of most human colon cancer cell lines by inhibiting cell proliferation and enhancing differentiation and apoptosis (204, 205).  $\beta$ -oxidation of SCFA occurs in the mitochondria to regulate cell proliferation and apoptosis in the intestinal mucosa (206). This was shown using a knock-out mouse model of short chain acyl dehydrogenase, the enzyme responsible for encoding the mitochondrial  $\beta$ -oxidation of SCFA (207). When the ability to efficiently metabolise SCFA was reduced, the level of apoptosis in the colon was significantly reduced.

The potential anti-cancer properties of butyrate in the colon are linked with a small molecule signalling pathway involving butyrate and its cellular target histone deacetylase (HDAC) (208). Butyrate inhibits the enzymatic activity of HDAC, resulting in changes in the transcription of specific genes, including the induction of the cyclin-dependent kinase

inhibitor p21/Cip1/WAF1 (209). Both inhibitions of HDAC and expression of p21 result in cell cycle arrest and differentiation.

However, not all experiments support a chemopreventitive effect of butyrate (210). For example, a previous rodent study showed that administration of butyrate in the drinking water increased the number of colon tumours detected (211). Furthermore, an azoxymethane-induced colon cancer rodent study demonstrated that administration of slow release butyrate pellets did increase the colonic butyrate concentration but found no protective effect (212). Since there is lack of agreement, particularly between *in vivo* and *in vitro* studies, regarding butyrate and colon cancer, this discrepancy is termed the "butyrate paradox".

The effect of butyrate on colon carcinogenesis may be depended upon the timing of butyrate administration in relation to the stage of the cancer development. For example, butyrate inhibits HDAC. This suggests that in its presence, DNA would be in a more open form, which may be optimal if DNA damage has occurred as more repair mechanisms may be initiated to prevent mutations (213). In addition, concentration is another factor which may affect the physiological effects of the butyrate. Several studies have indicated that low concentrations of butyrate stimulate cell proliferation, but high concentration may inhibit it (214).

## 1.6 Colonic Mucus Barrier

Another defence mechanism of the colon from bacterial and genetic damage is the mucus layer. A mucus layer covers the surface of the gastrointestinal tract, protecting intestinal tissues from pathogens (**Fig. 1.4**). The mucus layer consists of mucins, gel

forming glycoproteins, and a large number of smaller glycoproteins, proteins, glycolipids, and lipids (215). In the large bowel, the layer is especially thick to prevent the large number of bacteria present from infecting tissues (216). Important physiological roles of the mucus include lubrication (217), and a stabiliser for the mucosal association of some enteric bacteria (218). Degradation of the gel layer is likely to result in impairment of these functions. *In vivo* studies have shown that the mucus layer is a dual-layer and protects the mucosa through two mechanisms (216, 219, 220). The outer layer is a nonadherent mucus layer and is believed to reduce the shear stress to the mucosa (221). The constant turnover of this layer may act to trap bacteria, secondary bile acids and endogenous protease. The inner layer acts as a size exclusion barrier to toxic luminal agents. This barrier allows uptake of water, salt and SCFA, while preventing access of larger molecules to the mucosa (220). This dual-layer system is necessary to prevent digestion of the colonic wall by both hosts and bacterial enzymes.

The thickness of the colonic mucus layer is known to be influenced by diet (222). A recent study demonstrated that there was a significant decrease in mucus thickness and secretory response when rats were fed fibre-deficient diets (222). In addition, another rodent study showed that RS contributed to the maintenance of the colonic mucus layer thickness (143). This study further showed that colonic genetic damage was strongly linked with decreased mucus layer thickness.

NOTE: This figure is included on page 37 of the print copy of the thesis held in the University of Adelaide Library.

#### Figure 1.4: The mucosal barrier

The mucosal barrier separates the internal milieu from the luminal environment. Function of the barrier depends on factors such as the endothelium and epithelium lining, mucosal blood flow, reactivity of dynamic defence system, epithelial secretions and immunocompetent cells. (Based on Callicott & Womack 2006)

### 1.6.1 Mucins and MUC genes

Mucins, which form the major component of mucus, are large carbohydrate-rich glycoproteins and have a high content of cluster oligosaccharides O-glycosidically linked to tandem repeat peptides (mainly threonine, serine and proline) (223, 224). There are two structurally and functionally different classes of mucins: secreted gelforming mucins

**Research Background** 

(encoded by the following genes, MUC 2, MUC5AC, MUC5B, and MUC 6) and transmembrane mucins (MUC 1, MUC3A, MUC3B, MUC4, MUC12, MUC17). Besides their protective function in the normal colon, alternations in mucins are a common feature of colonic neoplasia. Moreover, expression of several MUC genes is linked with colon cancer. MUC 1 expression is increased in colon cancer (223), while several studies have found that expression of MUC2 is generally decreased in colorectal adenocarcinoma (225, 226). Also, MUC2 knock-out mice develop adenomas in the small intestine that progress to invasive adenocarcinoma, as well as rectal tumours (227), suggesting that MUC2 gene is involved in the suppression of CRC. Furthermore, MUC5AC which is normally expressed in the surface epithelium of the normal stomach, and not in the normal colon, is often expressed in adenomas and colon cancers (228, 229).

## 1.7 Biomarkers of Large Bowel Health

There are two major categories of biomarkers for CRC research. They involve changes in colonic epithelium or changes in faecal composition. Biomarkers of colonic epithelium provide greater information than faecal biomarkers. However, to obtain a sample of epithelium in clinical studies requires invasive methods such as pinch biopsy, or can only be collected at autopsy. Wide-scale screening using faecal occult blood testing results in up to 33% reduction in colorectal adenocarcinoma mortalities, but requires many unnecessary colonoscopies (230-232). Therefore, stool DNA is emerging as a potentially important new approach for the early detection of colorectal neoplasia (233). The most common genetically based faecal tests applies the concepts of chromosomal instabilities with mutations progressively accumulating in the APC, p53 tumour suppressor genes, and

K-ras oncogene (234). Moreover, a recent study demonstrated that exfoliated cells from the surface of the stool can be used to measure DNA adducts (57). However, measurements of human colonocytes via biopsy or autopsy still provide the best indication of large bowel health as exfoliated cells in general have very low cell viability. Other colon cancer biomarkers used in previous studies include proliferation, apoptosis, faecal mutagenicity and total and secondary faecal bile acid excretion.

#### **1.7.1 Proliferation and Apoptosis**

A measurement of proliferation provides an indication of how a dietary component or chemical may increase colon cancer risk. Proliferation, a process in which cells divide, increases the chance of genetic errors related to the carcinogenic process. Therefore, proliferation is often linked with risk of cancer (235, 236). In contrast, apoptosis provides information on the way in which cells with genetic mutation can be removed. However, the rate of apoptosis is very low in the colon and meaningful measurements can only be obtained if a chemical carcinogen is administered to increase the rate of apoptosis (237).

#### 1.7.2 Comet Assay

Colonic genetic damage is thought to be prerequisite for sporadic cancer initiation. The alkaline comet assay (single-cell electrophoresis) is used frequently in genotoxicity testing and population biomonitoring (238, 239). This is due to its sensitivity for detecting single-strand DNA breaks, ease of use and rapid performance. Furthermore, this assay requires a relatively small number of cells compared to other genotoxicity assays. In previous studies, the comet assay has been successfully used to determine DNA damage in rat colonocytes (143, 240). The comet assay is a useful method to measure and quantify DNA damage (strand breaks) in individual cells (241-243) due to the higher mobility of smaller DNA fragments during electrophoresis. The intensity of the DNA tail increases in proportion to the extent of DNA damage (**Fig. 1.5**) (244). A revised version of the method has also been developed to measure double-strand DNA breaks independent of the presence of single-strand breaks (245, 246). Furthermore, specific base damage can be identified by incubating lysed cells with base damage-specific endonucleases before electrophoresis (247). It is these reasons that led to this assay being used in the experiments presented in this thesis.



**Figure 1.5** Image of a typical cell used for comet assay. 200x magnification, section stained with propidium iodide.

## 1.8 HYPOTHESES AND AIMS

Recently, there is an increased public interest on high protein diets as means of weight control. However, effects of these diets on large bowel health need better understanding. Epidemiological and experimental evidence suggest that high protein diets have adverse effects on the large bowel. In contrast, dietary fibre, RS in particular, appears to be protective against large bowel diseases. The present thesis investigated the effect of several different types of protein sources (red meat, white meat, soy, whey and casein) on colonic DNA damage and interaction with RS.

### **1.8.1 General Hypotheses**

- Diets high in dietary protein increase colonic DNA damage and reduce the colonic mucus barrier
- Dietary red meat increases colonic DNA damage compared to other protein types
- RS protects large bowel health by increasing colonic fermentation and SCFA production (butyrate in particular)
- RS dose-dependently protects colonic DNA damage
- Red and white meat dose-dependently increases colonic DNA damage

### 1.8.2 Specific Aims and Objective of This Thesis

The aim of this thesis was to gain insights into how different types of proteins influence large bowel health, with particular respect to CRC risk. To investigate this aim,

animal studies were designed to determine:

- 1. The effect of red meat on CRC risk and thickness of the mucus layer
- 2. Whether RS protective against red meat-induced colonic DNA damage
- 3. Whether special types of proteins (whey and soy) cause similar colonic DNA damage as casein
- 4. Whether RS protects protein induced colonic DNA damage in a dose-dependent manner
- 5. Whether white meat increases colonic DNA damage similar to casein and red meat
- 6. Whether red meat increases colonic DNA damage in a dose-dependent manner
- Whether butyrate is the major component responsible for the protective effects of RS

# **CHAPTER 2**

## RESISTANT STARCH PREVENTS COLONIC DNA DAMAGE INDUCED BY HIGH DIETARY COOKED RED MEAT OR CASEIN IN RATS

S. Toden<sup>\*</sup>, A.R. Bird, D.L. Topping and M.A. Conlon

CSIRO Human Nutrition, Kintore Avenue, Adelaide, Australia;

CSIRO Food Futures National Research Flagship, Australia.

\*Discipline of Physiology, School of Molecular Science,

University of Adelaide, Adelaide, Australia;

**Cancer Biology and Therapy, 2006, 5(3),** 267-72

#### **Publication 1:**

The aim of this publication was to determine if dietary cooked red meat can cause colonic DNA damage and whether RS is effective in reducing such damage. The rats used in this study were fed diets containing 15% or 25% casein and 25% red meat, with or without 48% high amylose maize starch.

The results showed that red meat diet induced significant higher colonic DNA damage compared to casein diet in the absence of RS. These damages were reversed by inclusion of dietary RS in the diet. Several colonic parameters were measured to identify the changes occurred with these diets. The rats fed RS showed increased digesta weights and SCFA pools (including butyrate) in caecum and faeces, along with lowered pH of the faeces and caecal digesta, and increased weight of the caecum and length of the colon. Furthermore, high dietary casein and red meat reduced the thickness of the colonic mucus layer compared to the low casein diet in the absence of RS. The addition of RS attenuated the loss of colonic mucus thickness induced by both high protein diets.

These results suggest that the inclusion of RS can effectively attenuate the increase DNA damage and reversed thinning of the colonic mucus barrier induced by high protein diets (both red meat and casein). Therefore, food contains RS may provide protection against bowel diseases such as colorectal cancer.

Toden, S., Bird, A.R., Topping, D.L. and Conlon, M.A. (2006): Resistant Starch Prevents Colonic DNA Damage Induced by High Dietary Cooked Red Meat or Casein in Rats.

Cancer Biology and Therapy, v. 5 (3), pp. 267-272, March 2006.

NOTE: This publication is included on pages 45 -50 in the print copy of the thesis held in the University of Adelaide Library.

# **CHAPTER 3**

## DIFFERENTIAL EFFECTS OF DIETARY WHEY, CASEIN AND SOY ON COLONIC DNA DAMAGE AND LARGE BOWEL SCFA IN RATS FED DIETS LOW AND HIGH IN RESISTANT STARCH

S. Toden<sup>\*</sup>, A.R. Bird, D.L. Topping and M.A. Conlon

CSIRO Human Nutrition, Kintore Avenue, Adelaide, Australia;

CSIRO Food Futures National Research Flagship, Australia.

