



LATE EOCENE MOLLUSCA AND RELATED COMPOSITE SPECIES
FROM
SOUTHERN AUSTRALIA

by

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APPENDICES

PUBLISHED PAPERS

BIBLIOGRAPHIC REFERENCES

PLATES

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A P P E N D I X A

LOCALITIES



All localities mentioned in this study are shown in Figs. 1-5 and in Table XIII.

The main areas dealt with in this study are the Adelaide Plains, the Noarlunga, and the Willunga SubBasins situated on the Eastern side of the St. Vincent Basin. All three of them represent classical localities in the Cainozoic geology and paleontology of Australia.

In particular, the Willunga SubBasin, with the type-section of the Aldingan stage outcropping in Maslin and Aldinga Bays, constitutes the fulcrum around which this thesis develops. The stratigraphic distribution of Tate's species and the comparison of the matrix still attached to their types revealed that Tate's classical locality 'Aldinga', the type of the 'Lower Aldinga Series' (Tate, 1879), is composite. It includes the Middle-Late Eocene deposits of Maslin Bay and of the northern tract of Aldinga Bay, and the Oligocene-Middle Miocene outcrops of the southern Aldinga Bay, from Port Willunga to Sellick Hills.

The Adelaide Plains material is derived entirely from cores; the Willunga material is almost entirely from outcrops, with only a few specimens from the subsurface; the Noarlunga material is represented by few specimens from outcrops, kept in the S.A. Museum Collections.

In order to obtain a better definition of the species and to attempt some interbasin correlation, some samples from two bores in the Murray Basin were also examined initially. To these two bores another four were added during an ongoing project on the Late Eocene Mollusca from the Padthaway Ridge for the S.A. Department of Mines and Energy. This project leads ^{to} a revision of the Late Eocene stratigraphic units in the Murray Basin and their relationships, and, therefore, the results will be presented elsewhere. (Buonaiuto, in prep.; M.F. Buonaiuto & W.K. Harris, in prep.)

Finally, part of the material concerning Tertiary composite species with Late Eocene representatives is from younger classical localities

TABLE XIII

BASINS	SUB-BASINS, EMBAYMENTS	LOCALITIES	KEY	STRATIGRAPHIC UNITS	AGE	PURPOSE OF STUDY	REFERENCES	
S A I L I N T V I L L I N G E N T	A D E L A I D E	Adelaide Children's Hospital Bores 5, 1, 2, Hd Yatala, Town Acre 717, corner Kermode St.-Sir Edwin Smith Ave.		Blanche Point Formation	Late Eocene (late P15 middle P16)	Palaeontological	Lindsay, 1969; Buonaluto, 1975	
		Adelaide (Kent Town) Bore, E & WS Bore 5, N.E. Parklands (1881-82)		as above	as above	as above	Tate, 1882; Lindsay, 1969; Ludbrook, 1973.	
	P L A I N S	Adelaide Metropolitan Subway, Bore CH 3 (DDH), North Bank of Torrens Lake, opposite Kintore Ave.		as above	as above	as above	Buonaluto, 1975; Lindsay, in prep.	
		West End Brewery, western Hindley St., Bore CH 2.		as above	as above	as above	this study; Lindsay, in prep.	
	S U B.	South Parklands, Bore CH 1A		as above	as above	as above	this study; Lindsay, in prep.	
		Longyear Bore 50, Hd Noarlunga, Sect. 82		'North Maslin Sands' 'South Maslin Sands' 'Tortachilla Limestone' Blanche Point Formation Chinaman Gully Formation Port Willunga Formation (Aldinga Member)	e-ly M.Eoc. late M.Eoc. e-ly L.Eoc. mid L. Eoc. mid/L.Eoc. Latest Eoc.	Stratigraphy	this study.	
	N O R T H A R S I L A N D S	Wilton Bluff, at Christie Beach; Port Noarlunga Jetty; Onkaparinga River mouth; River Road; Onkaparinga River, at the Fleming Bridge and opposite the Oval; intersection Honeypot Rd-South Road; northern slope of Ochre Point, at Moana Beach.		'North Maslin Sands' 'South Maslin Sands' Tortachilla Limestone Blanche Point Formation Chinaman Gully Formation Aldinga Member and Ruwaring Member* (Port Willunga Formation)	as above *Oligocene	Palaeontology and Stratigraphy	This study; Lindsay, 1970; Stuart, 1969; Glaessner & Wade, 1958; Ludbrook, 1969; Crespin, 1954.	
		Maslin Bay (type section)		'North Maslin Sands' 'South Maslin Sands' Tortachilla Limestone Blanche Point Formation	as above	Palaeontology and Stratigraphy	Reynolds, 1953; Crespin, 1954; Glaessner & Wade, 1958; Ludbrook, 1969; Lindsay, 1967; McGowan et al., 1970; Buonaluto, 1977a; Cooper, 1977a, 1978; Stuart, 1969.	
		Aldinga Bay (type section)		Blanche Point Formation Chinaman Gully Formation Aldinga and Ruwaring Members (Port Willunga Formation)				
	W I L L I N G A	Willunga Bore WLG 42, Hd Willunga, sect. 174, on the boundary with sect. 175.		'South Maslin Sands' Tortachilla Limestone Blanche Point Formation	as above	Stratigraphy	Cooper, 1977a, 1978.	
	Willunga Bore WLG 40, Sect. 412.		Gull Rock Member (Blanche Point Formation) Aldinga Member (Port Willunga Formation)	as above	Palaeontology	Cooper, 1978; Buonaluto, in prep.		
	Willunga Bore WLG 37, Sect. 477.		Gull Rock Member (Blanche Point Formation)	as above	Palaeontology	Cooper, 1978.		
	Chaffey's Road, Just South of Coriole and Seaview Wineries, McLaren Vale		Tortachilla Limestone 'Transitional Marl' Member (Blanche Point Formation)	as above	Stratigraphy	Cooper, 1978; this study.		
	Y O R K I N E	Stansbury		Port Vincent Limestone	Oligo-Miocene	Palaeontology	Crawford, 1965; Stuart, 1970.	
	Point Turton			Point Turton Limestone	Late Oligocene	Palaeontology	Ludbrook, 1967; Crawford, 1965; Lindsay (herein).	
M U R R A Y	P A D T H A W A Y R I D G E	B.Q. Butler Bore 4, Co Buccleuch, Hd Kirkpatrick, Sect. 8.		Buccleuch Beds	Latest Eocene	Stratigraphy and Palaeontology	Ludbrook, 1969; Buonaluto, (herein; in prep.)	
		Kiki Town Bore, Co Buccleuch, Hd Livingstone, sect. 51		Upper Knight Group Buccleuch Beds	mid L.Eoc. Latest Eoc.	as above	Ludbrook, 1969; Buonaluto, (herein; in prep.)	
		EWS Coonalpyn Bore 2, Co Buccleuch, Hd Coneybeer, Sect. 56 (type section)		Upper Knight Group Buccleuch Beds	as above	as above	Ludbrook, 1961; Buonaluto, (herein; in prep.)	
		PBD Tintinara Area School Bore 12, Township Allotment, Hd Coombe, Co Cardwell		Buccleuch 'B' Beds	as above	as above	Buonaluto (herein; in prep.)	
		Old S.A. Railways Tintinara Bore, as above		Buccleuch Beds	as above	as above	Clarke, 1896; Tate, 1898; Buonaluto (in prep.)	
	C E N T R A L	Waikerie Observation Bore 2, Hd Waikerie, Sect 692		Buccleuch Beds	as above	as above	Lindsay & Bonnett, 1973;	
		'River Murray Cliffs', Morgan Hd Cadell, Sect. C (type section)		Cadell Marl Lens (Morgan Limestone)	Middle Miocene	Palaeontology	Tate, 1885; Ludbrook, 1961, 1973.	
E U C L A		Wilton Bluff (type section); Weebubbe Cave; Abrakurrie Cave; Toolinna Cove; Hag Cave		Wilton Bluff Limestone	mid M.Eoc.	Palaeontology and Stratigraphy	Lowry, 1970; Ludbrook, 1973; McGowan, 1978a,b,c; Quilty, 1974, 1977.	
		Toolinna Cove		Toolinna Limestone	Late Eoc.	Palaeontology	as above	
O T W A Y	GAMBIER EMB.	Pritchard's Quarry, 12.9 km N.W. of Mount Gambier (S.A.)		Gambier Limestone	Late Oligocene	Palaeontology	Ludbrook, 1971; McGowan, et al., 1971; Abele et al., 1976.	
		Browns Creek, at Johanna River mouth (Aire District, Vict.) (type section)		Johanna River Sand Drowns Creek Formation	Mid.Eoc. Late Eoc.	Stratigraphy and Palaeontology	Ludbrook, 1973; Abele et al., 1976.	
		Castle Cove, Aire District, (type section)		Castle Cove Limestone	Latest Eoc. E-ly Olig.	Stratigraphy	as above	
		'Gellibrand River'		Gellibrand Marls	E-ly Mloc.	Palaeontology	as above	
		Maude		Lower Maude Limestone	Late Olig.- E-ly Mloc.	Palaeontology	as above; Abele & Page, 1974.	
		T A R R A	Muddy Creek, near Hamilton (Vict.)		Muddy Creek Marls	Mid, Mloc.	Palaeontology	Abele et al., 1976; Ludbrook, 1973
		Grange Burn, near Hamilton		Grange Burn Coquina	Latest Mloc. -E. Plioc.	Palaeontology	as above	
		TORQUAY EMB.	Bird Rock, 'Spring Creek'		Jan Juc Formation Puebla Formation	Late Olig. E-ly Mloc.	Palaeontology Palaeontology	as above as above
		PORT PHILLIP EMB.	Balcombe Bay		Balcombe Clay (Fyansford Formation)	Middle Miocene	Palaeontology	as above
	B A S I N S	TABLE CAPE	'Fossil Bluff', Freestone Cove, Table Cape		Freestone Cove Sandstone	Early Miocene	Palaeontology	Ludbrook, 1967; 1973; Quilty, 1966, 1972, 1974.

