

09 PH
F 3322



Studies on HIV-1 Virion Infectivity Factor

Feng Feng

M.Sc.

Infectious Diseases Laboratories

Institute of Medical and Veterinary Science

Adelaide

South Australia

School of Molecular & Biomedical Science

University of Adelaide

Adelaide

South Australia

*A thesis submitted to the University of Adelaide in fulfilment of the
requirements for the degree of Doctor of Philosophy*

October, 2004

Contents

Abstract.....	vi
Acknowledgments.....	viii
Declaration of Originality.....	ix
Abbreviations.....	x
Publications Related to this Study.....	xii
Chapter 1. Introduction.....	1
1.1 Historical Background of HIV/AIDS.....	1
1.2 Overview of Human Immunodeficiency Virus.....	4
1.2.1 Classification of HIV	4
1.2.2 HIV virion structure.....	5
1.2.3 Genetic organisation of HIV.....	5
1.2.4 HIV replication Cycle.....	6
1.2.4.1 Viral attachment and viral fusion	7
1.2.4.2 Reverse transcription.....	8
1.2.4.3 Viral DNA integration.....	9
1.2.4.4 Transcription and translation.....	10
1.2.4.5 Assembly of virus and viral budding.....	11
1.2.5 HIV gene expression.....	12
1.2.5.1 HIV proviral genome transcription - overview	12
1.2.5.2 Regulation of transcription.....	13
1.2.6 HIV encoded proteins and their functions.....	15
1.2.6.1 The major structure proteins.....	15
1.2.6.2 Regulatory proteins/accessary proteins.....	18
1.2.7 HIV-1 Virion infectivity factor (Vif).....	24
1.2.7.1 HIV-1 Vif localisation and biological activity.....	24
1.2.7.2 Vif is required for efficient reverse transcription.....	25
1.2.7.3 Vif interacts with viral proteins and RNA.....	27
1.2.7.4 The effect of Vif is cell type dependent and involves cellular inhibitors of HIV replication.....	29
1.2.7.5 Host cellular factors that interact with Vif	30
1.2.8 Pathogenesis of HIV-1 Infection.....	34
1.2.8.1 Signs, symptoms of primary HIV-1 infection.....	34
1.2.8.2 Aspects of virus-host interactions affecting pathogenesis.....	35

1.2.8.3 Developments in viral pathogenicity.....	37
1.3 HIV/AIDS vaccine developments.....	39
1.4 HIV/AIDS treatment.....	40
1.5 Summary.....	43
1.6 Hypotheses.....	44
1.7 Specific Aims.....	45
Chapter 2. Materials and methods.....	46
2.1 Materials	46
2.1.1 Cells and cell culture.....	46
2.1.2 Bacterial culture.....	47
2.1.3 Yeast AH109 and cell culture.....	48
2.1.4 Plasmid vectors.....	48
2.1.5 Oligonucleotide sequences.....	49
2.1.6 Antibodies.....	51
2.1.7 Commonly used buffers and solutions.....	51
2.2 Methods.....	54
2.2.1 Investigation of Vif/Triad 3au or NVBP interactions in yeast.....	54
2.2.2 Production of Triad 3au DNA probe.....	55
2.2.3 RNA extraction and Northern blot analysis.....	56
2.2.4 Polymerase chain reaction (PCR).....	57
2.2.5 Determination of the full-length Triad 3au cDNA sequence	58
2.2.6 DNA constructions.....	59
2.2.7 Transient cell transfection.....	60
2.2.8 Western blotting.....	61
2.2.9 Co-immunoprecipitation analyses.....	62
2.2.10 Immunofluorescent staining	63
2.2.11 GST fusion protein pull-down assay.....	64
2.2.12 Production of HIV NL4.3 stocks.....	66
2.2.13 Single-cycle viral infectivity assay.....	66
2.2.14 Viral titres (TCID ₅₀).....	67
2.2.15 Extent of reverse transcription in newly infected target cells.....	68
2.2.16 Site-directed mutagenesis of HIV-1 Vif.....	69
Chapter 3. Interaction between HIV-Vif and Triad 3au.....	71
3.1 Introduction.....	71