<sup>\*</sup>Discipline of Physiology, School of Molecular Science,

University of Adelaide, Adelaide, Australia;

**British Journal of Nutrition, 2007** 

#### **Publication 2:**

The results shown in the Publication 1 provided evidence that consumption of a high protein diet, as both casein and red meat, increases colonic DNA damage. It also highlighted that this damage was attenuated by the addition of RS in the diet. This increase in colonic DNA damage induced by high protein diets supports the findings of a meta-analysis study that a high protein diet increases risk of colorectal cancer (100). However, the role of dietary proteins in cancer aetiology in general is poorly understood and it remains to be established whether other protein types have similar effects. Therefore, the aims of this second publication was to determine whether high dietary dairy (casein or whey) or plant (soy) protein have adverse effects on the large bowel and whether dietary RS was protective against these different protein sources.

In this experiment the effects of three different types of dietary proteins, casein, whey and soy were examined with relation to colonic DNA damage, colonic mucus layer thickness and SCFA levels in the caecum and faeces of rats. High dietary casein and soy (but not whey) increased colonic DNA damage. DNA damage was highest with soy when fed 15 or 25% protein in the absence of RS. Caecal total SCFA and butyrate pools were higher in rats fed RS compared with digestible starch.

These results suggest that diets high in protein affected colonic DNA damage and the mucus layer thickness. However, proteins differ in their effects on these indices. Furthermore, these protein induced changes were reversed by inclusion of dietary RS. Therefore, RS has protective effect against the colonic DNA damage produced by variety of protein sources. Toden, S., Bird, A.R., Topping, D.L. and Conlon, M.A. (2007) Differential effects of dietary whey casein and soya on colonic DNA damage and large bowel SCFA in rats fed diets low and high in resistant starch.

British Journal of Nutrition, v.97 (3) pp. 535-543, March 2007

NOTE: This publication is included on pages 53 - 61 in the print copy of the thesis held in the University of Adelaide Library.

It is also available online to authorised users at:

http://dx.doi.org/10.1017/S0007114507336817

# **CHAPTER 4**

## DOSE-DEPENDENT REDUCTION OF DIETARY PROTEIN-INDUCED COLONOCYTE DNA DAMAGE BY RESISTANT STARCH IN RATS CORRELATES MORE HIGHLY WITH CAECAL BUTYRATE THAN WITH OTHER SHORT CHAIN FATTY ACIDS

S. Toden<sup>\*</sup>, A.R. Bird, D.L. Topping and M.A. Conlon

CSIRO Human Nutrition, Kintore Avenue, Adelaide, Australia;

CSIRO Food Futures National Research Flagship, Australia.

<sup>\*</sup>Discipline of Physiology, School of Molecular Science,

University of Adelaide, Adelaide, Australia;

Cancer Biology and Therapy, 2007

#### **Publication 3:**

The first two publications have demonstrated the effects of various protein types on colonic DNA damage and the reversal effects of RS. In these studies, 48% HAMS diets were used as the source of RS, but 48% HAMS is too high for human to consume as diets. This next publication sought to determine whether there was a dose relationship between RS intake and amelioration of the effects of high dietary protein in the form of casein. Rats were fed a diet containing 25% casein with 0%, 10%, 20%, 30% or 40% HAMS for 4 weeks, as shown to produce significant increase in colonic DNA damage in publications 1 and 2. The single-strand comet assay was used to determine the colonic DNA damage and this was combined with the use of endonuclease III to ascertain whether formation of 8-hydroguanosine was involved.

Colonic DNA damage was greatest and the mucus barrier was thinnest in rats fed 0% HAMS. DNA damage was reduced and thickness of the mucus barrier increased in a logarithmic dose-dependent manner by HAMS. However, there was no significant difference in 8-hydroguanosine between dietary groups indicating that 8-hydroguanosine formation was independent of dietary HAMS level. As expected caecal and faecal SCFA pools were elevated with increased levels of dietary HAMS.

This publication extends current literature to confirm the protective effects of RS through SCFA production, most likely through butyrate production. Furthermore, inclusion of 10% HAMS was found to be sufficient to oppose colonic DNA damage, and to increase caecal and faecal SCFA.

Toden, S., Bird, A.R., Topping, D.L. and Conlon, M.A. (2007): Dose-Dependent Reduction of Dietary Protein-Induced Colonocyte DNA Damage by Resistant Starch in Rats Correlates More Highly with Caecal Butyrate than with Other Short Chain Fatty Acids.

Cancer Biology and Therapy, v. 6 (2), pp. 253-258, February 2007.

NOTE: This publication is included on pages 64 - 69 in the print copy of the thesis held in the University of Adelaide Library.

# **CHAPTER 5**

## HIGH RED MEAT DIETS INDUCE GREATER NUMBERS OF COLONIC DNA DOUBLE-STRAND BREAKS THAN WHITE MEAT IN RATS: ATTENUATED BY HIGH AMYLOSE MAIZE STARCH

## Toden S, Bird AR, Topping DL and Conlon MA

CSIRO Human Nutrition, Kintore Avenue, Adelaide, Australia;

CSIRO Food Futures National Research Flagship, Australia.

CSIRO Preventitive Health National Research Flagship, Australia

<sup>\*</sup>Discipline of Physiology, School of Molecular Science,

University of Adelaide, Adelaide, Australia;

Carcinogenesis, 2007

#### **Publication 4:**

The previous published chapters have shown that increases in dietary protein, as red meat, casein or soy, increased colonic DNA damage in rats. These damages were prevented when RS was added to the diet. The aims of this final publication were to determine the genotoxic burdens of dietary red and white meats on large intestine by measuring single-strand DNA breaks (SSB) and double-strand DNA breaks (DSB) and whether addition of RS in the diet provides protection.

Until now there have been limited studies on the effects of the high dietary red meat on colonic DNA damage. Recent epidemiological studies indicated that high red meat consumption increases the risk of colorectal cancer, but not white meat (87, 89, 91, 117). In contrast, dietary fibre is thought to have protective effects (81). The previous publications presented have shown that high levels of dietary protein significantly increased colonic DNA damage as SSB. In this publication we measured SSB and DSB, a more potent genetic damage. DSB differs from other types of DNA damage as both DNA strands of the double helix are damaged, which can be a potent inducer of chromosomal aberrations. Rats were fed diets containing 15, 25 or 35% of cooked beef or chicken at levels to provide equivalent amounts of protein as beef (13, 22 or 30%), and with or without 20% HAMS for 4 weeks.

Increased consumption of protein as either red or white meat increased colonic DNA damage. However, there was a significantly greater colonic SSB and DSB damage for the rats that consumed red meat. This increased colonic DNA damage was attenuated by the addition of RS in the diet. Thinning of the mucus barrier was again observed with high protein diets and this was reversed with addition of dietary RS. Caecal and faecal measurements indicated that RS consumption increased SCFA pools in caecam and faeces.

This publication demonstrated that dietary red meat had significantly greater

genetic damage than white meat, which corresponds with finding of epidemiological studies. Furthermore, the results of this publication again confirm that RS can effectively reduce the colonic DNA damage caused by high protein diets.

Toden, S., Bird, A.R., Topping, D.L. and Conlon, M.A. (2007) High Red Meat Diets Induce Greater Numbers of Colonic DNA Double-Strand Breaks than White Meat in Rats: Attenuation by High Amylose Maize Starch. *Carcinogenesis, (published online on October 4, 2007)* 

NOTE: This publication is included on pages 73 - 103 in the print copy of the thesis held in the University of Adelaide Library.

It is also available online to authorised users at:

http://dx.doi.org/10.1093/carcin/bgm216

# **CHAPTER 6: CONCLUSIONS**

The aim of these studies was to compare the deleterious effects of high levels of specific dietary proteins on the integrity of the colonic mucosa of a model animal species, the rat. The studies also aimed at determining the protective effect of resistant starch (RS) fed as a high amylose maize starch against protein-induced damage. As the chapters in this thesis are presented in a manuscript format, the conclusions and implications of the 4 major publications presented in this thesis are discussed below.

## 6.1 Conclusions and Implications

#### 6.1.1 Differential Effects of Various Protein Types

The proposition that greater protein intake was linked to increased CRC risk has been strengthened by several recent, large epidemiological studies showing that consumption of red meat correlated with higher rates of CRC (89-91). The original hypothesis of this thesis was that the increased risk of CRC is associated with red and processed meat may be a result of increased consumption of protein *per se*. The study described in Chapter 2 of this thesis demonstrated for the first time that consumption of high levels of dietary protein, as red meat, increased colonic DNA damage compared to equivalent levels of casein. In humans these effects could translate to increased risk of CRC. Furthermore, there was a thinning of the mucus layer correlated with increased colonic SSB DNA damage. This is a particularly important finding as it could link to IBD where breakdown of the mucus barrier occurs leading to greater potential for bacterial translocation and an inflammatory response. IBD is also a risk factor for CRC.

Collectively, these data show that dietary protein could increase genetic damage in the absence of a cytotoxic agent such as AOM. However, the data show also that the effect was due to protein and not to red meat alone.

In order to determine whether various protein sources have differential effects on the integrity of colonocyte DNA, the experiments reported in Chapter 3 of this thesis extended the study described in Chapter 2. Whey and soy protein were substituted for red meat as protein sources specifically to compare a plant protein isolate with another protein of animal origin. The absence of any loss of colonocyte integrity with whey protein confirmed previous rodent studies indicating that it could provide protection for the large bowel (93, 248). Soy protein is often regarded as a health food product (249), and is used by vegetarians as a dietary protein source. In contrast to whey, greater dietary soy protein increased genetic damage. Therefore, these current results indicate that whey protein is more protective against colonic DNA damage compared to other protein sources, such as casein, soy or red meat. This is supported by a previous study using rodents with chemically-induced cancer which showed whey protein to be the most protective against tumour formation and soy protein to be the least protective compared with casein and red meat (250). Furthermore, a rodent study of AOM-induced tumours demonstrated that bovine lactoferrin, a major whey component, reduced colon carcinogenesis with a significant lowering in adenocarcinoma incidence (251). One of the possible protective mechanisms of whey is its capacity to increase tissue glutathione. Whey protein can raise colonic glutathione concentration in rats and this may facilitate deactivation of xenobiotics via glutathione transferase activities (147, 156).

While the consumption of whey protein may be beneficial for large bowel health it is highly unlikely that people will consume enough to produce these effects independently. However, whey protein is usually consumed in supplements, especially for weight control and body building. Under these circumstances enough might be consumed where its lack of genotoxic effect could become significant.

One of the other striking effects of the experiment described in Chapter 3 of this thesis is the significant increase in the colonic DNA damage induced by high consumption of soy. This occurred despite any reduction in the thickness of the colonic mucus layer as found with other protein types. The data regarding soy consumption and CRC risk are contradictory. A previous rodent study showed that soy-fed rats showed lesser tumour formation compared to those fed without soy (252). It was suggested that minor components (eg isoflavones and saponins) may be responsible for the attenuation of the cancer formation by inhibiting proliferation (253). Furthermore, previous studies suggest that antioxidant properties of soy isoflavones may prevent breast cancer (254, 255). In addition, Mitchell and Collins (1999) (256) demonstrated using the comet assay that soy supplements decrease DNA damage in human lymphocytes. However, other studies indicate that consumption of dietary soy increases damage to epithelial cells in the large intestine (155, 156). Furthermore, soy saponins are known to increase bile acid and steroid excretion and secondary bile acids are cytotoxic within the large bowel (257-259). In addition, soy also increases faecal fat and this may contribute to the increase in damage Whatever the other reported effects of soy, the current data confirm that DNA (156). damage was induced by a non-animal protein source and that the capacity of individual proteins to induce strand breaks needs to be examined on a case-by-case basis.

Chapter 5 of this thesis compared the effects of dietary red and white meat on colonic DNA damage and other indices of bowel health. The rats consuming red meat caused a significantly greater number of colonic DNA SSB and DSB than white meat.

These findings are consistent with epidemiological data (89-91, 117, 260). Red meat contains greater amounts of heme in comparison to white meat and heme is known to stimulate production of genotoxic endogenous N-nitrosocompounds (NOC) in the human gut (129, 131). Hence differences in heme might explain the observed differences in DNA damage between the meat sources. Furthermore, endogenous N-nitrosation can lead to formation of promutagenic and toxic DNA adducts such as O<sup>6</sup>-carboxymethyl guanine (58).

In conclusion, this thesis demonstrated that different protein types have differential effects on the integrity of colonocyte DNA. However, further investigation is necessary to identify the mechanisms by which different protein types affect the integrity of colonocyte DNA.