distributed along the eastern southern Australian margin. These localities, mentioned in the systematic part, are indicated in Fig. 1 and Table XIII. Stratigraphic units and age of these localities are included in the descriptions of the species. For further information the reader is referred to Ludbrook (1973), who offers an updated and thorough discussion.

The descriptions of the Cainozoic St. Vincent and Murray Basins are purposefully not outlined, and limited to figures because both are well known and overdescribed by ^{previous} authors. The intracratonic St. Vincent Basin has been dealt with by Glaessner (1953), Glaessner & Wade (1958), Ludbrook (1969), Stuart (1969, 1970), Wopfner (1972), Daily, Firman, Forbes & Lindsay (1976), etc. The pericratonic Murray Basin has been described by Ludbrook (1957, 1958, 1961, 1969), O'Driscoll (1960), Wopfner (1972), Abele et al. (1976).

A P P E N D I X B

DESCRIPTION OF THE MEASURED DETAILED SECTIONS

Fourteen detailed sections have been measured in total. Nine are located in the Willunga SubBasin (Figs. 3c, 10, 12) at Chaffeys Road, McLaren Vale (Section A); at Maslin Bay (Section B, C, D, V, W, X); at Blanche Point (Section Y); at Chinaman Gully, Aldinga Bay (Section Z). Four are situated in the Noarlunga SubBasin (Figs. 3b, 10); at Witton Bluff, Christies Beach (Section E, F); along the River Road, Onkaparinga River (Section G, H); on the river bank, opposite the Noarlunga oval (Section I). The fourteenth section was measured on the continuous core of the Longyear Bore 50, Hd Noarlunga, Adelaide Plains SubBasin (Fig. 3a). Its stratigraphic column and lithological description are shown in Fig. 11.

WILLUNGA SUB-BASIN

SECTION A

Chaffey's Road, McLaren Vale, road cut on the knoll, just south of Coriole and Seaview Wineries. Total thickness 160 cm. Dip sub-horizontal, N-S. From bottom to top.

Tortachilla Limestone :

- Breccia; matrix of mottled yellow and green glauconitic sands; intraformational clasts highly limonitic with moulds of gastropods and bivalves. 30 cm thick.
- Yellowish sands with green glauconitic intercalations. 10 cm thick.
- Yellowish sands with vertical bodies of glauconite (? dissolved karsted horizon). 40 cm thick. The glauconitic bodies (pit filling?) are referable to the overlying,

'Transitional Marls' :

- Intraformational breccia: sandy glauconitic matrix; very angular limonitic clasts (\emptyset -5-10 cm).

SECTION B

'Uncle Tom's Cabin'; total thickness about 3.25 m; dip 4° S 65° W; from bottom to top.

'South Maslin Sands' :

- purplish, unconsolidated, well sorted, quartzitic limonitic fine sands; fossil content: limonitized pellets, foraminifera moulds, fragments of shells; in erosional contact with:

Tortachilla Limestone :

- red-brownish, unconsolidated, quartzitic limonitic microconglomerate: poorly clayey sandy matrix prevailing on clasts; clasts of 1-10 mm diameter, spheroidal or well rounded the quartzitic, well rounded often elongated and/or flattened the limonitic; the latter are both from inorganic origin and from moulds of organic origin; fossil content: limonitized pellets, foraminifera molds, sharks' teeth; thickness about 70

cm; passing gradually to:

- microconglomeratic limestone: biomicrudite composed chiefly of Bryozoa with very frequent irregular vertical and horizontal patches of microconglomeratic sands, crossing and following the layering; the lower part is richer in bioclasts; nature of clasts: fragment of shell or limonitic the organic, quartzitic and limonitic the inorganic; fossil content: limonitized pellets worn benthonic foraminifera, Bivalvia and Gastropoda (generally as moulds), echinoids, brachiopods, barnacles, bryozoa, sharks' teeth, worms; thickness about 30-45 cm; unconformably underlying with erosional contact :
- yellowish brownish unconsolidated microconglomerate with finer intercalations: clasts as the lower; poorly clayey sandy matrix; fossil content: limonitized pellets, worn benthonic foraminifera, bivalves, gastropods, scaphopods, echinoids, brachiopods, barnacles, crabs' claws, sharks' teeth, bryozoa, worms; thickness about 40 cm; passing gradually to:
- greenish unconsolidated coarse microconglomeratic sands, regularly interbedded with thin pinkish bioclastic intercalations, mainly made from bryozoa; clasts finer than the lower ones; fossil content as the lower: thickness about 45 cm; passing gradually to:
- mottled greenish pinkish unconsolidated or spotty cemented microconglomeratic sands with some galls of the same lithology as the 'Transitional Marls'; fossil content, as the lower; thickness about 55 cm; passing gradually to:
- microconglomeratic limestone: biomicrudite, chiefly composed of bryozoa; richer in bioclasts than the lower limestone; hollowed out by deep irregular subvertical pits, filled by the greenish glauconitic sands from the overlying 'Transitional Marls'; fossil content as the lower without crabs' claws; thickness about 65 cm; in erosional karst contact with:

'Transitional Marls' (Blanche Point Formation)

- Greenish unconsolidated glauconitic sands; fossil content as the lower; thickness about 25 cm.

SECTION C.

Reynolds Cave, total thickness about 1.95 m; roughly same lithology, same dip; from bottom to top.

