

# **INSULIN-LIKE GROWTH FACTOR RECEPTORS IN COLORECTAL CANCER**

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&

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## ***Abstract***

The IGF system is a crucial regulator of normal growth and development, however dysregulation of the system on multiple levels is associated with the incidence of a wide variety of malignancies including the breast, thyroid, lung, and colon, making the IGF system an important anti-cancer therapeutic target. Due to its role in mediating cellular proliferation, protection from apoptosis, and metastasis, traditional focus has been set on examining the role of the type 1 IGF receptor [IGF1R] in cancer. However there is mounting evidence to suggest the insulin receptor [IR] may also be involved in the potentiation and pathogenesis of cancers.

The observation that IGF-II is overexpressed, compared to normal tissues, by cancers suggests signaling via target receptors by this ligand has important implications on cancer pathogenesis. Indeed, both the IGF1R and IR have been demonstrated to be up-regulated in a variety of malignancies. In regards to IR isoform, the IGF-II binding IR-A is preferentially expressed by a number of cancer cell types. Together with the observation that an autocrine proliferative loop exists between IGF-II and the IR-A in malignant thyrocytes and cultured breast cancer cells, suggests signaling via the IR-A may play a role in cancer cell growth and survival. However, very few studies on the IR-A have been conducted in cells co-expressing the IGF1R. This is mainly due to the difficulties associated with discrimination between signaling arising from IGF1R homodimers, IR-A homodimers, and IGF1R/IR-A hybrid receptors.

It is not known how the IR-A interacts, and functions in conjunction with the other receptors of the IGF system to signal biologically relevant outcomes, especially in terms of anti-cancer therapeutics that aim to block and down-regulate the IGF1R. Current anti-cancer therapies targeting the IGF system have concentrated on blocking IGF signaling via the IGF1R, due mostly to the functional properties of the receptor, but also in part due to the metabolic consequences associated with blockade and inhibition of the IR. This individual targeting of the IGF1R potentially leaves a pathway by which IGF-II secreted by the tumour can circumvent current IGF1R based therapies. Consequently, this thesis investigated whether the IR-A could compensate for the targeted loss of the IGF1R and how the IR-A interacts with the IGF1R in cells co-expressing these two receptors. In addition, the individual ability of the IR isoforms to signal biological outcomes in response to IGF stimulation was assessed.

The main experimental techniques used throughout this body of work included; assessment of protein expression and activation by Western blot, siRNA mediated gene silencing, and measures of cell proliferation, survival, and migration.

The key areas of investigation included:

1. Investigation of the individual ability of the IR isoforms to signal biological outcomes in response to IGF stimulation
2. Identification of an appropriate cell line model in which to investigate the interactions between the IR-A and IGF1R
3. Optimisation of siRNA mediated knock-down of the IR-A and IGF1R in SW480 colorectal adenocarcinoma cells
4. Determination of the biological role of the IR-A in SW480 cells co-expressing the IGF1R

The key findings from this work included:

1. The IR-A could not compensate for IGF1R depletion in SW480 cells
2. Dual silencing of the IR-A and IGF1R indicated signaling via the IGF1R was dominant to signaling via the IR-A in SW480 cells
3. Signaling via IR-A/IGF1R hybrid receptors may not be as potent as signaling via IGF1R homodimers
4. IGF-I at physiological concentrations can stimulate biological responses via both isoforms of the IR.

## ***Declaration***

This thesis contains no material that has been accepted for the award of any other degree or diploma in any University. To the best of my knowledge and belief, it contains no material that has previously been published by any other person except where due reference is made.

I give consent for my thesis, when deposited in the library of the University of Adelaide to be available for loan and photocopying

Gemma Victoria Brierley

## **Acknowledgements**

Here we are, at the end of it all. A year ago I thought I would be struggling to write a very different thesis. What now forms this thesis makes for a pleasant and welcome surprise. I'd like to start by acknowledging and thanking Prof. John Wallace [Department of Biochemistry, University of Adelaide], Dr. Leah Cosgrove [CSIRO, Molecular and Health Technologies, Adelaide], and Dr. Lance Macaulay [CSIRO, Molecular and Health Technologies, Melbourne] for their supervision. Particular thanks to Lance for his thoughtful and detailed feedback throughout the preparation of this document. I also acknowledge the input of Dr. Briony Forbes at the beginning of this project.

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Throughout the last four years, the encouragement and support I have received from my family and friends has been unrelenting. "Thank you", doesn't quite seem adequate enough. One question I have been asked a lot over the last few weeks is "if you could go back, would you do it all again?" The answer is "yes - but I would approach it differently". My reasons for beginning a PhD in the first place have not changed, and through circumstance of location I have been extremely fortunate to forge strong friendships with several fun-loving, smart, inspirational, like-minded people. To them I am especially thankful and grateful. For the opportunity to have crossed paths with them, I would do it all again.