#### 6.1.2 Protective effects of RS

The role of dietary fibre in CRC is not clear-cut. Several epidemiological studies showed that dietary fibre intake is negatively associated with CRC risk (78, 81, 140). These include a large European multi-centre prospective study which showed a dose-dependent reduction in incidence with greater fibre intake. However, other studies have failed to show any protective effects. An early meta-analysis showed that RS, and not fibre NSP, contributes to protection against CRC (100). RS is highly fermentable in the large bowel and leads to SCFA production rather more than NSP (74). The studies in this thesis demonstrate that RS consumption reduces colonic DNA damage, increases SCFA production in the colon and restores the thickness of the colonic mucus barrier. This occurred in the presence of wheat bran as an NSP source.

One of the basic hypotheses in this thesis was that higher large bowel SCFA levels (produced from RS fermentation) correlated with greater protection against proteininduced DNA damage. As expected from the large body of literature, greater RS consumption led to overall increases in caecal and faecal SCFA. The majority of the studies showed that total and individual SCFA pools correlated with colonic mucus layer thickness and inversely correlated with colonic DNA damage for both SSB and DSB. Furthermore, of all SCFA, butyrate showed the strongest correlation with colonic DNA damage. This finding supports many previous studies which show that butyrate is the key SCFA for maintenance of large bowel health (74, 261-263). The precise mechanism whereby this protection occurs needs further study, especially in the context of the mechanism whereby dietary proteins induce that damage.

It needs to be recognised that dietary RS (and fibre) may protect against CRC by mechanisms other than SCFA production. One of the most widely discussed is one which was proposed several decades ago by Dennis Burkitt (195) and suggested that the greatest benefit of fibre was through faecal bulking which diluted luminal toxic compounds and minimised contact between them and the colonic wall. The studies in this thesis support this hypothesis with strong inverse correlations between caecal bulk and colonic DNA damage. As indicated, greater SCFA production is a further mechanism of protection. The data presented in Chapter 4 of this thesis demonstrate that SCFA, especially butyrate, showed strong inverse correlation to colonocyte DNA damage. SCFA are thought to have multiple effects on colonic epithelial cells at different stages in growth, development, transformation and cell death that may explain a decrease in cancer risk (264). Although SCFA have a trophic effect on the normal colon, they seem to exert opposite effects on cancer cell lines by increasing apoptosis and decreasing proliferation (265). Butyrate is a

preferred metabolic fuel for normal colonic epithelial cells but inhibits proliferation and stimulates differentiation of human colon cancer cell lines (204, 205, 265, 266). Moreover, *in vivo* studies have shown that butyrate influences colonic mucus secretion (267-269). The studies in this thesis demonstrated a strong correlation between caecal butyrate and the colonic mucosal barrier. In addition, RS consumption increased overall apoptosis levels of colonocytes, which supports the findings of a previous study (270).

#### 6.1.2.1 Dose-Response Effects of RS

One of the main criticisms of the studies conducted in chapters 2 and 3 was the high content of RS in the diet (48% HAMS) which is rather higher than that considered to be representative of human intakes. Chapter 4 of this thesis tackled this issue by conducting a study on various levels of RS on a high protein diet to determine the effectiveness of RS at lower dosages. The results showed for the first time that the reduction in comet tail moment by increased dietary RS was a dose-dependent one. Furthermore, it was found that even an inclusion of 10% HAMS significantly increased colonic butyrate production and was able to significantly reduce the high protein induced colonic DNA damage. A previous dietary study suggests this dose is achievable in a human diet (271).

#### 6.1.2.2 Oxidative Damage

Another potential protective mechanism investigated in this thesis was oxidative damage. Oxidative damage through free radical generation was proposed as a potential contributor to experimental carcinogenesis by the heme iron in red meat (247, 272). It has

been shown in an *in vitro* (cellular) study that both acetate and butyrate reduced oxidative damage but isobutyrate and propionate were ineffective (273). However, there appear to have been no previous studies of the effectiveness of RS in reducing oxidative damage. In chapter 4, the endonuclease III FLARE comet assay was used to determine whether dietary RS induces changes in oxidative damage. Subtraction of the baseline values from those with endonuclease III showed that the contribution of 8-hydroguanosine formation was small and that tail moment was independent of dietary RS level. Therefore, it appears that RS does not lower colonic DNA damage through lowering oxidative damage. This leaves other potential protective mechanisms of RS including absorption of carcinogens, modification of intestinal microflora, alteration of faecal bile salt excretion and lowering of pH (204).

#### 6.1.3 Mucus Layer Thickness

Thickness of the mucus barrier has been identified as a key protective element in the health of the large bowel. Intestinal mucins, secreted by goblet cells, forms a highly hydrated mucus gel coating the epithelial surface of the intestinal tract (274). As mucins play a key protective role to underlying epithelium, any quantitative change in mucus secretion may modify this defensive barrier (275, 276). Furthermore, loss of barrier function is a prominent feature of inflammatory bowel diseases (277). For these reasons, the thickness of the colonic mucus layer was measured in every experiment throughout this thesis. With the exception of soy protein, the combined results indicate that increased DNA damage correlated with thinning of the mucus layer thickness. The reason for this differential effect of soy is unclear. However, in every case where there was mucus

thinning of the mucus layer, it was countered by inclusion of RS in the diet. One possible mechanism for this protection may be increased fermentation and therefore, the production of SCFA. Previous investigations suggested that dietary supplementation with fibre may modify the mucin composition favourably or increase the number of goblet cells (278, 279). Furthermore, *in vivo* studies indicate that SCFA increase colonic mucin secretion (267-269). Furthermore, a human biopsy study showed butyrate in particular reduced sulphide induced mucosal hyper-proliferation (280). SCFA may inhibit colonic mucin fermentation by reducing sulphide production while enhancing mucin secretion. The interactions between dietary resistant starch and secretory activity of goblet cells may provide a means for the manipulation of the mucus barrier to improve its protective functions.

#### 6.1.4 Red Meat and DNA DSB

The experiments described in chapter 5 revisited the effect of red meat on CRC risk. This time consumption of red meat was compared to that of white meat and their dose-dependencies (15, 25 and 35% protein contents) in terms of colonic DNA damage. Both DSB and SSB were measured in colonocytes. DSB is a more potent form of DNA damage compared to SSB and is considered to be a better indicator of CRC risk. The results demonstrated that increases in dietary red meat led to increased DSB as well as SSB. Furthermore, this increase in DSB was seen only with dietary red meat consumption and not white meat. DSB can result in chromosomal aberrations as they can affect many genes (52) leading to the malfunctioning of cells and cell death. Furthermore, several studies have shown a strong link between impaired DSB repair systems and predisposition

to cancer (281, 282). The results of this study suggest strongly that red meat consumption changes the integrity of colonocyte DNA more than white meat and that similar changes in humans could be a contributing factor to the differential risks of red and white meat consumption on CRC as indicated by epidemiological studies (87, 89, 91).

In summary, this work has demonstrated that the consumption of high protein diets (as casein, soy and red meat) increased colonic DNA damage. The effects of red meat support the findings of the previous studies (89, 90, 107, 140). The studies demonstrated that there were differential effects of protein induced colonic DNA damage depending on the types of proteins. In addition, there was a strong correlation between the colonic DNA damage and the colonic mucus barrier thickness and the caecal SCFA production.

## 6.2 Future Directions

Future research investigating the effect of dietary protein on CRC risk needs to focus on the specific mechanisms involved and how these findings relate to human large bowel health.

## 6.2.1 Animal and Human Models

The research conducted in this thesis indicates a strong link between increased consumption of several types of protein consumption and increased DNA damage in the colon. Further investigations in other animals (i.e. pigs) which resemble humans more closely are needed to confirm the findings of this thesis. Rodents are valuable for genotoxicity testing since rodents and humans have similar biological functions. However, rodents and humans have many differences including, lifespan, body weight, gut microflora and gene regulations. A meta-analysis analysed the suitability of rodent models of carcinogenesis in predicting efficacy in humans using aspirin,  $\beta$ -carotene, calcium and wheat bran (283). The results indicated that, for studies on carcinogen induction of tumours, rats matched humans for aspirin,  $\beta$ -carotene, calcium and wheat bran. This indicates that rodents can be used to predict effects in humans. However, one of the disadvantages for using a rodent model in gastrointestinal research is the considerably greater relative size of the rat caecum in proportion to that of humans. Pigs are generally considered as a better model for large bowel study than rodents, due to their comparable size and structure to the human colon (284). However, pigs are significantly more expensive than rats and so tend to be used only after preliminary rodent studies.

There are several hurdles to overcome in order to conduct successful clinical studies on high dietary protein and RS consumption on large bowel health. The main concern for a clinical study is that human colonocytes are very difficult to obtain (285-287). Colonoscopy is one of the few techniques which can be used to obtain intact colonocytes. However, the procedures are invasive with the attendant risk of major complications and can be costly (285, 287, 288). Furthermore, isolating colonocytes from mucus and colon tissues from colonoscopy samples can be challenging. An alternative method for obtaining colonocytes is to collect exfoliated cells from faeces. Lewin *et al.* (57) showed that it is possible to collect exfoliated cells from the colon and measure the damage of the cells. However, the main concern with this method is that majority of exfoliated cells are generally not viable and this means that genotoxicity assays, such as

the comet or micronucleus assays, may not be sufficiently robust to measure genetic damage in these cells. Recently, instead of collecting exfoliated cells, DNA has been isolated from faecal samples to measure chromosomal instability and mutations (234). This method requires a large-volume faecal sample and the genetic mutation sites are detected and quantified with real time polymerase chain reaction (233, 234). However, sensitivities for detecting adenoma and adenocarcinoma are still significantly weaker than those obtained with colonoscopy (289). Nevertheless, faecal DNA biomarkers for cancer screening and early detection tools for large populations look promising.

#### 6.2.2 Mechanisms of Protein Induced Colonic DNA Damage

This research project has demonstrated that there are differential effects on protein induced colonic DNA damage depending on the protein type utilised. It was originally hypothesised that increases in colonic DNA damage were the effect of protein *per se*. However, the findings of the chapter 3 of this thesis indicated that the protein types can influence the extent of colonic DNA damage. Furthermore, it was shown that red meat significantly increased DSB as well as SSB when compared to corresponding white meat protein concentration in the absence of RS. All of the studies conducted in this thesis suggest that the mucus layer thickness correlated with colonic DNA damage. The sole exception was soy, for reasons which are unclear. These results suggest that genotoxins may reduce the thickness of the mucus barrier by influencing the turnover rate or the production rate of mucus by epithelial cells. Hence, it is likely that the mechanisms of protein induced colonic DNA damage on the large bowel health may be more complex than previously anticipated. This thesis focused on DNA damage as single and double strand DNA breaks. However, other types of DNA damage may be involved eg base loss, chemical modifications, replication errors and cross link formation. The comet assay was used to measure the DNA damage of the colonocytes, but there are some limitations attached to this method. The main one is that interpretation of the data can be complicated by the fact that there is no simple relationship between the amount of DNA damage caused by a specific chemical and its biological impact (239, 290). Furthermore, if the samples contain predominantly necrotic and apoptotic cells, accurate information cannot be obtained (239). Further studies are needed to clarify the mechanisms of how certain protein sources increase colonic DNA damage.

#### 6.2.3 Protective Mechanisms of RS

Whether the RS protects large bowel health via faecal bulking effects or increased SCFA production is yet to be determined definitively. Since the majority of SCFA are produced during the fermentation process, it is difficult to determine whether faecal bulk or SCFA production provides large bowel protection. Furthermore, it is most likely that faecal bulking and SCFA production protect the colon at a different stage of carcinogenesis. Faecal bulking will dilute toxins and carcinogens in the faeces and reduce intestinal transit time, thus minimising mucosal exposure to potential carcinogens or tumour promoters (74, 165, 166). However, it is unlikely that this dilution and reduced transit time are beneficial at adenoma or carcinoma stages of carcinogenesis. In contrast SCFA, butyrate in particular, have been shown to increase apoptosis and reduce proliferation (262, 291). It has been shown that RS consumption increased apoptosis in

carcinogen damaged rat colonocytes (292). Furthermore, SCFA modifies the composition of the microflora, especially by stimulating beneficial bacteria including bifidobacteria and lactobacilli (292-294).

Recent introduction of modified starch sources may clarify the protective effects of RS. These modified starches are acylated with specific SCFA which are released by bacterial action in the colon (295, 296). Therefore, these starches will be able to determine the effects of SCFA on colonic DNA damage independently of faecal bulking and of fermentation. The complexity of the large bowel system makes identification of the specific mechanisms difficult. Colonic SCFA production is known to change the bacterial populations and there are still large numbers of micro-organisms in the gut which are yet to be identified. It is possible that RS alters cancer risk by modifying bacterial metabolism (eg carcinogen production) by subtle changes which need not involve SCFA.