'South Maslin Sands'

- as in Section A, but displaying shrinkage structures in erosional contact with:

Tortachilla Limestone

- microconglomerate as in Section A; thickness about 20-30 cm; passing gradually to:
- microconglomeratic limestone: biomicrudite as the lower in section B with microconglomeratic glauconitic lenses; thickness 65 cm; (?) gradually passing to:
- greenish microconglomeratic glauconitic sands; thickness about 25 cm; gradually passing to:
- pitted limestone as the upper in Section A; thickness about 45 cm in erosional karst contact with:

'Transitional Marls'

- glauconitic sands more clayey than in Section A; thickness 30 cm.
- In this section the fossiliferous content appears similar as in Section A, but more scarce.

SECTION D.

15-20 m southward Section B, dip subhorizontal; total thickness measured 1.60 m; from bottom to top:

Tortachilla Limestone

- microconglomerate as the lower in Sections A and B; in contact with:
- barren microconglomeratic sands, interpreted as residue of the Tortachilla Limestone after dissolution of the calcareous content; thickness about 55 cm; passing gradually to:

- same microconglomerate but with very frequent lentils and patches of glauconitic sands; thickness 35 cm; passing gradually to:
- glauconitic clayey sands with relicts of calcareous microconglomerates; fossil content: bivalves, gastropods, brachiopods, bryozoa, worms, crabs, sharks' teeth, barnacles. Passing disconformably to:

'Transitional Marls'

- glauconitic clays and marls.

SECTION V

Southern Cliffs of Maslin Bay, first gully 200 m south of 'Uncle Tom's Cabin'. Dip as in Tortachilla Limestone at 'Uncle Tom's Cabin' (from bottom to top).

Tortachilla Limestone

Karsted surface.

'Transitional Marls'

- Green glauconitic, sandy, microconglomeratic limestone, highly fossiliferous, nodular in the upper part. 35 cm thick, unconformably overlain by:
- mottled yellowish greenish, sandy marl with bioclasts and few lithoclasts. fossils observed: Chlamys flindersi, Pycnodonte tatei, Spondylus tortachillensis, Turritella sp., gastropods, echinoids, brachiopods. 30 cm thick. Unconformably overlain by:
- Coquiña lens, cemented, with Bryozoa, C. flindersi, C. peroni, P. tatei, brachiopods, echinoids. 10 cm thick, gradually passing to:
- mottled greyish-green, glauconitic soft marls (Hantkenina horizon), very rich in glauconitic moulds of mollusca. 55 cm thick. Gradually passing to:
- hard glauconitic limestone with C. flindersi, P. tatei, Dimya sigillata, Turbo sp., brachiopods, echinoids, sharks' teeth, Spirocolpus aldingae, Bryozoa. 60 cm thick.
- Whitish-yellowish marl with bioclastic thin lenses and C. flindersi, C. peroni, S. aldingae, Brachiopoda, Bryozoa, Echinoidea, Lentipecten sp.

D. sigillata, Yermes. 40 cm thick.

- Hard greyish-buff marly limestone, silicified in its top part. 45 cm thick.
- Soft glauconitic mottled green-white marl, with high shell dissolution. 1.45 cm thick.
- Glauconitic marly limestone. 30 cm thick.

'Gull Rock Member'

- Whitish soft marls. 20 cm thick.
- Whitish marly limestone. 18 cm thick.
- Whitish marls with harder nodules (Phygraea tarda horizon). 40 cm thick.
- Grey soft marls. 15 cm thick.
- Hard limestone with some glauconite. 50 cm thick. Unconformably overlain by:

'Hallett Cove Sandstone' (Pliocene)

SECTION W

Maslin Bay, southern cliffs, 100 m about, north of Reynolds Cave.

Gull Rock Member

Top Phygraea tarda horizon.

- alternance of siliceous soft marls, and hard siliceous marls or limestone. 3.10 m thick.
- Observations. In the lowest hard band, were noted a mould of Pleurotomaria s.l., Bryozoa, Spirocolpus, Arca s.l. sp., a muricid, Bivalvia, Praehyalocylis annulata, Emarginula sp., Chlamys, moulds of siliquariids.

SECTION X

Maslin Bay, detritus conoid, just south of Reynolds Cave.

'Transitional Marls'

- Hard marly limestone, partly silicified.
- Dissolution front.

Gull Rock Member

- Whitish soft marls, with siliceous patches. 80 cm thick.

- Phygraea tarda horizon, whitish soft marls, 70 cm thick, two thin lenses of P. tarda, one at the bottom and one on the top of the unit.
- Soft glauconitic marls, with siliceous patches. 70 cm thick.
- Hard glauconitic marly limestone. 10 cm thick.
- Glauconitic marls with siliceous patches. 2.30 m thick.
- Glauconitic marls. 40 cm thick.
- Glauconitic limestone. 50 cm thick.
- Glauconitic marls. 30 cm thick.
- Glauconitic limestone. 10 cm thick.
- Alternance of glauconitic limestones and marls with Spirocolpus aldingae. 3.10 m thick.

'Soft Marls'

- greyish glauconitic marls. 1.50 m thick.
- detritus.

Hallett Cove Sandstone (Pliocene)

SECTION Y

Blanche Point, 1st and 2nd amphitheatre, southern slope of Blanche Point. Dip about 3⁰.

'Gull Rock Member'

- Phygraea tarda horizon, whitish marls. 40 cm thick.
- Whitish marls, bioturbated, with patches of glauconitic pellets and Bryozoa. .5 m thick.
- Siliceous limestone or marl. 20 cm thick.
- Whitish marls, with Anellida, Bryozoa, rare Chlamys and Turritella moulds. 40 cm thick.
- Siliceous limestone. 10 cm thick.
- Alternance of grey soft marls and hard siliceous limestone with siliceous nodules. 11.70 m thick. Spirocolpus assemblages associated with abundant sponge spicules 3.25 m above top P. tarda horizon.

Gradually passing to:

'Soft Marls'

- Soft marls very rich in siliceous sponges and Mollusca. 80 cm thick.
- hard horizon of siliceous nodules imbedded in marls similar to the ones above and below (20 cm thick).
- Soft grey to black marls, extremely rich in sponge spicules and Mollusca. 16.50 m thick. Nautilus sp. in patches, 8m above the last hard nodular horizon. Upper part, with shells subjected to intensive dissolution.

SECTION Z

Aldinga Bay, cliff just on the southern side of Chinaman Gully (from bottom to top).

'Soft Marls'

- Lower unit, in erosional contact with:

Chinaman Gully Formation

- Dark grey clay, stained by limonite, 50 cm. thick.
- Very fine alternance of grey fine clayey sands and red clays. 50 cm thick. In erosional contact with:
- Quartz angular sands (2-3 cm) with scarce matrix of finer sand. 15 cm thick.
- Fine whitish clayey quartz sands with thin lenses of red limonitic sands. 25 cm thick.
- Yellow clayey fine sands. 10 cm thick.
- Banded brick-red and grey silty clays. 20 cm thick.
- Grey silty clays. 10 cm thick. In erosional contact with:
- Banded brick-red/grey clays with coarse sand nodules. 10 cm thick. In erosional contact with:
- Grey silty clays with sandy nodules. 5 cm thick. In erosional contact with:
- Brown silty clays with very thin grey horizons. Small scale deformations by compression. 10 cm thick. In erosional contact with:
- Grey coarse clayey sands, stained by limonite. 30 cm thick. In

erosional contact with:

Aldinga Member, Port Willunga Formation

- Quartz microconglomerate with sandy matrix. 15 cm thick.
- Sandstone gradually passing to a crossbedded Bryozoal limestone. 1.10 m thick.

Observations. On the cliffs, just north of Chinaman Gully, the Bryozoal limestone is underlain by a thin horizon of mottled whitish-green glauconitic marls.

NOARLUNGA SUB-BASIN

SECTION E

At Christies Beach, at the middle of the northern side of Witton Bluff; total thickness about 1.75 m; dip $10^{\circ}S$ $50^{\circ}W$; from bottom to top.