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He who learns must suffer,  
Even in our sleep, pain which cannot forget,  
Falls drop by drop upon the heart,  
Until, in our own despair,  
Against our will,  
Comes wisdom

- Aeschylus

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## **List of Publications**

*Peer reviewed publications [see appendix]*

\* Denotes joint first authorship

Denley A\*, Carroll JM\*, **Brierley GV**, Cosgrove LJ, Wallace JC, Forbes BE, Roberts CT Jr. Differential activation of insulin receptor substrates 1 and 2 by insulin-like growth factor-activated insulin receptors. *Molecular Cell Biology* 2007 May; 27[10]: 3569 - 77

Denley A, **Brierley GV**, Carroll JM, Lindenberg A, Booker GW, Cosgrove LJ, Wallace JC, Forbes BE, Roberts CT Jr. Differential activation of insulin receptor isoforms by insulin-like growth factors is determined by the C domain. *Endocrinology* 2006 Feb; 147[2]: 1029 – 1036

### *Conference Proceedings*

**Brierley GV**, Forbes BE, Macaulay L, Siddle K, Wallace JC, Cosgrove LJ. The biological role of insulin-like growth factors signalling via IR-A and IR-A/IGF1R hybrid receptors in cell survival, migration, and invasion. *The role of the IGF system in Cancer – Taormina, Sicily, Italy. November 2005*

**Brierley GV**, Forbes BE, Wallace JC, Cosgrove LJ. The biological role of insulin-like growth factors signalling via IR-A and IR-A/IGF1R hybrid receptors in cell survival, migration, and invasion. *Gordon Research Conference: Insulin-like growth factors in physiology and disease – Ventura, California, U.S.A. April 2005*

## Abbreviations

|                          |  |
|--------------------------|--|
| °C                       | degrees celcius  |
| µg                       | microgram  |
| µl                       | microlitre   |
| µm                       | micrometre   |
| 3'                       | three prime  |
| 5'                       | five prime   |
| 95% CI                   | ninety five percent confidence interval                      |
| bp                       | base pair  |
| Ab                       | antibody   |
| ACF                      | aberrant crypt foci  |
| APC                      | Adenomatosis Polyposis coli                                  |
| ASO                      | anti-sense oligonucleotide                                   |
| ATP                      | adenosine triphosphate                                       |
| BMI                      | body mass index  |
| BuA                      | butyrate   |
| BRET                     | bioluminescence resonance energy transfer                    |
| BSA                      | bovine serum albumin   |
| CO <sub>2</sub>          | carbon dioxide   |
| cDNA                     | complementary DNA  |
| c.f                      | compared with  |
| COX-2                    | cyclooxygenase-2   |
| CR                       | cystine-rich   |
| CRC                      | colorectal cancer  |
| CT                       | carboxy-terminal tail  |
| C-terminus               | Carboxyl-terminus  |
| dATP                     | 2'-Deoxyadenosine 5'-triphosphate                            |
| DEPC                     | diethyl pyrocarbonate  |
| DEPC-MQ H <sub>2</sub> O | dethyl pyrocarbonate milliQ water                            |
| DMEM                     | Dulbecco's modified eagles medium                            |
| DMSO                     | dimethyl sulfoxide   |
| DNA                      | deoxyribonucleic acid  |
| dL                       | decilitre  |
| dATP                     | 2'-deoxyadenosine 5'-triphosphate                            |
| dCTP                     | 2'-deoxycytidine 5'-triphosphate                             |
| dGTP                     | 2'-deoxyguanosine 5'-triphosphate                            |
| dTTP                     | 2'-deoxythymidine 5'-triphosphate                            |
| dNTPs                    | deoxynucleotide triphosphates                                |
| DTT                      | dithiothreitol   |
| EC <sub>50</sub>         | 50% effective concentration                                  |
| EDTA                     | ethylene diamine tetra-acetic acid                           |
| EGF                      | Epidermal Growth Factor                                      |
| EGFR                     | Epidermal Growth Factor Receptor                             |
| EPIC                     | European Prospective Investigation into Cancer and Nutrition |
| Erk-1/2                  | extracellular signalling-related kinase – 1/2                |
| Eu                       | Europium   |
| FACS                     | fluorescent-activated cell sorting                           |
| FCS                      | foetal calf serum  |
| FITC                     | fluorescein isothiocyanate                                   |
| F <sub>n</sub>           | fibronectin domain   |
| GH                       | growth hormone   |
| Grb2                     | Growth factor receptor-bound protein 2                       |
| H <sub>2</sub> O         | water  |
| HEPES                    | 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid           |
| HER-2                    | Human Epidermal Growth Factor-2                              |
| Hybrid-A                 | IGF1R/IR-A hybrid receptor                                   |
| Hybrid-B                 | IGF1R/IR-B hybrid receptor                                   |
| IB                       | immunoblot   |
| IC <sub>50</sub>         | concentration of inhibitor which reduces binding by 50%      |
| IGF1R                    | Type 1 IGF receptor  |
| IGF1R <sub>α</sub>       | Type 1 IGF receptor alpha-subunit                            |
| IGF1R <sub>β</sub>       | Type 1 IGF receptor beta-subunit                             |
| IGF2R                    | Type 2 IGF receptor  |
| IGFBPs                   | Insulin-like growth factor binding proteins                  |
| IGF-I                    | Insulin-like growth factor I                                 |
| IGF-I CII                | IGF-I containing the IGF-II C domain                         |
| IGF-I DII                | IGF-I containing the IGF-II D domain                         |
| IGF-I CIIDII             | IGF-I containing the IGF-II C and D domains                  |
| IGF-II                   | Insulin-like growth factor II                                |
| IGF-II CI                | IGF-II containing the IGF-I C domain                         |
| IGF-II DI                | IGF-II containing the IGF-I D domain                         |
| IGF-II CIDI              | IGF-II containing the IGF-I C and D domains                  |
| Ins                      | insert domain  |
| IP                       | immunoprecipitate  |
| IR                       | Insulin receptor   |