#### 6.2.4 Red Meat and Colorectal Cancer Risk

The dose-dependent increases in both DNA SSB and DSB with dietary red meat support the idea that this protein source has adverse effects on the integrity of colonocyte DNA. One of the common explanations for the association between increase risk of CRC and red meat consumption is increased formation of heterocyclic amines (HCA) while meat is cooked. The 4<sup>th</sup> chapter of this thesis cooked both red meat and white meat by the same procedure to minimise carcinogenic compounds linked to different cooking procedures and still showed significant differences in colonic DNA damage. Red and processed meat contains greater amounts of heme in comparison to poultry (129). Heme is known to stimulate the production of genotoxic endogenous NOC in the human gastrointestinal system (131). A human study showed that the high levels of red meat consumption (600 g a day) had the same magnitude of concentration of total NOC as cigarette smoke (129). Moreover, endogenous N-nitrosation can lead to the formation of promutagenic and toxic DNA adducts such as  $O^6$ -carboxymethyl guanine (58). Another possible mechanism for the red meat induced colonic DNA damage is the formation of hydrogen sulphide during bacterial metabolism of dietary proteins. Carbohydrates are the preferred substrate for most colonic bacteria compared with protein. In the absence of dietary carbohydrates, colonic bacteria may degrade host mucin by formation of hydrogen sulphide (297). Red meat is known to be high in sulphur amino acids and a human study has shown that red meat consumption dose-dependently increases faecal sulphide (298). Moreover, recent evidence indicates that increased protein fermentation as a consequence of a high protein diet can produce harmful compounds such as phenol, cresol, indoles, amines and ammonia (104). High concentrations of these compounds are toxic to the epithelium and may promote genomic instability. A recent rodent study of AOM-induced tumours demonstrated that undigested protein promoted adenocarcinoma in the small intestine (299). However, these toxic by-products of protein metabolism were reduced when RS was added to the diet (116, 300). The studies in this thesis support this and showed significant reduction of phenol and *p*-cresol, which are implicated in bowel cancer (116), in both caecum and faeces with RS diets. Accumulation of these harmful byproducts of protein metabolism may be reduced by the fermentation of carbohydrate. Moreover, lowering of colonic pH through acidification via increased SCFA may limit the production of ammonia. Further investigations are necessary to determine how increased DNA damage caused by red meat is associated with increased risk of CRC.

Finally, this thesis has contributed some new information on the interactive effects of high protein diets and RS *in vivo*. High dietary protein, red meat in particular, appears to be harmful to the health of the large bowel, but addition of RS to the diet protects against this damage possibly via SCFA production and the maintenance of the colonic mucus layer thickness. One of the unanswered questions is the absence of a substantial genotoxic effect of whey and the relatively smaller effect of white meat. The fact that soy induced SSB without loss of mucus barrier also needs to be investigated. The potential for RS to be effective in maintenance of the large bowel health is also of considerable interest and this and its interaction with dietary protein is an area needing more research.

# REFERENCES

Moran BJ, Jackson AA. Function of the human colon. Br J Surg 1992;79(11):1132 7.

2. Cummings JG. The colon: Absorptive, seccretory and metabolic functions. Digestion 1975;13(4):232-40.

3. Cummings JH. Absorption and secretion by the colon. Gut 1975;16(4):323-9.

4. Shetty K, Rybicki L, Brzezinski A, Carey WD, Lashner BA. The risk for cancer or dysplasia in ulcerative colitis patients with primary sclerosing cholangitis. Am J Gastroenterol 1999;94(6):1643-9.

5. Lashner BA, Silverstein MD, Hanauer SB. Hazard rates for dysplasia and cancer in ulcerative colitis. Results from a surveillance program. Dig Dis Sci 1989;34(10):1536-41.

6. Friedman S, Rubin PH, Bodian C, Goldstein E, Harpaz N, Present DH. Screening and surveillance colonoscopy in chronic Crohn's colitis. Gastroenterology 2001;120(4):820-6.

7. Shapiro BD, Lashner BA. Cancer biology in ulcerative colitis and potential use in endoscopic surveillance. Gastrointest Endosc Clin N Am 1997;7(3):453-68.

8. Jarnerot G, Jarnmark I, Nilsson K. Consumption of refined sugar by patients with Crohn's disease, ulcerative colitis, or irritable bowel syndrome. Scand J Gastroenterol 1983;18(8):999-1002.

9. D'Haens G. Risks and benefits of biologic therapy for inflammatory bowel diseases. Gut 2007;56(5):725-32.

10. Makharia GK. Rising incidence and prevalence of Crohn's disease in Asia: is it apparent or real? J Gastroenterol Hepatol 2006;21(6):929-31.

11. Yang SK, Loftus EV, Jr., Sandborn WJ. Epidemiology of inflammatory bowel disease in Asia. Inflamm Bowel Dis 2001;7(3):260-70.

12. Loftus EV, Jr. Clinical epidemiology of inflammatory bowel disease: Incidence, prevalence, and environmental influences. Gastroenterology 2004;126(6):1504-17.

13. Association ACaC. Canberra, Australia 2006.

Podolsky DK. The current future understanding of inflammatory bowel disease.Best Pract Res Clin Gastroenterol 2002;16(6):933-43.

15. Hanauer SB. Inflammatory bowel disease. N Engl J Med 1996;334(13):841-8.

16. Blonski W, Lichtenstein GR. Complications of biological therapy for inflammatory bowel diseases. Curr Opin Gastroenterol 2006;22(1):30-43.

17. Midgley R, Kerr D. Colorectal cancer. Lancet 1999;353(9150):391-9.

 Sung JJ, Lau JY, Goh KL, Leung WK. Increasing incidence of colorectal cancer in Asia: implications for screening. Lancet Oncol 2005;6(11):871-6.

19. Tamura K, Ishiguro S, Munakata A, Yoshida Y, Nakaji S, Sugawara K. Annual changes in colorectal carcinoma incidence in Japan. Analysis of survey data on incidence in Aomori Prefecture. Cancer 1996;78(6):1187-94.

20. Stewart BW, Kleihues, P. World cancer report. IARC 2003;Press.

21. (AACR) AIoHaWAaAAoC. Cancer in Australia 2004. Canberra, Australia 2006.

22. Potter JD. Nutrition and colorectal cancer. Cancer Causes Control 1996;7(1):127-46.

23. Doll R, Peto R. The causes of cancer: quantitative estimates of avoidable risks of cancer in the United States today. J Natl Cancer Inst 1981;66(6):1191-308.

24. Willett WC. Diet, nutrition, and avoidable cancer. Environ Health Perspect 1995;103 Suppl 8:165-70.

25. Allinger UG, Johansson GK, Gustafsson JA, Rafter JJ. Shift from a mixed to a lactovegetarian diet: influence on acidic lipids in fecal water--a potential risk factor for colon cancer. Am J Clin Nutr 1989;50(5):992-6.

26. Glinghammar B, Venturi M, Rowland IR, Rafter JJ. Shift from a dairy product-rich to a dairy product-free diet: influence on cytotoxicity and genotoxicity of fecal water--potential risk factors for colon cancer. Am J Clin Nutr 1997;66(5):1277-82.

27. Venturi M, Hambly RJ, Glinghammar B, Rafter JJ, Rowland IR. Genotoxic activity in human faecal water and the role of bile acids: a study using the alkaline comet assay. Carcinogenesis 1997;18(12):2353-9.

28. Bignold LP. The mutator phenotype theory of carcinogenesis and the complex histopathology of tumours: support for the theory from the independent occurrence of nuclear abnormality, loss of specialisation and invasiveness among occasional neoplastic lesions. Cell Mol Life Sci 2003;60(5):883-91.

29. Vogelstein B, Fearon ER, Hamilton SR, Kern SE, Preisinger AC, Leppert M, et al.
Genetic alterations during colorectal-tumor development. N Engl J Med 1988;319(9):52532.

30. Sharma RA, Manson MM, Gescher A, Steward WP. Colorectal cancer chemoprevention: biochemical targets and clinical development of promising agents. Eur J Cancer 2001;37(1):12-22.

31. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. Cell 1990;61(5):759-67.

32. Narayan S, Roy D. Role of APC and DNA mismatch repair genes in the development of colorectal cancers. Mol Cancer 2003;2:41.

33. Kinzler KW, Vogelstein B. Lessons from hereditary colorectal cancer. Cell

References

1996;87(2):159-70.

34. Augenlicht LH, Mariadason JM, Wilson A, Arango D, Yang W, Heerdt BG, et al. Short chain fatty acids and colon cancer. J Nutr 2002;132(12):3804S-3808S.

35. Fodde R, Kuipers J, Rosenberg C, Smits R, Kielman M, Gaspar C, et al. Mutations in the APC tumour suppressor gene cause chromosomal instability. Nat Cell Biol 2001;3(4):433-8.

36. He TC, Sparks AB, Rago C, Hermeking H, Zawel L, da Costa LT, et al. Identification of c-MYC as a target of the APC pathway. Science 1998;281(5382):1509-12.

37. Hermeking H, Rago C, Schuhmacher M, Li Q, Barrett JF, Obaya AJ, et al.
Identification of CDK4 as a target of c-MYC. Proc Natl Acad Sci U S A 2000;97(5):222934.

38. Carson DA, Ribeiro JM. Apoptosis and disease. Lancet 1993;341(8855):1251-4.

39. Preston-Martin S, Pike MC, Ross RK, Jones PA, Henderson BE. Increased cell division as a cause of human cancer. Cancer Res 1990;50(23):7415-21.

40. Campbell SL, Khosravi-Far R, Rossman KL, Clark GJ, Der CJ. Increasing complexity of Ras signaling. Oncogene 1998;17(11 Reviews):1395-413.

41. Soussi T. The p53 tumor suppressor gene: from molecular biology to clinical investigation. Ann N Y Acad Sci 2000;910:121-37; discussion 137-9.

42. Nakagama H, Nakanishi M, Ochiai M. Modeling human colon cancer in rodents using a food-borne carcinogen, PhIP. Cancer Sci 2005;96(10):627-36.

43. Gorgoulis VG, Vassiliou LV, Karakaidos P, Zacharatos P, Kotsinas A, Liloglou T, et al. Activation of the DNA damage checkpoint and genomic instability in human precancerous lesions. Nature 2005;434(7035):907-13.

44. Bartkova J, Horejsi Z, Koed K, Kramer A, Tort F, Zieger K, et al. DNA damage response as a candidate anti-cancer barrier in early human tumorigenesis. Nature 2005;434(7035):864-70.

45. Kastan MB, Bartek J. Cell-cycle checkpoints and cancer. Nature 2004;432(7015):316-23.

46. Shiloh Y. ATM and related protein kinases: safeguarding genome integrity. Nat Rev Cancer 2003;3(3):155-68.

47. Hoeijmakers JH. Genome maintenance mechanisms for preventing cancer. Nature 2001;411(6835):366-74.

48. Grady WM. Genomic instability and colon cancer. Cancer Metastasis Rev 2004;23(1-2):11-27.

49. de la Chapelle A. Microsatellite instability. N Engl J Med 2003;349(3):209-10.

50. Richards RI. Fragile and unstable chromosomes in cancer: causes and consequences. Trends Genet 2001;17(6):339-45.

51. Young GP, Hu Y, Le Leu RK, Nyskohus L. Dietary fibre and colorectal cancer: a model for environment--gene interactions. Mol Nutr Food Res 2005;49(6):571-84.

52. van Gent DC, Hoeijmakers JH, Kanaar R. Chromosomal stability and the DNA double-stranded break connection. Nat Rev Genet 2001;2(3):196-206.

53. Lengauer C, Kinzler KW, Vogelstein B. Genetic instabilities in human cancers. Nature 1998;396(6712):643-9.

Beland FA, Marques MM. DNA adducts of nitropolycyclic aromatic hydrocarbons.
 IARC Sci Publ 1994(125):229-44.

55. Hochstein P, Atallah AS. The nature of oxidants and antioxidant systems in the inhibition of mutation and cancer. Mutat Res 1988;202(2):363-75.

56. Harrison KL, Jukes R, Cooper DP, Shuker DE. Detection of concomitant formation of O6-carboxymethyl- and O6-methyl-2'-deoxyguanosine in DNA exposed to nitrosated glycine derivatives using a combined immunoaffinity/HPLC method. Chem Res Toxicol 1999;12(1):106-11.