South Maslin Sands

- red purplish unconsolidated quartzitic limonitic fine sands intercalated with frequent greenish clayey very fine sands marking cross-bedding; a minor fainter cross-bedding is displayed too; clasts: angular quartz, mica, limonite, organic calcite; fossil content: pellets, limonitic the smaller, calcite the larger, moulds of foraminifera, arenaceous foraminifera; fragments of Chlamys sp; in erosional contact with:

Tortachilla Limestone

- red brown unconsolidated very fine sands with very frequent intercalations of greenish veins like as the lower, greenish clayey galls, lentils which in the upper part are graded white greenish at the bottom and limonitic at the top. In the upper part an undulating limonitic ripple mark-like interbedding; clasts: round quartz, limonitic organic aggregates, organic calcite; fossil content: Bryozoa,

limonitized pellets, moulds of badly preserved benthonic foraminifera, traces of intensive bioturbation from burrowing organisms, with borings filled by finer greenish sands; thickness about 90-100; in erosional contact with:

- intraformational conglomerate with angular large galls of sand or of microconglomerate and abundant microconglomeratic lentils; the secondary calcitic cement gradually increases upward from the underlying horizon; clast as the lower; fossil content: limonitic pellets; thickness c. 10-15 cm in erosional contact with:
- green glauconitic fine sands with frequent microconglomeratic and sand galls and microconglomeratic lenses irregularly shaped; clasts: quartz, limonite from organic origin, organic calcite; fossil content: simple or lumped pellets, glauconitic the larger, limonitized the smaller; moulds of foraminifera, calcitic tubules from worms, moulds of molluscs; thickness c. 30 cm; passing gradually to:
- deeply pitted mottled brown greenish microconglomeratic limestone with abundant calcitic cement; clasts as the lower; fossil content: moulds of molluscs and burrowing traces filled of limonitic material; the pits are filled by the overlying glauconitic sands; thickness c. 30 cm.; unconformably underlying to:

'Transitional Marls'

- green glauconitic sands very fossiliferous, nearly a coquina; fossil content: bivalves, gastropods, echinoids, brachiopods, etc., all generally of very large dimensions; thickness c. 60 cm.

SECTION E

At the foot of Witton Bluff; total thickness estimated 1.50-1.60 m.; the lower contact is not observable; dip as in Section D; from bottom to top.

- Tortachilla Limestone
- Same red-brownish bioturbated sands as the lower in Section D; unconformably underlying to:
- intraformational conglomerate with arenaceous lentils in the lower part and angular sand galls occurring in all the bed but more frequent upward; thickness 30-40 cm; in erosional contact with:
- greenish glauconitic microconglomeratic limestone deeply pitted with sand- and microconglomerate-galls; pits filled by the overlying green sands; fossil content: bivalves, gastropods, echinoids etc. thickness c. 40 cm; unconformably underlying by karst surface to:

Transitional Marls

- green glauconitic sandy coquiña as in Section D; thickness 10-25 cm.

SECTIONS G, H

Along River Road, roadcut on the northern bank of the Onkaparinga River. Total thickness 4.10 m. Dip subhorizontal; crossbedding dipping westward.

? 'North Maslin Sands'

- Red yellowish clayey sands, medium size; dispersed quartz pebbles (15 mm) and small clay galls. 60 cm thick.
- Sands with more clayey matrix; regular thin clayey intercalations. 25 cm thick. Brusquely passing laterally to and overlain by,
- Coarser angular purple sands with abundant clay galls. 40 cm thick (max.). Unconformably overlain by:

? 'South Maslin Sands' - Tortachilla Limestone undifferentiated.

- Thin ferruginous silty clays, varva-like, gradually passing to alunite westward. 10 cm thick.
- Coarse angular purple sands with dispersed clay galls (ϕ 2-3 cm); alunite galls on the upper part; large cross-bedded horizons of alunite, gradually passing to sands and clay galls. 90 cm thick.

- Ferruginous clayey sands 30 cm thick.
- Yellow clayey sands with clay galls. 10 cm thick.

'Tortachilla Limestone'

- Yellowish cemented, highly microconglomeratic sands with rare intraformational clasts; cross-bedding in places. 40 cm thick.
Unconformably overlain by:
- Red conglomeratic sands, unconformably passing to:

'Transitional Marls'

Green glauconitic sands, very rich in fossil moulds: Turritella, Chlamys, and other Bivalvia, large rounded relicts (\emptyset 20-30 cm) from the underlain units. Section G, 10 m westward of Section H, is represented by the lowermost unit of Section H and by the above described 'Transitional Marl' unit, containing the relicts from 'Tortachilla Limestone'.

SECTION I

At Noarlunga Oval; total thickness c. 5.85 m; dip $5^{\circ}\text{S } 80^{\circ}\text{W}$;
from bottom to top:

South Maslin Sands

- sandy clays, brownish in the lower and grey in the upper with 5 cm; brick-red clay at the top; rich in limonite, angular or nearly rounded quartz; and very thin lentils of concretionary calcite; fossil content: worm tubules, gastropods, fragments of Chlamys sp; thickness only 70 cm. outcropping; in erosional contact with:
- red-brownish cross-bedded sands with hard limonitic clayey intercalations at the top of each cross-bed; the intercalations often contain veins of concretionary calcite; clasts: well rounded quartz, limonite also from organic moulds, mica, organic and concretionary calcite; fossil content; limonitic moulds of foraminifera, calcitic and limonitic pellets, calcitic and limonitic moulds of

gastropods, fragments of Chlamys sp, echinoids' radioli; thickness c. 3.60 m gradually passing to:

- cross-bedded microconglomeratic sands with abundant clay-galls and bioturbation traces; clasts as the lower; fossil content: limonitized pellets, poorly preserved benthonic foraminifera in moulds or with their shells; thickness c. 95 cm gradually passing to:
- regularly layered compacted rather coarse sands, of the usual nature; fossil content as the lower; thickness 10-15 cm, unconformably underlying to:
- intraformational conglomerate with angular clay- and sand- and microconglomerate-galls imbedded in abundant sandy matrix poorly consolidated in the lower part; thickness 25-50 cm; unconformably underlying to:
- green unconsolidated glauconitic sands; clasts as the lower; fossil content: limonitic and glauconitic pellets, moulds of gastropods, fragments of molluscs, crinoids, bryozoa, molds and fragments of benthonic foraminifera; thickness c. 15 cm; gradually passing to:
- microconglomeratic limestone (biomicrudite) deeply pitted as usual; fossil content: bryozoa, molluscs, brachiopods, echinoids; thickness c. 65 cm; in erosional karst contact with:

'Transitional Marls'

- green glauconitic fossiliferous sands; thickness c. 20-30 cm.

A P P E N D I X C

SAMPLE CHECK-LISTS

TORTACHILLA LIMESTONE

(Section B, 'Uncle Tom's Cabin')

TL2

	<u>No. spec.</u>					<u>%</u>
GASTROPODA						
<i>Cirsotrema (Cirsotrema) mariae</i> (Tate)	7					1.84
<i>Turritella</i> sp.	several					-
<i>Calyptraea</i> sp.	4					1.05
<i>Siliquaria</i> s.l. sp.	several					-
<i>Emarginula</i> sp.	2					0.52
<i>Trochus</i> sp.	4					1.05
? <i>Patelloida</i> sp.	1					0.26
<i>Gastropoda</i> sp.	1					0.26
<i>Gastropoda</i> sp.	1					0.26
<i>Gastropoda</i> sp.	1					0.26
	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>no. total spec.</u>	<u>%</u>
BIVALVIA						
<i>Pycnodonte tatei</i> (Suter)	31	8			39	10.24
<i>Limatula margaritata</i> Buonaiuto		1		8	9	2.36
<i>Lima maslinensis</i> Buonaiuto	3				3	0.79
<i>Divarilima</i> cf. <i>polyactina</i> (Tate)				1	1	0.26
<i>Chlamys (Chlamys)</i> <i>aldingensis</i> (Tate)	24	12		26	62	16.27
<i>C. (Chlamys) flindersi</i> (Tate)	18	9	1	17	45	11.81
<i>C. (Chlamys) peroni</i> (Tate)	16	12		12	40	10.50
<i>Dimya sigillata</i> Tate	2	5			7	1.84
<i>Barbatia (Barbatia)</i> cf. <i>limatella</i> (Tate)	1	2			3	0.79
<i>Cucullaea (Cucullaea)</i> <i>adelaidensis</i> Tate	10	3			13	3.41
<i>Spondylus (Spondylus)</i> <i>tortachillensis</i> sp.nov.				7	7	1.84
<i>Pteria</i> sp.	3				3	0.79