|   |  |
|---|--|
| IR <sub>α</sub>                               | Insulin receptor alpha-subunit   |
| IR <sub>β</sub>                               | Insulin receptor beta-subunit  |
| IR-A  | A isoform of the human insulin receptor [exon 11-]                                 |
| IR-B  | B isoform of the human insulin receptor [exon 11+]                                 |
| IRS-1   | Insulin Receptor Substrate -1  |
| IRS-2   | Insulin Receptor Substrate -2  |
| JM  | juxtamembrane domain   |
| kDa   | kilodalton   |
| L1  | large domain 1   |
| L2  | large domain 2   |
| L-chain                                       | light chain  |
| LHS   | left-hand side   |
| LOI   | loss of imprinting   |
| M   | molar  |
| mAb   | monoclonal antibody  |
| MALDI-TOF                                     | matrix-assisted laser desorption/ionisation – time of flight                       |
| MAPK  | Mitogen-activated protein kinase   |
| Max.  | maximum  |
| mg  | milligram  |
| MgCl <sub>2</sub>                             | magnesium chloride   |
| ml  | millilitre   |
| mM  | millimolar   |
| MMP   | matrix metalloproteinase   |
| MOPS  | 3-(N-morpholino)propanesulfonic acid   |
| mRNA  | messenger ribonucleic acid   |
| N/A   | not applicable   |
| NaCl  | sodium chloride  |
| NaF   | sodium fluoride  |
| Na <sub>4</sub> O <sub>7</sub> P <sub>2</sub> | sodium orthovanadate   |
| Na <sub>3</sub> PO <sub>4</sub>               | sodium pyrophosphate   |
| ng  | nanogram   |
| NH <sub>4</sub>                               | ammonia  |
| nm  | nanometer  |
| nM  | nanomolar  |
| n/s   | not significant  |
| NSAIDs  | non-steroidal anti-inflammatory drugs  |
| nt  | nucleotide   |
| N-terminus                                    | amino-terminus   |
| OD <sub>260</sub>                             | optical density at 260 nm  |
| OD <sub>280</sub>                             | optical density at 280 nm  |
| p53   | protein 53   |
| pAb   | polyclonal antibody  |
| PAGE  | polyacrylamide gel electrophoresis   |
| PBS   | phosphate buffered saline  |
| PCR   | polymerase chain reaction  |
| PFA   | paraformaldehyde   |
| PI3K  | phosphoinositide kinase-3  |
| pIRS-1  | phosphorylated IRS-1   |
| pIRS-2  | phosphorylated IRS-2   |
| PKB   | protein kinase B   |
| PMSF  | phenylmethanesulphonyl fluoride  |
| PTEN  | phosphatase and tensin homologue   |
| R <sup>-</sup>                                | IGF1R negative NIH 3T3-like mouse fibroblasts                                      |
| R <sup>+</sup> IGF1R                          | IGF1R negative NIH 3T3-like mouse fibroblasts recombinantly expressing human IGF1R |
| R <sup>+</sup> IRA                            | IGF1R negative NIH 3T3-like mouse fibroblasts recombinantly expressing human IR-A  |
| R <sup>+</sup> IRB                            | IGF1R negative NIH 3T3-like mouse fibroblasts recombinantly expressing human IR-B  |
| RHS   | right-hand side  |
| RISC  | RNA-inducing silencing complex   |
| RNA   | ribonucleic acid   |
| RNAi  | RNA interference   |
| rpm   | revolutions per minute   |
| RR  | relative risk  |
| RT-PCR  | reverse-transcriptase PCR  |
| SCFA  | short-chain fatty acid   |
| SDS   | sodium dodecyl sulphate  |
| SH2   | Src-homology domain  |
| siRNA   | short interfering RNA  |
| SEM   | standard error of the mean   |
| TAE   | Tris-acetate EDTA  |
| TK  | tyrosine-kinase domain   |
| Tris-Cl                                       | Tris (hydroxymethyl) aminomethane  |
| Tyr   | tyrosine   |
| VEGF  | vascular epidermal growth factor   |
| vs.   | versus   |
| WB  | Western blot   |
| WCL   | Whole cell lysate  |