57. Lewin MH, Bailey N, Bandaletova T, Bowman R, Cross AJ, Pollock J, et al. Red meat enhances the colonic formation of the DNA adduct O6-carboxymethyl guanine: implications for colorectal cancer risk. Cancer Res 2006;66(3):1859-65.

58. Shuker DE, Margison GP. Nitrosated glycine derivatives as a potential source of O6-methylguanine in DNA. Cancer Res 1997;57(3):366-9.

59. Challis BC. Chemistry and biology of nitrosated peptides. Cancer Surv 1989;8(2):363-84.

60. Mirvish SS. Role of N-nitroso compounds (NOC) and N-nitrosation in etiology of gastric, esophageal, nasopharyngeal and bladder cancer and contribution to cancer of known exposures to NOC. Cancer Lett 1995;93(1):17-48.

61. Gottschalg E, Scott GB, Burns PA, Shuker DE. Potassium diazoacetate-induced p53 mutations in vitro in relation to formation of O6-carboxymethyl- and O6-methyl-2'-deoxyguanosine DNA adducts: relevance for gastrointestinal cancer. Carcinogenesis 2007;28(2):356-62.

62. Chang WL, Chapkin RS, Lupton JR. Fish oil blocks azoxymethane-induced rat colon tumorigenesis by increasing cell differentiation and apoptosis rather than decreasing cell proliferation. J Nutr 1998;128(3):491-7.

63. Khanna KK, Jackson SP. DNA double-strand breaks: signaling, repair and the cancer connection. Nat Genet 2001;27(3):247-54.

64. Berwick M, Vineis P. Markers of DNA repair and susceptibility to cancer in

humans: an epidemiologic review. J Natl Cancer Inst 2000;92(11):874-97.

65. Milner JA, McDonald SS, Anderson DE, Greenwald P. Molecular targets for nutrients involved with cancer prevention. Nutr Cancer 2001;41(1-2):1-16.

66. German JB, Roberts MA, Fay L, Watkins SM. Metabolomics and individual metabolic assessment: the next great challenge for nutrition. J Nutr 2002;132(9):2486-7.

67. Ahn J, Gammon MD, Santella RM, Gaudet MM, Britton JA, Teitelbaum SL, et al. Associations between breast cancer risk and the catalase genotype, fruit and vegetable consumption, and supplement use. Am J Epidemiol 2005;162(10):943-52.

68. Ferrari P, Al-Delaimy WK, Slimani N, Boshuizen HC, Roddam A, Orfanos P, et al. An approach to estimate between- and within-group correlation coefficients in multicenter studies: plasma carotenoids as biomarkers of intake of fruits and vegetables. Am J Epidemiol 2005;162(6):591-8.

69. Ames BN. DNA damage from micronutrient deficiencies is likely to be a major cause of cancer. Mutat Res 2001;475(1-2):7-20.

70. Michaud DS, Pietinen P, Taylor PR, Virtanen M, Virtamo J, Albanes D. Intakes of fruits and vegetables, carotenoids and vitamins A, E, C in relation to the risk of bladder cancer in the ATBC cohort study. Br J Cancer 2002;87(9):960-5.

71. Genkinger JM, Platz EA, Hoffman SC, Comstock GW, Helzlsouer KJ. Fruit, vegetable, and antioxidant intake and all-cause, cancer, and cardiovascular disease mortality in a community-dwelling population in Washington County, Maryland. Am J Epidemiol 2004;160(12):1223-33.

72. Kiefer I, Prock P, Lawrence C, Wise J, Bieger W, Bayer P, et al. Supplementation with mixed fruit and vegetable juice concentrates increased serum antioxidants and folate in healthy adults. J Am Coll Nutr 2004;23(3):205-11.

References

73. Waladkhani AR, Clemens MR. Effect of dietary phytochemicals on cancer development (review). Int J Mol Med 1998;1(4):747-53.

74. Topping DL, Clifton PM. Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. Physiol Rev 2001;81(3):1031-64.

75. Obrador A. Fibre and colorectal cancer: a controversial question. Br J Nutr 2006;96 Suppl 1:S46-8.

76. Rock CL. Primary dietary prevention: is the fiber story over? Recent Results Cancer Res 2007;174:171-7.

77. Schatzkin A, Mouw T, Park Y, Subar AF, Kipnis V, Hollenbeck A, et al. Dietary fiber and whole-grain consumption in relation to colorectal cancer in the NIH-AARP Diet and Health Study. Am J Clin Nutr 2007;85(5):1353-60.

78. Fuchs CS, Giovannucci EL, Colditz GA, Hunter DJ, Stampfer MJ, Rosner B, et al. Dietary fiber and the risk of colorectal cancer and adenoma in women. N Engl J Med 1999;340(3):169-76.

79. Jacobs ET, Lanza E, Alberts DS, Hsu CH, Jiang R, Schatzkin A, et al. Fiber, sex, and colorectal adenoma: results of a pooled analysis. Am J Clin Nutr 2006;83(2):343-9.

80. Wark PA, Weijenberg MP, van 't Veer P, van Wijhe G, Luchtenborg M, van Muijen GN, et al. Fruits, vegetables, and hMLH1 protein-deficient and -proficient colon cancer: The Netherlands cohort study. Cancer Epidemiol Biomarkers Prev 2005;14(7):1619-25.

81. Bingham SA, Day NE, Luben R, Ferrari P, Slimani N, Norat T, et al. Dietary fibre in food and protection against colorectal cancer in the European Prospective Investigation into Cancer and Nutrition (EPIC): an observational study. Lancet 2003;361(9368):1496-501.

82. Wakai K, Date C, Fukui M, Tamakoshi K, Watanabe Y, Hayakawa N, et al. Dietary fiber and risk of colorectal cancer in the Japan collaborative cohort study. Cancer Epidemiol Biomarkers Prev 2007;16(4):668-75.

Kim YI. AGA technical review: impact of dietary fiber on colon cancer occurrence.Gastroenterology 2000;118(6):1235-57.

84. Cassidy A, Bingham, S.A., Cummings, J.H. Starch intake and colorectal cancer risk: an international comparison. Br J Cancer 1994(May;69(5)):937-42.

85. Bartsch H, Nair J, Owen RW. Exocyclic DNA adducts as oxidative stress markers in colon carcinogenesis: potential role of lipid peroxidation, dietary fat and antioxidants. Biol Chem 2002;383(6):915-21.

86. Bartsch H, Nair J, Owen RW. Dietary polyunsaturated fatty acids and cancers of the breast and colorectum: emerging evidence for their role as risk modifiers. Carcinogenesis 1999;20(12):2209-18.

87. Norat T, Lukanova, A., Ferrari, P., Riboli, E. Meat consumption and colorectal cancer risk: an estimate of attributable and preventable fractions. IARC Sci Publ 2002;156:223-5.

88. Rieger MA, Parlesak, A., Pool-Zobel, B.L., Rechkemmer, G., Bode, C. A diet high in fat and meat but low in dietary fibre increases the genotoxic potential of 'faecal water'. Carcinogenesis 1999;20(12)(Dec):2311-6.

89. Chao A, Thun MJ, Connell CJ, McCullough ML, Jacobs EJ, Flanders WD, et al. Meat consumption and risk of colorectal cancer. Jama 2005;293(2):172-82.

90. Norat T, Bingham S, Ferrari P, Slimani N, Jenab M, Mazuir M, et al. Meat, fish, and colorectal cancer risk: the European Prospective Investigation into cancer and nutrition. J Natl Cancer Inst 2005;97(12):906-16.

91. Larsson SC, Rafter J, Holmberg L, Bergkvist L, Wolk A. Red meat consumption and risk of cancers of the proximal colon, distal colon and rectum: the Swedish Mammography Cohort. Int J Cancer 2005;113(5):829-34.

92. Visek WJ, Clinton SK, Imrey PB, Thursh DR, Truex CR, Alster JM, et al. Dietary protein and chronic toxicity of 1,2-dimethylhydrazine fed to mice. J Toxicol Environ Health 1991;32(4):383-413.

93. Belobrajdic DP, McIntosh, G.H., Owens, J.A. Whey proteins protect more than red meat against azoxymethane induced ACF in Wistar rats. Cancer Lett 2003;198(1)(Jul 30):43-51.

94. Yang L, Mutanen, M., Cheng, Y., Duan, R.D. Effects of fat, beef and fiber in diets on activities of sphingomyelinase, ceramidase and caspase-3 in rat colonic mucosa. Med Princ Pract 2002;11(3)(Jul-Sep):150-6.

95. McKeown-Eyssen GE, Bright-See, E. Dietary factors in colon cancer: international relationships. An update. Nutr Cancer 1985;7(4):251-3.

96. McIntosh GH, Le Leu, R.K. The influence of dietary proteins on colon cancer risk. Nutr Res 2001(Jul;21(7)):1053-1066.

97. Skov AR, Toubro S, Ronn B, Holm L, Astrup A. Randomized trial on protein vs carbohydrate in ad libitum fat reduced diet for the treatment of obesity. Int J Obes Relat Metab Disord 1999;23(5):528-36.

98. Morita T, Kasaoka S, Kiriyama S. Physiological functions of resistant proteins: proteins and peptides regulating large bowel fermentation of indigestible polysaccharide. J AOAC Int 2004;87(3):792-6.

99. Le Leu RK, Brown IL, Hu Y, Morita T, Esterman A, Young GP. Effect of Dietary Resistant Starch and Protein on Colonic Fermentation and Intestinal Tumourigenesis in Rats. Carcinogenesis 2006.

100. Cassidy A, Bingham SA, Cummings JH. Starch intake and colorectal cancer risk: an international comparison. Br J Cancer 1994;69(5):937-42.

101. Gibson JA, Sladen GE, Dawson AM. Protein absorption and ammonia production: the effects of dietary protein and removal of the colon. Br J Nutr 1976;35(1):61-5.

102. Porter JW, Rolls BA. Some aspects of the digestion of proteins. Proc Nutr Soc 1971;30(1):17-25.

103. Lin HC, Visek WJ. Large intestinal pH and ammonia in rats: dietary fat and protein interactions. J Nutr 1991;121(6):832-43.

104. Visek WJ. Diet and cell growth modulation by ammonia. Am J Clin Nutr 1978;31(10 Suppl):S216-S220.

105. Russell JB, Sniffen CJ, Van Soest PJ. Effect of carbohydrate limitation on degradation and utilization of casein by mixed rumen bacteria. J Dairy Sci 1983;66(4):763-75.

106. McBurney MI, Van Soest PJ, Jeraci JL. Colonic carcinogenesis: the microbial feast or famine mechanism. Nutr Cancer 1987;10(1-2):23-8.

107. Bingham SA. Meat, starch, and nonstarch polysaccharides and large bowel cancer.Am J Clin Nutr 1988;48(3 Suppl):762-7.

108. Gibson GR, Roberfroid, M.B. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. J Nutr 1995;125(6)(Jun):1401-12.

109. Ferguson LR. Meat consumption, cancer risk and population groups within New Zealand. Mutat Res 2002;506-507:215-24.

110. Potter JD. Colorectal cancer: molecules and populations. J Natl Cancer Inst 1999;91(11):916-32.

111. Silvester KR, Bingham SA, Pollock JR, Cummings JH, O'Neill IK. Effect of meat and resistant starch on fecal excretion of apparent N-nitroso compounds and ammonia from the human large bowel. Nutr Cancer 1997;29(1):13-23.

112. Hill MJ. Diet and the human intestinal bacterial flora. Cancer Res 1981;41(9 Pt 2):3778-80.

113. Pool-Zobel BL, Lotzmann N, Knoll M, Kuchenmeister F, Lambertz R, Leucht U, et al. Detection of genotoxic effects in human gastric and nasal mucosa cells isolated from biopsy samples. Environ Mol Mutagen 1994;24(1):23-45.

114. Boutwell RK, Bosch DK. The tumor-promoting action of phenol and related compounds for mouse skin. Cancer Res 1959;19(4):413-24.

115. Bryan GT. The role of urinary tryptophan metabolites in the etiology of bladder cancer. Am J Clin Nutr 1971;24(7):841-7.

116. Bone E, Tamm A, Hill M. The production of urinary phenols by gut bacteria and their possible role in the causation of large bowel cancer. Am J Clin Nutr 1976;29(12):1448-54.

117. Jain M, Cook GM, Davis FG, Grace MG, Howe GR, Miller AB. A case-control study of diet and colo-rectal cancer. Int J Cancer 1980;26(6):757-68.