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>no. total spec.</u>	<u>%</u>
<i>Glycymeris (Glycymeris) cf. kaurna</i> sp. nov.	3	2			5	1.31
<i>G. (Glycymeris) cf. lenticularis</i> (Tate)	1	1			2	0.52
<i>Gari</i> sp.	7	5			12	3.15
<i>Hiatella (Hiatella)?vera</i> (Deshayes)	15	31			46	12.07
<i>Glans</i> sp. <u>A</u>		1			1	0.26
<i>Glans</i> sp. <u>B</u>	3	2			5	1.31
<i>Dosina</i> sp.	8	9	1		18	4.72
<i>Mactra</i> sp.	2	3			5	1.31
<i>Corbula</i> sp.			1		1	0.26
<i>Nemocardium</i> sp.	7	7			14	3.67
<i>Periglypta</i> sp.		1		6	7	1.84
<i>Nuculana</i> sp.	1				1	0.26
<i>Clavagella lirata</i> (Tate)			2		2	0.52
<i>Jouannetia (Pholadopsis) cuneata</i> (Tate)			1		1	0.26
<i>Myadora</i> sp.		1			1	0.26
? <i>Cuna</i> sp.		1			1	0.26
<i>Herella</i> sp.	2	1			3	0.79
<i>Divalucina</i> sp.		1			1	0.26
<i>Katelsysia</i> sp.	1				1	0.26

ALIA

Echinoids, Brachiopoda, Cirripedia (two forms), shark teeth, Vermes,

Bryozoa.

DIVERSITY INDEXES			<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Tot.</u>
			<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	
N=381	N _s =42	Specimens	21	5.51	-	-	360	94.49	381
S _s =42	S _g =39	Species	10	23.81	-	-	32	76.19	42
D _s =.9170	D _g =.9954	Genera	10	25.64	-	-	29	74.36	39

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>VV</u>	<u>Total</u>	BIVALVIA	
Epifaunal	124	83	1	71	279		<u>%</u>
Infaunal	35	35	5	6	81	Epifaunal	77.5
uncertain	-	-	-	-	-	Infaunal	22.5
Total	159	118	6	77	360		
%	(44.17)	(32.78)	(1.67)	(21.39)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEX			
	<u>spec.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty ind.</u>
Infaunal	76	0.0004	Epifaunal	278	0.0304	0.0150
Epifaunal	278	0.0000	Infaunal	76	0.0274	0.0052
aggregate	354	0.0002	aggregate	354	0.0252	0.0095

TL3

	<u>no. spec.</u>	<u>%</u>
GASTROPODA		
<i>Siliquaria</i> s.l. sp.	14	1.01
<i>Calyptraea</i> sp.	7	0.51
? <i>Cellana</i> sp.	1	0.07
<i>Emarginula</i> sp.	6	0.43
<i>Cirsotrema mariae</i> (Tate)	17	1.23
<i>Natica</i> sp.	9	0.65
? <i>Vexillum</i> sp.	6	0.43
<i>Turritella</i> sp.	9	0.65
<i>Marginella</i> sp.	1	0.07
<i>Cypraea</i> sp.	2	0.14
<i>Trochus</i> sp.	19	1.38
<i>Olivella</i> sp.	1	0.07
<i>Scaphander</i> sp.	2	0.14
<i>Gastropoda</i> sp. ind. & juv.	27	1.96
<i>Latirus</i> sp.	1	0.07
SCAPHOPODA		
<i>Dentalium</i> s.l. sp.	11	0.80

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.spec.</u>	<u>%</u>
BIVALVIA						
<i>Pycnodonte tatei</i> (Suter)	3	4			7	0.51
<i>Pycnodonte tatei</i> juv. morphā <u>A</u>	3	7	1		11	0.80
<i>Pycnodonte tatei</i> juv. morphā <u>B</u>	11	9			20	1.45
<i>Ostrea</i> sp.				2	2	0.14
<i>Ostrea</i> sp. juv.	1				1	0.07
<i>Limatula margaritata</i> Buonaiuto	1			56	57	4.13
<i>Ctenoides</i> sp. nov. aff. <i>linguliformis</i> (Tate)		2			2	0.14
<i>Lima maslinensis</i> Buonaiuto	3	2		1	6	0.43
<i>C. (Chlamys) aldingensis</i> (Tate)	8	2		14	24	1.74
<i>C. (Chlamys) peroni</i> (Tate)	29	14		68	111	8.04
<i>C. (Chlamys) flindersi</i> (Tate)	2	3		20	25	1.81
<i>Spondylus tortachillensis</i> sp. nov.	6	1		26	33	2.39
<i>Dimya sigillata</i> Tate	297	368	5	21	691	50.07
<i>Glycymeris</i> cf. <i>kaurna</i> sp. nov.	13	5		8	26	1.88
<i>C. (Cucullaea)</i> cf. <i>adelaidensis</i> Tate	5	2		1	8	0.58
<i>B. (Barbatia)</i> cf. <i>limatella</i> Tate	5	6			11	0.80
<i>Allasinazella</i> gen.nov. cf. <i>equidens</i> (Tate)	1	1			2	0.14
<i>Nucula</i> sp.	1	2			3	0.22
<i>H. (Hiatella) ?vera</i> Deshayes	6	28			34	2.46
<i>Limopsis</i> cf. <i>zitteli</i> Ihering	1				1	0.07
<i>Propeamussium</i> (<i>Parva-</i> <i>mussium</i>) sp.				11	11	0.80
<i>Crenella</i> cf. <i>globularis</i> Tate	1	1			2	0.14

	<u>LY</u>	<u>RV</u>	<u>BY</u>	<u>vy</u>	<u>tot.no.spec.</u>	<u>%</u>	(20)
<i>Limea (Isolimea) alticosta</i> Tate				14	14	1.01	
<i>Gari</i> sp.	13	9		6	28	2.03	
<i>Clavagella lirata</i> (Tate)			2		2	0.14	
? <i>Pteria</i> sp.		2			2	0.14	
<i>Mactra</i> sp.	4	4			8	0.58	
<i>Dosina</i> sp.	13	9			22	1.59	
<i>Myadora</i> sp.		1			1	0.07	
<i>Corbula</i> sp.	1	1			2	0.14	
<i>Lucina</i> sp.	3	2			5	0.36	
<i>Dosinia</i> sp.	2	3			5	0.36	
<i>Herella</i> sp.	3				3	0.22	
<i>Bivalve</i> sp. ind.				6	6	0.43	
<i>Propeleda</i> sp.	1				1	0.07	
<i>Nemocardium</i> sp.	15	16		11	42	3.04	
<i>Glans</i> sp. <u>A</u>	1				1	0.07	
<i>Glans</i> sp. <u>B</u>	6	7		2	15	1.09	
<i>Nuculana</i> sp.	1				1	0.07	

ALIA

Brachiopoda, Echinoids, Cirripedia (two forms), Crustacea (crab claws), shark teeth, Vermes, Bryozoa.