3.1.1 NVBP.....	72
3.1.2 Triad 3au.....	72
3.2 Interaction in yeast between HIV-1 Vif and Triad 3au or NVBP	75
3.3 Interaction between Triad 3au and HIV Vif in mammalian cells.....	77
3.4 Effect of Triad 3au on the cellular localisation of Vif....	79
3.5 Triad 3au mRNA is expressed in both permissive and primary non-permissive cells.....	81
3.6 Identification of ZIN and preparation of the ZIN cDNA.....	83
Chapter 4. Interaction between HIV-1 Vif and ZIN, and Implications for Viral Replication.....	87
4.1 Introduction.....	87
4.2 ZIN interacts with purified Vif <i>in vitro</i>	88
4.3 ZIN interacts with HIV-1 Vif in mammalian cells.....	90
4.4 Co-localisation of Vif and ZIN in transfected cells....	91
4.5 Effect of ZIN overexpression on production and infectivity of HIV-1.....	94
4.5.1 TCID ₅₀ analysis of wild-type HIV produced in the presence or absence of co-transfected ZIN.....	94
4.5.2 Single-cycle viral infectivity assay of wild type and Δvif HIV in the presence or absence of co-transfected ZIN.....	95
4.5.3 Real time PCR analysis of viral DNA synthesis after infection.....	97
4.5.4 Discussion.....	98
Chapter 5. Vif Site-Directed Mutagenesis.....	100
5.1 Introduction	100
5.2 Experimental principle.....	101

5.3 Result and discussion.....	102
Chapter 6. General Discussion and Future Work.....	104
6.1 Discussion.....	104
6.2 Future work.....	114
Reference.....	118

Abstract

Virion Infectivity Factor (Vif) protein of human immunodeficiency virus type 1 (HIV-1) is essential for the productive viral infection of primary human CD4 T lymphocytes and macrophages. Recently, it has been reported that Vif overcomes the HIV-inhibitory effects of the cellular factor CEM15/APOBEC3G, which has cytidine deaminase activity. Using the yeast two-hybrid system, our laboratory previously reported the identification of a Vif-interacting ring finger protein called Triad 3 (renamed Triad 3au in this study), from a human leukocyte cDNA library. The full-length cellular protein homologue of Triad 3 has been recently identified as the zinc finger protein inhibiting NF κ B (ZIN). In this study, biological characteristics of Triad 3au/ZIN were investigated. Sequence analysis indicated that Triad 3au protein contains all 4 major ring-like motifs of ZIN. RNA was extracted from A3.01 cells and RT PCR was then performed using the ZIN gene specific primers. A single product of 1,464 bps was obtained and subsequently confirmed as the full-length coding region of ZIN by sequence analysis. GST fusion protein pull-down experiments confirmed that overexpressed ZIN binds to purified Vif *in vitro* suggesting a direct interaction between ZIN and Vif. Next, in studies of co-transfected human 293T cells, Triad 3au/ZIN and Vif were shown to interact using co-immunoprecipitation and confocal microscopy demonstrated co-localisation of Triad 3au and Vif in cytoplasm and membrane while co-localisation of ZIN and Vif in nuclei.

To test the biological relevance of the Vif-ZIN interaction, infectious HIV-1 NL4.3 virus stocks were produced in 293T cells expressing Flag tagged ZIN or Flag tag

alone from transfected plasmids. The virus stocks produced in the presence of exogenously expressed ZIN were less infectious in both single-cycle infectivity assay and end-point titration compared with virus produced in the absence of exogenous ZIN. Real Time PCR analyses showed that cells infected with HIV NL4.3 virus stocks produced in the presence of exogenously expressed ZIN, showed reduced levels of early viral DNA synthesis. This reduction in viral reverse transcription and the reduction in single-cycle viral infectivity were shown to be dependent on the presence of Vif in the virus producer cells. The possible mechanisms by which presence of ZIN in producer cells reduces HIV-1 reverse transcription and replication in target cells are discussed.