118. Willett WC, Stampfer MJ, Colditz GA, Rosner BA, Speizer FE. Relation of meat, fat, and fiber intake to the risk of colon cancer in a prospective study among women. N Engl J Med 1990;323(24):1664-72.

119. Suzuki K, Mitsuoka T. Increase in faecal nitrosamines in Japanese individuals given a Western diet. Nature 1981;294(5840):453-6.

120. Hill MJ, Drasar DS. Bacteria and the aetiology of human cancer. Br J Cancer 1973;28(1):94.

121. Layton DW, Bogen KT, Knize MG, Hatch FT, Johnson VM, Felton JS. Cancer risk of heterocyclic amines in cooked foods: an analysis and implications for research. Carcinogenesis 1995;16(1):39-52.

Berg I, Overvik E, Gustafsson JA. Effect on cooking time on mutagen formation in smoke, crust and pan residue from pan-broiled pork. Food Chem Toxicol 1990;28(6):4216.

123. Pfau W, Martin FL, Cole KJ, Venitt S, Phillips DH, Grover PL, et al. Heterocyclic aromatic amines induce DNA strand breaks and cell transformation. Carcinogenesis 1999;20(4):545-51.

124. Ito N, Hasegawa R, Sano M, Tamano S, Esumi H, Takayama S, et al. A new colon and mammary carcinogen in cooked food, 2-amino-1-methyl-6-phenylimidazo[4,5b]pyridine (PhIP). Carcinogenesis 1991;12(8):1503-6.

125. Burnouf D, Miturski R, Nagao M, Nakagama H, Nothisen M, Wagner J, et al. Early detection of 2-amino-1-methyl-6-phenylimidazo (4,5-b)pyridine(PhIP)-induced mutations within the Apc gene of rat colon. Carcinogenesis 2001;22(2):329-35.

126. Ransohoff DF, Lang CA. Screening for colorectal cancer. N Engl J Med 1991;325(1):37-41.

127. Hecht SS. Biochemistry, biology, and carcinogenicity of tobacco-specific Nnitrosamines. Chem Res Toxicol 1998;11(6):559-603.

128. Hecht SS, Hoffmann D. N-nitroso compounds and man: sources of exposure, endogenous formation and occurrence in body fluids. Eur J Cancer Prev 1998;7(2):165-6.

129. Bingham SA, Pignatelli B, Pollock JR, Ellul A, Malaveille C, Gross G, et al. Does increased endogenous formation of N-nitroso compounds in the human colon explain the association between red meat and colon cancer? Carcinogenesis 1996;17(3):515-23.

130. Bingham SA, Hughes R, Cross AJ. Effect of white versus red meat on endogenous N-nitrosation in the human colon and further evidence of a dose response. J Nutr 2002;132(11 Suppl):3522S-3525S.

131. Cross AJ, Pollock JR, Bingham SA. Haem, not protein or inorganic iron, is responsible for endogenous intestinal N-nitrosation arising from red meat. Cancer Res 2003;63(10):2358-60.

132. van Maanen JM, Moonen EJ, Maas LM, Kleinjans JC, van Schooten FJ. Formation of aromatic DNA adducts in white blood cells in relation to urinary excretion of 1-hydroxypyrene during consumption of grilled meat. Carcinogenesis 1994;15(10):2263-8.

133. Cross AJ, Sinha R. Meat-related mutagens/carcinogens in the etiology of colorectal cancer. Environ Mol Mutagen 2004;44(1):44-55.

134. Culp SJ, Gaylor DW, Sheldon WG, Goldstein LS, Beland FA. A comparison of the tumors induced by coal tar and benzo[a]pyrene in a 2-year bioassay. Carcinogenesis 1998;19(1):117-24.

135. Babbs CF. Free radicals and the etiology of colon cancer. Free Radic Biol Med 1990;8(2):191-200.

136. Nelson RL. Dietary iron and colorectal cancer risk. Free Radic Biol Med 1992;12(2):161-8.

137. Sesink AL, Termont DS, Kleibeuker JH, Van der Meer R. Red meat and colon cancer: the cytotoxic and hyperproliferative effects of dietary heme. Cancer Res 1999;59(22):5704-9.

138. Sawa T, Akaike T, Kida K, Fukushima Y, Takagi K, Maeda H. Lipid peroxyl radicals from oxidized oils and heme-iron: implication of a high-fat diet in colon carcinogenesis. Cancer Epidemiol Biomarkers Prev 1998;7(11):1007-12.

139. Pierre F, Tache S, Petit CR, Van der Meer R, Corpet DE. Meat and cancer: haemoglobin and haemin in a low-calcium diet promote colorectal carcinogenesis at the aberrant crypt stage in rats. Carcinogenesis 2003;24(10):1683-90.

140. Giovannucci E, Rimm EB, Stampfer MJ, Colditz GA, Ascherio A, Willett WC. Intake of fat, meat, and fiber in relation to risk of colon cancer in men. Cancer Res 1994;54(9):2390-7.

141. Singh PN, Fraser GE. Dietary risk factors for colon cancer in a low-risk population.Am J Epidemiol 1998;148(8):761-74.

142. Corpet DE, Chatelin-Pirot V. Cooked casein promotes colon cancer in rats, may be because of mucosal abrasion. Cancer Lett 1997;114(1-2):89-90.

143. Toden S, Bird AR, Topping DL, Conlon MA. Resistant starch attenuates colonic DNA damage induced by higher dietary protein in rats. Nutr Cancer 2005;51(1):45-51.

144. Corpet DE, Stamp D, Medline A, Minkin S, Archer MC, Bruce WR. Promotion of colonic microadenoma growth in mice and rats fed cooked sugar or cooked casein and fat. Cancer Res 1990;50(21):6955-8.

145. Corpet DE, Cassand P. Lack of aberrant crypt promotion and of mutagenicity in extracts of cooked casein, a colon cancer-promoting food. Nutr Cancer 1995;24(3):249-56.

146. Kennedy RS, Konok GP, Bounous G, Baruchel S, Lee TD. The use of a whey protein concentrate in the treatment of patients with metastatic carcinoma: a phase I-II clinical study. Anticancer Res 1995;15(6B):2643-9.

147. Bounous G. Whey protein concentrate (WPC) and glutathione modulation in cancer treatment. Anticancer Res 2000;20(6C):4785-92.

148. Meister A, Anderson ME. Glutathione. Annu Rev Biochem 1983;52:711-60.

149. Dekant W. Biosynthesis and cellular effects of toxic glutathione S-conjugates. Adv

Exp Med Biol 1996;387:297-312.

150. Meister A, Anderson ME, Hwang O. Intracellular cysteine and glutathione delivery systems. J Am Coll Nutr 1986;5(2):137-51.

151. Grodstein F, Newcomb PA, Stampfer MJ. Postmenopausal hormone therapy and the risk of colorectal cancer: a review and meta-analysis. Am J Med 1999;106(5):574-82.

152. Rossouw JE, Anderson GL, Prentice RL, LaCroix AZ, Kooperberg C, Stefanick ML, et al. Risks and benefits of estrogen plus progestin in healthy postmenopausal women: principal results From the Women's Health Initiative randomized controlled trial. Jama 2002;288(3):321-33.

153. Messina MJ, Persky V, Setchell KD, Barnes S. Soy intake and cancer risk: a review of the in vitro and in vivo data. Nutr Cancer 1994;21(2):113-31.

154. Hakkak R, Korourian S, Ronis MJ, Johnston JM, Badger TM. Soy protein isolate consumption protects against azoxymethane-induced colon tumors in male rats. Cancer Lett 2001;166(1):27-32.

155. Govers MJ, Lapre JA, De Vries HT, Van der Meer R. Dietary soybean protein compared with casein damages colonic epithelium and stimulates colonic epithelial proliferation in rats. J Nutr 1993;123(10):1709-13.

156. McIntosh GH, Regester GO, Le Leu RK, Royle PJ, Smithers GW. Dairy proteins protect against dimethylhydrazine-induced intestinal cancers in rats. J Nutr 1995;125(4):809-16.

157. Wu AH, Yang D, Pike MC. A meta-analysis of soyfoods and risk of stomach cancer: the problem of potential confounders. Cancer Epidemiol Biomarkers Prev 2000;9(10):1051-8.

158. Spector D, Anthony M, Alexander D, Arab L. Soy consumption and colorectal

cancer. Nutr Cancer 2003;47(1):1-12.

159. Badger TM, Ronis MJ, Simmen RC, Simmen FA. Soy protein isolate and protection against cancer. J Am Coll Nutr 2005;24(2):146S-149S.

160. Burkitt DP. Relationship as a clue to causation. Lancet 1970;2(7685):1237-40.

161. Burkitt DP. Some diseases characteristic of modern Western civilization. Br Med J1973;1(5848):274-8.

162. McIntyre A, Young GP, Taranto T, Gibson PR, Ward PB. Different fibers have different regional effects on luminal contents of rat colon. Gastroenterology 1991;101(5):1274-81.

163. Folino M, McIntyre A, Young GP. Dietary fibers differ in their effects on large bowel epithelial proliferation and fecal fermentation-dependent events in rats. J Nutr 1995;125(6):1521-8.

164. Topping DL, Illman RJ. Bacterial fermentation in the human large bowel. Time to change from the roughage model of dietary fibre? Med J Aust 1986;144(6):307-9.

165. Stephen AM, Cummings JH. Mechanism of action of dietary fibre in the human colon. Nature 1980;284(5753):283-4.

166. Birkett A, Muir, J., Phillips, J., Jones, G., O'Dea, K. Resistant starch lowers fecal concentrations of ammonia and phenols in humans. Am J Clin Nutr 1996;63(5)(May):766-72.

167. Savage DC. Gastrointestinal microflora in mammalian nutrition. Annu Rev Nutr 1986;6:155-78.

168. Topping DL, Clifton, P.M. Short-chain fatty acids and human colonic function:
roles of resistant starch and nonstarch polysaccharides. Physiol Rev 2001(Jul;81(3)):103164.

169. Jacobs LR. Fiber and colon cancer. Gastroenterol Clin North Am 1988;17(4):747-60.

170. Thornton JR. High colonic pH promotes colorectal cancer. Lancet 1981;1(8229):1081-3.

171. Walker AR, Walker BF, Walker AJ. Faecal pH, dietary fibre intake, and proneness to colon cancer in four South African populations. Br J Cancer 1986;53(4):489-95.

172. Giovannucci E. Insulin and colon cancer. Cancer Causes Control 1995;6(2):164-79.

173. Weiderpass E, Gridley G, Nyren O, Ekbom A, Persson I, Adami HO. Diabetes mellitus and risk of large bowel cancer. J Natl Cancer Inst 1997;89(9):660-1.

174. Potter JD, Slattery ML, Bostick RM, Gapstur SM. Colon cancer: a review of the epidemiology. Epidemiol Rev 1993;15(2):499-545.

175. Giovannucci E, Willett WC. Dietary factors and risk of colon cancer. Ann Med 1994;26(6):443-52.

176. Steinmetz KA, Potter JD. Vegetables, fruit, and cancer prevention: a review. J Am Diet Assoc 1996;96(10):1027-39.

177. Englyst HN, Trowell H, Southgate DA, Cummings JH. Dietary fiber and resistant starch. Am J Clin Nutr 1987;46(6):873-4.

178. Englyst HN, Kingman SM, Cummings JH. Classification and measurement of nutritionally important starch fractions. Eur J Clin Nutr 1992;46 Suppl 2:S33-50.

179. Topping DL, Fukushima M, Bird AR. Resistant starch as a prebiotic and synbiotic: state of the art. Proc Nutr Soc 2003;62(1):171-6.

180. Ahmed R, Segal I, Hassan H. Fermentation of dietary starch in humans. Am J Gastroenterol 2000;95(4):1017-20.

181. Segal I, Solomon A, Hunt JA. Emergence of diverticular disease in the urban South

African black. Gastroenterology 1977;72(2):215-9.

Bourne LT, Langenhoven ML, Steyn K, Jooste PL, Laubscher JA, Van der VyverE. Nutrient intake in the urban African population of the Cape Peninsula, South Africa.The Brisk study. Cent Afr J Med 1993;39(12):238-47.

183. Le Leu RK, Brown, I.L., Hu, Y., Young, G.P. Effect of resistant starch on genotoxin-induced apoptosis, colonic epithelium, and lumenal contents in rats. Carcinogenesis 2003;24(8)(Aug):1347-52.