DIVERSITY INDEXES			<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Tot.</u>	
			<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>		
N = 1380	N _s = 51		Specimens	122	8.84	11	.80	1247	90.36	1380
S _s = 51	S _g = 46		Species	14	27.45	1	2.78	36	70.59	51
D _s = .7357	D _g = .9961		Genera	14	30.43	1	2.17	31	67.39	46

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Total</u>	BIVALVIA	<u>%</u>
Epifaunal	382	453	6	234	1.075	Epifaunal	86.21
Infaunal	65	58	2	41	166	Infaunal	13.31
Uncertain	-	-	-	6	6	Uncertain	.48
<hr/>							
Total	447	511	8	282	1.247		
(%)	(35.85)	(40.98)	(0.6)	(22.61)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty</u>
						<u>index</u>
Infaunal	164	0.0005	Infaunal	1.069	0.0781	0.0084
Epifaunal	1.069	0.00002	Epifaunal	164	0.0275	0.0074
Aggregate	1.241	0.00001	Aggregate	1.241	0.0584	0.0064

TL4

GASTROPODA	<u>Tot. no. specs.</u>	<u>%</u>
<i>Cirsotrema mariae</i> (Tate) juvs. & ads.	42	2.69
<i>Calyptraea</i> sp.	2	0.13
<i>Cypraea</i> sp.	16	1.02
<i>Olivella</i> sp.	5	0.32
? <i>Terebra</i> sp.	1	0.06
<i>Scaphander</i> sp.	3	0.19
<i>Emarginula</i> sp.	4	0.26
<i>Latirus</i> sp.	3	0.19
<i>Siliquaria</i> s.l. sp.	11	0.70
<i>Natica</i> sp.	1	0.06
<i>Voluta</i> sp.	3	0.19
<i>Turritella</i> sp.	6	0.38
<i>Trochus</i> sp.	7	0.45
<i>Turbonilla</i> sp.	1	0.06
Gastropoda sp.	40	2.56

SCAPHOPODA		<u>tot.no.specs.</u>				<u>%</u>	
<i>Dentalium</i>	s.l. sp.			10		0.64	
BIVALVIA		<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Limea (Isolimea) alticosta</i>	(Tate)	44	53		140	237	15.16
<i>Dimya sigillata</i>	Tate	276	281	5	119	681	43.57
<i>Limatula margaritata</i>	Buonaiuto	5	5		36	46	2.94
<i>Pycnodonte tatei</i>	(Suter) juvs.	31	37			68	4.35
<i>Chlamys</i>	sp.	6	3		21	30	1.92
<i>C. (Chlamys) aldingensis</i>	(Tate)	4			14	18	1.15
<i>C. (Chlamys) flindersi</i>	(Tate)	1	1		4	6	0.38
<i>C. (Chlamys) peroni</i>	(Tate)	7			49	56	3.58
<i>Spondylus tortachillensis</i>	sp. nov.	3	1		5	9	0.60
<i>Propeamussium (Parvamussium)</i>	sp.		1		59	60	3.84
<i>Lima maslinensis</i>	Buonaiuto				5	5	0.32
<i>Cuspidaria (Rhinoelama)</i>	sp.	1	1			2	0.13
<i>Solamen (Exosiperna) cf. globularis</i>	(Tate)	3				3	0.19
<i>Hiatella (Hiatella) ?vera</i>	(Deshayes)	14	9			23	1.47
<i>Tivelina</i>	sp.	2	3			5	0.32
<i>Clavagella lirata</i>	Tate			1		1	0.06
<i>Cuspidaria (Cuspidaria)</i>	sp.	1				1	0.06
<i>Corbula</i>	sp.	1				1	0.06
? <i>Lucina</i>	sp.		1			1	0.06
<i>Mactra</i>	sp.	8				8	0.51
<i>Tellina</i>	sp.	11	6		11	28	1.79
<i>Dosina</i>	sp.	6	10			16	1.02

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot. no. specs.</u>	<u>%</u>
<i>Glycymeris</i> sp				11	11	0.70
<i>Nemocardium</i> sp	15	15		4	34	2.18
<i>C. (Cucullaea) cf. adelaidensis</i> Tate	2	1			3	0.19
<i>Arca</i> sp.	3	3			6	0.38
<i>Pronucula</i> sp.			1		1	0.06
<i>Bivalve</i> gen. & sp. ind. & juv.	15	15		18	48	3.07

ALIA

Brachiopoda, Cirripedia (two forms), Crustacea (Crabs' claws), Echinoids, Shark teeth and embryonic teeth, Vermes, Bryozoa.

DIVERSITY INDEXES		<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Total</u>	
		<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>		
N=1.563	N _s =43	Specimens	145	9.28	10	0.64	1408	90.08	1563
S=43	S _g =39	Species	15	34.88	1	2.33	27	62.79	43
D _s =.7778	D _g =.9956	Genera	15	38.46	1	2.56	23	58.97	39

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	<u>BIVALVIA</u>	<u>%</u>
Infaunal	51	30	2	26	109	Infaunal	7.74
Epifaunal	399	395	5	452	1.251	Epifaunal	88.85
Uncertain	15	15	-	18	48	Uncertain	3.41
Total	465	440	7	496	1.408		
(%)	(33.03)	(31.25)	(0.5)	(35.23)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty index</u>
Epifaunal	1.246	0.00001	Epifaunal	1.246	0.0509	0.0266
Infaunal	107	0.0037	Infaunal	107	0.0360	0.0108
Aggregate	1.401	0.0004	Aggregate	1.401	0.0406	0.0214

	<u>no. specs.</u>	<u>%</u>
GASTROPODA		
<i>Cirsotrema mariae</i> (Tate) juvs & ads	18	1.33
<i>Emarginula</i> sp.	9	0.66
<i>Calyptreaea</i> sp.	3	0.22
<i>Turbo</i> sp.	28	2.07
<i>Trochus</i> sp.	15	1.11
<i>Latirus</i> sp.	4	0.30
<i>Turritella</i> sp.	13	0.96
<i>Gastropoda</i> ind.	4	0.30
<i>Cypraea</i> sp.	6	0.45
<i>Calliostoma</i> (Fautor) <i>allasinazi</i> sp. nov.	1	0.07
<i>Guildfordia</i> (<i>Pseudostraliium</i>) <i>maslinensis</i> sp. nov.	1	0.07
<i>Siliquaria</i> s.l. sp.	13	0.96

SCAPHOPODA

<i>Dentalium</i> sp.	30	2.22
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	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Tot. no. specs.</u>	<u>%</u>
<i>Limatula margaritata</i> Buonaiuto	14	14		24	52	3.76
<i>Limea</i> (<i>Isolimea</i>) <i>alticosta</i> (Tate)	42	48		43	133	9.61
<i>Divarilima polyactina</i> (Tate)	1	2			3	0.22
<i>Dimya sigillata</i> (Tate)	195	162		47	404	29.19
<i>Pycnodonte tatei</i> (Suter)	18	17		4	39	2.82
<i>Pycnodonte</i> sp.				1	1	0.07
<i>Lima maslinensis</i> Buonaiuto	2	2		4	8	0.58
<i>Spondylus tortachillensis</i> sp. nov.		1		1	2	0.14
<i>C.</i> (<i>Chlamys</i>) <i>aldingensis</i> (Tate)	10	3		21	34	2.46
<i>C.</i> (<i>Chlamys</i>) <i>flindersi</i> (Tate)	2	2		3	7	0.51
<i>C.</i> (<i>Chlamys</i>) <i>peroni</i> (Tate)	12	11		19	42	3.03
<i>Chlamys</i> sp.	9			23	32	2.31
<i>Propeamussium</i> (<i>Parvamussium</i>) sp.				14	14	1.01
<i>Nemocardium</i> sp.	61	33		23	117	8.45

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Tot.no.specs.</u>	<u>%</u>
<i>Clavagella lirata</i> (Tate)			28		28	2.02
<i>Macra</i> sp.	12	9			21	1.52
<i>Corbula</i> sp.	2	2			4	0.29
<i>Gari</i> sp.	11	4		3	18	1.30
<i>H. (Hiatella)?vera</i> (Deshayes)	14	21			35	2.53
<i>Glans</i> sp. B	2	1			3	0.22
<i>Herella</i> sp.	2	2			4	0.29
<i>C. (Cucullaea) adelaidensis</i> (Tate)	18	12		3	33	2.38
<i>Dosina</i> sp.	11	4			15	1.08
<i>Dosinia</i> sp.	5	8			13	0.94
<i>Glycymeris</i> sp.	8	7		2	17	1.23
<i>B. (Barbatia) cf. limatella</i> (Tate)	8	8			16	1.16
? <i>Porterius (Ludbrookella) cf. spinosus</i> sp. nov.	2	2			4	0.29
<i>Arca</i> sp.		1			1	0.07
<i>Arcopsis cf. dissimilis</i> (Tate)		1			1	0.07
<i>Limopsis zitteli</i> Ihering	2				2	0.14
<i>Pronucula</i> sp.	2	2			4	0.29
<i>Bivalvia</i> sp. ind.	37	30	2	36	103	7.44

ALIA

Barachiopoda, Echinoids, Vermes, Cirripedia (two forms), crabs' claws, shark teeth, embryonic teeth, Bryozoa.

DIVERSITY INDEXES			<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Tot.</u>
			<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	<u>no.</u>	<u>%</u>	
N = 1384	N = 47	Specimens	119	8.59	30	2.16	1.235	89.23	1.384
S _s = 47	S _g = 43	Species	12	25.53	1	2.12	34	72.39	47
D _s = .8851	D _g = .9963	Genera	12	27.90	1	2.32	30	69.76	43

VALVES						BIVALVIA	
	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Total</u>		<u>%</u>
Infaunal	118	72	28	51	269	Infaunal	21.78
Epifaunal	347	307	-	207	861	Epifaunal	69.72
Uncertain	37	30	2	36	105	Uncertain	8.50
Total	502	409	30	294	1.235		
(%)	(40.65)	(33.12)	(2.43)	(23.81)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty index</u>
Infaunal	241	0.0131	Infaunal	241	0.0552	0.0081
Epifaunal	861	0.0000	Epifaunal	861	0.0709	0.0089
Aggregate	1.205	0.0005	Aggregate	1.205	0.0326	0.0053

TL6

GASTROPODA

Turbo sp.no. specs.%

1

0.85

Natica sp.

2

1.71

Turritella sp.

10

8.55

Gastropoda sp. ind.

4

3.42

SCAPHOPODA

Dentalium sp.

1

0.85

BIVALVIA

LVRVBVvvtot.no.specs.%*Bivalvia* sp. ind.

1

4

15

20

17.09

Ostrea sp.

2

2

1.71

Dosina sp.

8

2

1

4

15

12.82

C. (Chlamys) peroni (Tate)

2

2

1.71

Lima maslinensis Buonaiuto

2

2

1.71

Chlamys sp.

1

1

30

32

27.35

Limea (Isolimea) alticosta (Tate)

3

3

2.56

Dimya sigillata Tate

2

2

1.71

C. (Chlamys) flindersi (Tate)

6

6

5.13

Limatula margaritata Buonaiuto

1

1

2

4

3.42

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Corbula</i> sp.	1	1			2	1.71
<i>Clavagella lirata</i> (Tate)			1		1	0.85
<i>Gari</i> sp.		1		1	2	1.71
<i>Cardium</i> sp.				1	1	0.85
? <i>Pteria</i> sp.		1			1	0.85
<i>Glycymeris</i> sp.				1	1	0.85
<i>C. (Cucullaea) cf. adelaidensis</i> Tate	1	1			2	1.71
<i>Arca</i> sp.	1				1	0.85

ALIA

Brachiopoda, Echinoids, Vermes, Sharks' teeth, Cirripedia (1 form), Bryozoa.

DIVERSITY INDEXES		<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Total</u>
		no.	%	no.	%	no.	%	
$N = 117$	$N = 23$	Specimens	17 14.52	1 0.85	99 84.61	117		
$S_s = 23$	$S_g = 21$	Species	4 17.39	1 4.34	18 78.26	23		
$D_s = .8712$	$D_g = .9960$	Genera	4 19.08	1 4.76	16 76.19	21		

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Total</u>	BIVALVIA	<u>%</u>
Epifaunal	4	4	-	50	58	Infaunal	21.21
Infaunal	9	4	2	6	21	Epifaunal	58.59
Uncertain	1	4	-	15	20	Uncertain	20.20
Total	14	12	2	71	99		
(%)	(14.14)	(12.12)	(2.02)	(71.71)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty index</u>
Infaunal	19	0.0000	Infaunal	19	0.1637	0.0351
Epifaunal	58	0.0000	Epifaunal	58	0.0000	0.2771
Aggregate	97	0.0000	Aggregate	97	0.0318	0.1222

BLANCHE POINT FORMATION'Transitional Marl Member'

POCKETS

(Section B - 'Uncle Tom's Cabin')

GASTROPODA					<u>no. specs.</u>	<u>%</u>
<i>Turbo</i> sp.					2	2.20
<i>Turritella</i> sp.					18	19.78
<i>Cypraea</i> sp.					1	1.10
BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>C. (Chlamys) peroni</i> (Tate)	1	2		4	7	7.69
<i>C. (Chlamys) flindersi</i> (Tate)				3	3	3.30
<i>Chlamys</i> sp.	1	1		12	14	15.38
<i>Limea (Isolimea) alticosta</i> (Tate)		1		2	3	3.30
<i>Dosina</i> sp.	1	4		7	12	13.19
<i>Bivalvia</i> sp. ind. & juvs				5	5	5.49
<i>Lima maslinensis</i> Buonaiuto				2	2	2.20
<i>Limatula margaritata</i> Buonaiuto		1		3	4	4.40
<i>Clavagella lirata</i> (Tate)			1		1	1.10
<i>Ostrea</i> sp.				1	1	1.10
<i>Mactra</i> sp.				1	1	1.10
<i>Dimya sigillata</i> Tate		4		3	7	7.69
<i>C. (Cucullaea) cf. adelaidensis</i> Tate	2	2			4	4.40
<i>Area</i> sp				3	3	3.30
<i>Nemocardium</i> sp.	1				1	1.10
<i>Glans</i> sp. B		1			1	1.10
<i>Gari</i> sp.				2	2	2.20
<i>Nucula</i> sp.	2				2	2.20

ALIA

Echinoids, Brachiopods, Cirripedia, Bryozoa.

DIVERSITY INDEXES		Gastropoda		Scaphopoda		Bivalvia		Total
N = 91	N = 21	n.	%	no.	%	no.	%	
$S_s = 21$	$S_s = 19$	Specimens	21 23.07	-	-	70	76.92	91
$D_s = .9343$	$D_s = .9952$	Species	3 14.28	-	-	18	85.72	21
		Genera	3 15.78	-	-	16	84.21	19

VALVES	LV	RV	BV	vv	Total	BIVALVIA	%
Epifaunal	4	11	-	33	48	Epifaunal	68.57
Infaunal	4	5	1	7	17	Infaunal	24.29
Uncertain	-	-	-	5	5	Uncertain	7.14
Total	8	16	1	45	70		
(%)	(11.43)	(22.85)	(1.43)	(64.29)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	no. specs.	index		no. specs.	index	uncertainty index
Infaunal	16	0.0000	Infaunal	16	0.0083	0.1833
Epifaunal	48	0.0000	Epifaunal	48	0.0009	0.0762
Aggregate	69	0.0000	Aggregate	69	0.0000	0.0503

TMI
(Section D-Reynolds Cave)