184. Falk PG, Hooper LV, Midtvedt T, Gordon JI. Creating and maintaining the gastrointestinal ecosystem: what we know and need to know from gnotobiology. Microbiol Mol Biol Rev 1998;62(4):1157-70.

185. Guarner F, Malagelada JR. Gut flora in health and disease. Lancet 2003;361(9356):512-9.

186. Rafter J, Glinghammar B. Interactions between the environment and genes in the colon. Eur J Cancer Prev 1998;7 Suppl 2:S69-74.

187. Rafter J. Lactic acid bacteria and cancer: mechanistic perspective. Br J Nutr2002;88 Suppl 1:S89-94.

188. Onoue M, Kado S, Sakaitani Y, Uchida K, Morotomi M. Specific species of intestinal bacteria influence the induction of aberrant crypt foci by 1,2-dimethylhydrazine in rats. Cancer Lett 1997;113(1-2):179-86.

189. Horie H, Kanazawa K, Kobayashi E, Okada M, Fujimura A, Yamagiwa S, et al. Effects of intestinal bacteria on the development of colonic neoplasm II. Changes in the immunological environment. Eur J Cancer Prev 1999;8(6):533-7.

190. Singh J, Rivenson A, Tomita M, Shimamura S, Ishibashi N, Reddy BS. Bifidobacterium longum, a lactic acid-producing intestinal bacterium inhibits colon cancer

and modulates the intermediate biomarkers of colon carcinogenesis. Carcinogenesis 1997;18(4):833-41.

191. Pool-Zobel BL, Neudecker C, Domizlaff I, Ji S, Schillinger U, Rumney C, et al. Lactobacillus- and bifidobacterium-mediated antigenotoxicity in the colon of rats. Nutr Cancer 1996;26(3):365-80.

192. Kleessen B, Stoof G, Proll J, Schmiedl D, Noack J, Blaut M. Feeding resistant starch affects fecal and cecal microflora and short-chain fatty acids in rats. J Anim Sci 1997;75(9):2453-62.

193. Jacobasch G, Schmiedl D, Kruschewski M, Schmehl K. Dietary resistant starch and chronic inflammatory bowel diseases. Int J Colorectal Dis 1999;14(4-5):201-11.

194. Mulller V. Bacterial Fermentation. Encyclopedia of Life Sciences 2001:1-7.

195. Burkitt DP. Related disease--related cause? Lancet 1969;2(7632):1229-31.

196. Cook SI, Sellin JH. Review article: short chain fatty acids in health and disease. Aliment Pharmacol Ther 1998;12(6):499-507.

197. Scheppach W, Fabian C, Sachs M, Kasper H. The effect of starch malabsorption on fecal short-chain fatty acid excretion in man. Scand J Gastroenterol 1988;23(6):755-9.

198. Noakes M, Clifton PM, Nestel PJ, Le Leu R, McIntosh G. Effect of high-amylose starch and oat bran on metabolic variables and bowel function in subjects with hypertriglyceridemia. Am J Clin Nutr 1996;64(6):944-51.

199. van Munster IP, Tangerman A, Nagengast FM. Effect of resistant starch on colonic fermentation, bile acid metabolism, and mucosal proliferation. Dig Dis Sci 1994;39(4):834-42.

200. Andrieux C, Pacheco ED, Bouchet B, Gallant D, Szylit O. Contribution of the digestive tract microflora to amylomaize starch degradation in the rat. Br J Nutr

139

1992;67(3):489-99.

201. Bird AR, Hayakawa, T., Marsono, Y., Gooden, J.M., Record, I.R., Correll, R.L., Topping, D.L. Coarse brown rice increases fecal and large bowel short-chain fatty acids and starch but lowers calcium in the large bowel of pigs. J Nutr 2000;130(7)(Jul):1780-7.

202. Brown I, Warhurst M, Arcot J, Playne M, Illman RJ, Topping DL. Fecal numbers of bifidobacteria are higher in pigs fed Bifidobacterium longum with a high amylose cornstarch than with a low amylose cornstarch. J Nutr 1997;127(9):1822-7.

203. Topping DL, Gooden JM, Brown IL, Biebrick DA, McGrath L, Trimble RP, et al. A high amylose (amylomaize) starch raises proximal large bowel starch and increases colon length in pigs. J Nutr 1997;127(4):615-22.

204. Hague A, Elder DJ, Hicks DJ, Paraskeva C. Apoptosis in colorectal tumour cells: induction by the short chain fatty acids butyrate, propionate and acetate and by the bile salt deoxycholate. Int J Cancer 1995;60(3):400-6.

205. Heerdt BG, Houston MA, Augenlicht LH. Short-chain fatty acid-initiated cell cycle arrest and apoptosis of colonic epithelial cells is linked to mitochondrial function. Cell Growth Differ 1997;8(5):523-32.

206. Heerdt BG, Halsey HK, Lipkin M, Augenlicht LH. Expression of mitochondrial cytochrome c oxidase in human colonic cell differentiation, transformation, and risk for colonic cancer. Cancer Res 1990;50(5):1596-600.

207. Augenlicht LH, Anthony GM, Church TL, Edelmann W, Kucherlapati R, Yang K, et al. Short-chain fatty acid metabolism, apoptosis, and Apc-initiated tumorigenesis in the mouse gastrointestinal mucosa. Cancer Res 1999;59(23):6005-9.

208. Hassig CA, Tong JK, Schreiber SL. Fiber-derived butyrate and the prevention of colon cancer. Chem Biol 1997;4(11):783-9.

140

209. Chen JS, Faller DV, Spanjaard RA. Short-chain fatty acid inhibitors of histone deacetylases: promising anticancer therapeutics? Curr Cancer Drug Targets 2003;3(3):219-36.

210. Lupton JR. Butyrate and colonic cytokinetics: differences between in vitro and in vivo studies. Eur J Cancer Prev 1995;4(5):373-8.

211. Freeman HJ. Effects of differing concentrations of sodium butyrate on 1,2dimethylhydrazine-induced rat intestinal neoplasia. Gastroenterology 1986;91(3):596-602.

212. Caderni G, Luceri C, Lancioni L, Tessitore L, Dolara P. Slow-release pellets of sodium butyrate increase apoptosis in the colon of rats treated with azoxymethane, without affecting aberrant crypt foci and colonic proliferation. Nutr Cancer 1998;30(3):175-81.

213. Sealy L, Chalkley R. The effect of sodium butyrate on histone modification. Cell 1978;14(1):115-21.

214. Le Blay G, Blottiere HM, Ferrier L, Le Foll E, Bonnet C, Galmiche JP, et al. Shortchain fatty acids induce cytoskeletal and extracellular protein modifications associated with modulation of proliferation on primary culture of rat intestinal smooth muscle cells. Dig Dis Sci 2000;45(8):1623-30.

215. Zemel MB. Calcium utilization: effect of varying level and source of dietary protein. Am J Clin Nutr 1988(Sep;48(3 Suppl)):880-3.

216. Atuma C, Strugala, V., Allen, A., Holm, L. The adherent gastrointestinal mucus gel layer: thickness and physical state in vivo. Am J Physiol Gastrointest Liver Physiol 2001(May;280(5)):G922-9.

217. Allen A, Bell, A., Mantle, M., Pearson, J.P. The structure and physiology of gastrointestinal mucus. Adv Exp Med Biol 1982;144:115-33.

218. Rozee KR, Cooper, D., Lam, K., Costerton, J.W. Microbial flora of the mouse

ileum mucous layer and epithelial surface. Appl Environ Microbiol 1982(Jun;43(6)):1451-63.

219. Strugala V, Allen A, Dettmar PW, Pearson JP. Colonic mucin: methods of measuring mucus thickness. Proc Nutr Soc 2003;62(1):237-43.

220. Brownlee IA, Havler ME, Dettmar PW, Allen A, Pearson JP. Colonic mucus: secretion and turnover in relation to dietary fibre intake. Proc Nutr Soc 2003;62(1):245-9.

221. Allen A, Leonard AJ, Sellers LA. The mucus barrier. Its role in gastroduodenal mucosal protection. J Clin Gastroenterol 1988;10 Suppl 1:S93-8.

222. Brownlee IA, Havler, M.E., Dettmar, P.W., Allen, A., Pearson, J.P. Colonic mucus: secretion and turnover in relation to dietary fibre intake. Proc Nutr Soc 2003(Feb;62(1)):245-9.

223. Byrd JC, Bresalier RS. Mucins and mucin binding proteins in colorectal cancer. Cancer Metastasis Rev 2004;23(1-2):77-99.

224. Shirazi T, Longman RJ, Corfield AP, Probert CS. Mucins and inflammatory bowel disease. Postgrad Med J 2000;76(898):473-8.

225. Chang SK, Dohrman AF, Basbaum CB, Ho SB, Tsuda T, Toribara NW, et al. Localization of mucin (MUC2 and MUC3) messenger RNA and peptide expression in human normal intestine and colon cancer. Gastroenterology 1994;107(1):28-36.

226. Weiss AA, Babyatsky MW, Ogata S, Chen A, Itzkowitz SH. Expression of MUC2 and MUC3 mRNA in human normal, malignant, and inflammatory intestinal tissues. J Histochem Cytochem 1996;44(10):1161-6.

227. Velcich A, Yang W, Heyer J, Fragale A, Nicholas C, Viani S, et al. Colorectal cancer in mice genetically deficient in the mucin Muc2. Science 2002;295(5560):1726-9.

228. Byrd JC, Yan P, Sternberg L, Yunker CK, Scheiman JM, Bresalier RS. Aberrant

expression of gland-type gastric mucin in the surface epithelium of Helicobacter pyloriinfected patients. Gastroenterology 1997;113(2):455-64.

229. Bartman AE, Sanderson SJ, Ewing SL, Niehans GA, Wiehr CL, Evans MK, et al. Aberrant expression of MUC5AC and MUC6 gastric mucin genes in colorectal polyps. Int J Cancer 1999;80(2):210-8.

230. Mandel JS, Bond JH, Bradley M, Snover DC, Church TR, Williams S, et al. Sensitivity, specificity, and positive predictivity of the Hemoccult test in screening for colorectal cancers. The University of Minnesota's Colon Cancer Control Study. Gastroenterology 1989;97(3):597-600.

231. Kronborg O, Fenger C, Olsen J, Jorgensen OD, Sondergaard O. Randomised study of screening for colorectal cancer with faecal-occult-blood test. Lancet 1996;348(9040):1467-71.

232. Hardcastle JD, Chamberlain JO, Robinson MH, Moss SM, Amar SS, Balfour TW, et al. Randomised controlled trial of faecal-occult-blood screening for colorectal cancer. Lancet 1996;348(9040):1472-7.

233. Brenner DE, Rennert G. Fecal DNA biomarkers for the detection of colorectal neoplasia: attractive, but is it feasible? J Natl Cancer Inst 2005;97(15):1107-9.

234. Imperiale TF, Ransohoff DF, Itzkowitz SH, Turnbull BA, Ross ME. Fecal DNA versus fecal occult blood for colorectal-cancer screening in an average-risk population. N Engl J Med 2004;351(26):2704-14.

235. Kashtan H, Stern HS. Colonic proliferation and colon cancer risk. A review of clinical studies. Isr J Med Sci 1992;28(12):904-10.

236. Einspahr JG, Alberts DS, Gapstur SM, Bostick RM, Emerson SS, Gerner EW. Surrogate end-point biomarkers as measures of colon cancer risk and their use in cancer chemoprevention trials. Cancer Epidemiol Biomarkers Prev 1997;6(1):37-48.

237. Hu Y, Martin J, Le Leu R, Young GP. The colonic response to genotoxic carcinogens in the rat: regulation by dietary fibre. Carcinogenesis 2002;23(7):1131-7.

238. Olive PL. The comet assay. An overview of techniques. Methods Mol Biol 2002;203:179-94.

239. Olive PL, Banath JP. The comet assay: a method to measure DNA damage in individual cells. Nature Protocol 2006;1(1):23-29.

240. Toden S, Bird AR, Topping DL, Conlon MA. Resistant starch attenuates colonic DNA damage induced by a high protein diet in rats. Asia Pac J Clin Nutr 2003;12 Suppl:S13.

241. McKelvey-Martin VJ, Green MH, Schmezer P, Pool-Zobel BL, De Meo MP, Collins A. The single cell gel electrophoresis assay (comet assay): a European review. Mutat Res 1993;288(1):47-63.

242. Fairbairn DW, Olive PL, O'Neill KL. The comet assay: a comprehensive review. Mutat Res 1995;339(1):37-59.

243. Olive PL, Wlodek D, Durand RE, Banath JP. Factors influencing DNA migration from individual cells subjected to gel electrophoresis. Exp Cell Res 1992;198(2):259-67.

244. Tice R, Agurell, E., Anderson, D., Burlinson, B., Hartmann, A., Kobayashi, H., Miyamae, Y., Rojas, E., Ryu, J.C., Sasaki, Y.F. Single cell gel/comet assay: guidelines for in vitro and in vivo genetic toxicology testing. Environ Mol Mutagen 2000(35(3)):206-21.

245. Olive PL, Wlodek D, Banath JP. DNA double-strand breaks measured in individual cells subjected to gel electrophoresis. Cancer Res 1991;51(17):4671-6.

246. Wojewodzka M, Buraczewska I, Kruszewski M. A modified neutral comet assay: elimination of lysis at high temperature and validation of the assay with anti-single-

stranded DNA antibody. Mutat Res 2002;518(1):9-20.

247. Collins AR, Duthie SJ, Dobson VL. Direct enzymic detection of endogenous oxidative base damage in human lymphocyte DNA. Carcinogenesis 1993;14(9):1733-5.

248. Xiao R, Badger TM, Simmen FA. Dietary exposure to soy or whey proteins alters colonic global gene expression profiles during rat colon tumorigenesis. Mol Cancer 2005;4(1):1.

249. Friedman M, Brandon DL. Nutritional and health benefits of soy proteins. J Agric Food Chem 2001;49(3):1069-86.

250. McIntosh GH, Le Leu RK. The influence of dietary proteins on colon cancer risk. Nutr Res 2001;21(7):1053-1066.

251. Tsuda H, Sekine K, Ushida Y, Kuhara T, Takasuka N, Iigo M, et al. Milk and dairy products in cancer prevention: focus on bovine lactoferrin. Mutat Res 2000;462(2-3):227-33.

252. Guo JY, Li X, Browning JD, Jr., Rottinghaus GE, Lubahn DB, Constantinou A, et al. Dietary soy isoflavones and estrone protect ovariectomized ERalphaKO and wild-type mice from carcinogen-induced colon cancer. J Nutr 2004;134(1):179-82.

253. Macdonald RS, Guo J, Copeland J, Browning JD, Jr., Sleper D, Rottinghaus GE, et al. Environmental influences on isoflavones and saponins in soybeans and their role in colon cancer. J Nutr 2005;135(5):1239-42.

254. Stoll BA. Eating to beat breast cancer: potential role for soy supplements. Ann Oncol 1997;8(3):223-5.

255. Ruiz-Larrea MB, Mohan AR, Paganga G, Miller NJ, Bolwell GP, Rice-Evans CA. Antioxidant activity of phytoestrogenic isoflavones. Free Radic Res 1997;26(1):63-70.

256. Mitchell JH, Collins AR. Effects of a soy milk supplement on plasma cholesterol

levels and oxidative DNA damage in men--a pilot study. Eur J Nutr 1999;38(3):143-8.

257. Lee SO, Simons AL, Murphy PA, Hendrich S. Soyasaponins lowered plasma cholesterol and increased fecal bile acids in female golden Syrian hamsters. Exp Biol Med (Maywood) 2005;230(7):472-8.

258. Narisawa T, Magadia NE, Weisburger JH, Wynder EL. Promoting effect of bile acids on colon carcinogenesis after intrarectal instillation of N-methyl-N'-nitro-N-nitrosoguanidine in rats. J Natl Cancer Inst 1974;53(4):1093-7.

259. Reddy BS, Watanabe K, Weisburger JH, Wynder EL. Promoting effect of bile acids in colon carcinogenesis in germ-free and conventional F344 rats. Cancer Res 1977;37(9):3238-42.

260. English DR, MacInnis RJ, Hodge AM, Hopper JL, Haydon AM, Giles GG. Red meat, chicken, and fish consumption and risk of colorectal cancer. Cancer Epidemiol Biomarkers Prev 2004;13(9):1509-14.

261. Emenaker NJ, Basson MD. Short chain fatty acids differentially modulate cellular phenotype and c-myc protein levels in primary human nonmalignant and malignant colonocytes. Dig Dis Sci 2001;46(1):96-105.

262. Singh B, Halestrap AP, Paraskeva C. Butyrate can act as a stimulator of growth or inducer of apoptosis in human colonic epithelial cell lines depending on the presence of alternative energy sources. Carcinogenesis 1997;18(6):1265-70.

263. McIntyre A, Gibson PR, Young GP. Butyrate production from dietary fibre and protection against large bowel cancer in a rat model. Gut 1993;34(3):386-91.

264. Scheppach W, Bartram HP, Richter F. Role of short-chain fatty acids in the prevention of colorectal cancer. Eur J Cancer 1995;31A(7-8):1077-80.

265. Hague A, Manning AM, Hanlon KA, Huschtscha LI, Hart D, Paraskeva C. Sodium

butyrate induces apoptosis in human colonic tumour cell lines in a p53-independent pathway: implications for the possible role of dietary fibre in the prevention of large-bowel cancer. Int J Cancer 1993;55(3):498-505.

266. Barnard JA, Warwick G. Butyrate rapidly induces growth inhibition and differentiation in HT-29 cells. Cell Growth Differ 1993;4(6):495-501.

267. Shimotoyodome A, Meguro S, Hase T, Tokimitsu I, Sakata T. Short chain fatty acids but not lactate or succinate stimulate mucus release in the rat colon. Comp Biochem Physiol A Mol Integr Physiol 2000;125(4):525-31.

268. Cummings JH, Macfarlane GT. The control and consequences of bacterial fermentation in the human colon. J Appl Bacteriol 1991;70(6):443-59.

269. Barcelo A, Claustre J, Moro F, Chayvialle JA, Cuber JC, Plaisancie P. Mucin secretion is modulated by luminal factors in the isolated vascularly perfused rat colon. Gut 2000;46(2):218-24.

270. Le Leu RK, Brown IL, Hu Y, Young GP. Effect of resistant starch on genotoxininduced apoptosis, colonic epithelium, and lumenal contents in rats. Carcinogenesis 2003;24(8):1347-52.

271. Birkett AM, Jones GP, de Silva AM, Young GP, Muir JG. Dietary intake and faecal excretion of carbohydrate by Australians: importance of achieving stool weights greater than 150 g to improve faecal markers relevant to colon cancer risk. Eur J Clin Nutr 1997;51(9):625-32.

272. Collins AR. Oxidative DNA damage, antioxidants, and cancer. Bioessays 1999;21(3):238-46.

273. Abrahamse SL, Pool-Zobel BL, Rechkemmer G. Potential of short chain fatty acids to modulate the induction of DNA damage and changes in the intracellular calcium

concentration by oxidative stress in isolated rat distal colon cells. Carcinogenesis 1999;20(4):629-34.

274. Specian RD, Oliver MG. Functional biology of intestinal goblet cells. Am J Physiol 1991;260(2 Pt 1):C183-93.

275. Rhodes JM. Colonic mucus and mucosal glycoproteins: the key to colitis and cancer? Gut 1989;30(12):1660-6.

276. Smith AC, Podolsky DK. Colonic mucin glycoproteins in health and disease. Clin Gastroenterol 1986;15(4):815-37.

277. Jowett SL, Seal CJ, Pearce MS, Phillips E, Gregory W, Barton JR, et al. Influence of dietary factors on the clinical course of ulcerative colitis: a prospective cohort study. Gut 2004;53(10):1479-84.

278. Sharma R, Schumacher U, Ronaasen V, Coates M. Rat intestinal mucosal responses to a microbial flora and different diets. Gut 1995;36(2):209-14.

279. Lundin E, Zhang JX, Huang CB, Reuterving CO, Hallmans G, Nygren C, et al. Oat bran, rye bran, and soybean hull increase goblet cell volume density in the small intestine of the golden hamster. A histochemical and stereologic light-microscopic study. Scand J Gastroenterol 1993;28(1):15-22.

280. Christl SU, Eisner HD, Dusel G, Kasper H, Scheppach W. Antagonistic effects of sulfide and butyrate on proliferation of colonic mucosa: a potential role for these agents in the pathogenesis of ulcerative colitis. Dig Dis Sci 1996;41(12):2477-81.

281. de Boer J, Hoeijmakers JH. Nucleotide excision repair and human syndromes. Carcinogenesis 2000;21(3):453-60.

282. Rotman G, Shiloh Y. ATM: from gene to function. Hum Mol Genet 1998;7(10):1555-63.

283. Corpet DE, Pierre F. How good are rodent models of carcinogenesis in predicting efficacy in humans? A systematic review and meta-analysis of colon chemoprevention in rats, mice and men. Eur J Cancer 2005;41(13):1911-22.

284. Stevens CE, Argenzio RA, Roberts MC. Comparative physiology of the mammalian colon and suggestions for animal models of human disorders. Clin Gastroenterol 1986;15(4):763-85.

285. Winawer SJ, Fletcher RH, Miller L, Godlee F, Stolar MH, Mulrow CD, et al. Colorectal cancer screening: clinical guidelines and rationale. Gastroenterology 1997;112(2):594-642.

286. Winawer SJ, Zauber AG, O'Brien MJ, Ho MN, Gottlieb L, Sternberg SS, et al. Randomized comparison of surveillance intervals after colonoscopic removal of newly diagnosed adenomatous polyps. The National Polyp Study Workgroup. N Engl J Med 1993;328(13):901-6.

287. Winawer SJ, Zauber AG, Ho MN, O'Brien MJ, Gottlieb LS, Sternberg SS, et al. Prevention of colorectal cancer by colonoscopic polypectomy. The National Polyp Study Workgroup. N Engl J Med 1993;329(27):1977-81.

288. Song K, Fendrick AM, Ladabaum U. Fecal DNA testing compared with conventional colorectal cancer screening methods: a decision analysis. Gastroenterology 2004;126(5):1270-9.

289. Osborn NK, Ahlquist DA. Stool screening for colorectal cancer: molecular approaches. Gastroenterology 2005;128(1):192-206.

290. Olive PL, Johnston PJ. DNA damage from oxidants: influence of lesion complexity and chromatin organization. Oncol Res 1997;9(6-7):287-94.

291. Thompson CB. Apoptosis in the pathogenesis and treatment of disease. Science

## 1995;267(5203):1456-62.

292. Le Leu RK, Brown IL, Hu Y, Bird AR, Jackson M, Esterman A, et al. A Synbiotic Combination of Resistant Starch and Bifidobacterium lactis Facilitates Apoptotic Deletion of Carcinogen-Damaged Cells in Rat Colon. J Nutr 2005;135(5):996-1001.

293. Bird AR, Brown IL, Topping DL. Starches, resistant starches, the gut microflora and human health. Curr Issues Intest Microbiol 2000;1(1):25-37.

294. Duncan SH, Belenguer A, Holtrop G, Johnstone AM, Flint HJ, Lobley GE. Reduced dietary intake of carbohydrates by obese subjects results in decreased concentrations of butyrate and butyrate-producing bacteria in feces. Appl Environ Microbiol 2007;73(4):1073-8.

295. Bajka BH, Topping DL, Cobiac L, Clarke JM. Butyrylated starch is less susceptible to enzymic hydrolysis and increases large-bowel butyrate more than high-amylose maize starch in the rat. Br J Nutr 2006;96(2):276-82.

296. Annison G, Illman RJ, Topping DL. Acetylated, propionylated or butyrylated starches raise large bowel short-chain fatty acids preferentially when fed to rats. J Nutr 2003;133(11):3523-8.

297. McGarr SE, Ridlon JM, Hylemon PB. Diet, anaerobic bacterial metabolism, and colon cancer: a review of the literature. J Clin Gastroenterol 2005;39(2):98-109.

298. Magee EA, Richardson CJ, Hughes R, Cummings JH. Contribution of dietary protein to sulfide production in the large intestine: an in vitro and a controlled feeding study in humans. Am J Clin Nutr 2000;72(6):1488-94.

299. Le Leu RK, Brown IL, Hu Y, Morita T, Esterman A, Young GP. Effect of dietary resistant starch and protein on colonic fermentation and intestinal tumourigenesis in rats. Carcinogenesis 2007;28(2):240-5.

150

300. Govers MJ, Gannon NJ, Dunshea FR, Gibson PR, Muir JG. Wheat bran affects the site of fermentation of resistant starch and luminal indexes related to colon cancer risk: a study in pigs. Gut 1999;45(6):840-7.