	<u>no. specs.</u>	<u>%</u>
GASTROPODA		
<i>Trophonopsis hypsellus</i> (Tate)	1	0.22
<i>Cirsotrema mariae</i> (Tate)	1	0.22
<i>Cypraea</i> sp.	1	0.22
<i>Gastropoda</i> sp. A.	2	0.45
<i>Gastropoda</i> sp. B.	8	1.79
<i>Gastropoda</i> larvae spp. ind.	71	15.85
<i>Calyptreaea</i> sp.	1	0.22
<i>Turbo</i> ss. 11. sp.	7	1.56
<i>Calliostoma</i> (Fautor) <i>allasinazi</i> sp. nov.	2	0.45
<i>Guildfordia</i> (<i>Pseudastralium</i>) cf. <i>maslinensis</i> sp. nov.	1	0.22
<i>Emarginula</i> sp.	1	0.22
<i>Spirocolpus aldingae</i> (Tate)	210	46.87

SCAPHOPODA

					<u>no. specs.</u>	%
<i>Dentalium</i> sp.					2	0.45
BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Corbula</i> (<i>Cariocorbula</i>) <i>pyxidiata</i> (Tate)		1			1	0.22
<i>Paraglans</i> <i>latissima</i> (Tate)	1	1			2	0.45
<i>Nuculana</i> (<i>Saccella</i>) <i>chapmani</i> (Finlay)			1		1	0.22
<i>Limopsis</i> (<i>Limopsis</i>) <i>zitteli</i> Ihering		2		1	3	0.67
<i>Pycnodonte</i> <i>tatei</i> (Suter)	1	1		1	3	0.67
<i>Limatula</i> <i>margaritata</i> Buonaiuto				3	3	0.67
<i>Limea</i> (<i>Isolimea</i>) <i>alticosta</i> (Tate)	1			1	2	0.45
<i>Lima</i> <i>maslinensis</i> Buonaiuto	1			4	5	1.12
<i>Vulsella</i> (<i>Vulsella</i>) <i>laevigata</i> Tate	1	3		1	5	1.12
<i>Phygraea</i> <i>tarda</i> (Hutton) juv. (?)	1				1	0.22
<i>Ostrea</i> sp. A juv.	3				3	0.67
<i>Spondylus</i> <i>tortachillensis</i> sp. nov.			3	3	6	1.34
<i>Lentipecten</i> (<i>Lentipecten</i>) sp. aff. <i>hochstetteri</i> (Zittel)	1			3	4	0.89
<i>Ostrea</i> sp. B juv.			4		4	0.89
<i>Clavagella</i> <i>lirata</i> (Tate)			4		4	0.89
<i>Dimya</i> <i>sigillata</i> Tate	7		18	7	32	7.14
<i>Modiolus</i> (<i>Modiolus</i>) cf. <i>adelaidensis</i> (Tate)	2				2	0.45
<i>Tellina</i> (<i>Tellina</i>) sp. aff. <i>cainozoica</i> T. Woods	1				1	0.22
<i>Dosina</i> sp.	4	3		2	9	2.01
<i>Glycymeris</i> sp.				2	2	0.45
<i>Cardium</i> sp.				2	2	0.45
<i>Nemocardium</i> sp.		1		1	2	0.45
<i>Arca</i> (<i>Arca</i>) <i>pseudonavicularis</i> Tate	2				2	0.45
<i>Barbatia</i> (<i>Barbatia</i>) <i>limatella</i> Tate	1				1	0.22
<i>C.</i> (<i>Chlamys</i>) <i>peroni</i> (Tate)		2		12	14	3.12
<i>C.</i> (<i>Chlamys</i>) <i>flindersi</i> (Tate)	2	1		12	15	3.35
<i>Chlamys</i> sp. ind. & juvs.	1		1	9	11	2.46

ALIA

Echinoids, Brachiopoda, Cirripedia, Decapoda (crabs' claws), Bryozoa,
Shark teeth, embryonic teeth.

DIVERSITY INDEXES		Gastropoda		Scaphopoda		Bivalvia		total	
N = 440	N _s = 40	no.	%	no.	%	no.	%		
		Specimens	306	68.29	2	0.45	140	31.26	448
S _s = 40	S _g = 38	Species	12	30.00	1	2.50	27	67.50	40
D _s = .9640	D _g = .9940	Genera	12	32.43	1	2.70	24	64.86	37

VALVES	LV	RV	BV	vv	total	BIVALVIA	
							%
Epifaunal	24	7	26	56	113	Epifaunal	80.71
Infaunal	6	8	5	8	27	Infaunal	19.29
Uncertain	-	-	-	-	-	Uncertain	-
<hr/>							
Total	30	15	31	64	140		
(%)	(21.43)	(10.71)	(22.14)	(45.71)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	no.specs.	index		no.specs.	index	uncertainty index
Infaunal	122	0.0969	Infaunal	122	0.0260	0.0130
Epifaunal	87	0.0256	Epifaunal	87	0.0072	0.0545
			Aggregate	103	0.0063	0.0394

GASTROPODA	TM2	no. specs.	%
<i>Spirocolpus aldingae</i> (Tate)		102	21.75
<i>Emarginula</i> sp.		2	0.43
<i>Voluta</i> s.l. sp.		2	0.43
<i>Natica</i> s.l. sp.		2	0.43
<i>Calyptraea</i> sp.		1	0.21
<i>Turbo</i> s.l. sp.		24	5.12
<i>Trochus</i> s.l. sp.		2	0.43
<i>Gastropoda</i> spp. ind. & juvs.		8	1.71
? <i>Nototrivia</i> sp.		1	0.21

BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Dimya sigillata</i> Tate	39	77			116	24.73
<i>Goniocardium</i> sp.	3	1	1		5	1.07
<i>Vepricardium (Hedecardium) monilectum</i> (Tate)	4				4	0.85
<i>Dosina cf. multilamellata</i> (Tate)	17	6	2		25	5.33
<i>Paraglans latissima</i> (Tate)		2			2	0.43
<i>Cucullaea adalaidensis</i> Tate	1				1	0.21
<i>Glycymeris</i> sp.		1			1	0.21
<i>Bivalvia</i> spp. ind.	11	5	1	15	32	6.82
<i>Pycnodonte tatei</i> (Suter)	33	31		4	68	14.50
? <i>Lanternula</i> sp.		1			1	0.21
<i>Lima maslinensis</i> Buonaiuto		1			1	0.21
<i>Spondylus tortachillensis</i> Buonaiuto		1		4	5	1.07
<i>C. (Chlamys) peroni</i> (Tate)	1	2		20	23	4.90
<i>C. (Chlamys) flindersi</i> (Tate)	2	4		25	31	6.61
<i>Chlamys</i> sp.				10	10	2.13

DIVERSITY INDEXES		Gastropoda		Scaphopoda		Bivalvia		Total
N_s	N_g	no.	%	no.	%	no.	%	
$N_s = 469$	$N_g = 24$	144	30.29	-	-	325	69.71	469
$S_s = 24$	$S_g = 22$	9	37.50	-	-	15	62.50	24
$D_s = .8543$	$D_g = .9891$	9	40.91	-	-	13	59.09	22
VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	BIVALVIA		
Epifaunal	76	116	-	63	265	%		
Infaunal	25	11	3	-	38	Epifaunal	78.46	
Uncertain	11	5	1	15	32	Infaunal	11.69	
Total	111	132	4	78	325	Uncertain	9.85	
(%)	(34.15)	(40.62)	(1.23)	(24.00)				

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>no specs</u>	<u>index</u>		<u>no specs</u>	<u>index</u>	<u>uncertainty index</u>
Infaunal	38	0.0014	Infaunal	38	0.2063	0.0000
Epifaunal	255	0.0000	Epifaunal	255	0.0392	0.6255
Aggregate	321	0.0002	Aggregate	321	0.0286	0.0127

Check samples from
Section V, between 'Uncle Tom's Cabin' and Reynolds Cave

S. 1

Chlamys flindersi (Tate), *Dimya sigillata* (Tate), *Gastropoda* sp. ind.

S. 2

Chlamys sp., *Dimya sigillata* (Tate)

S. 3

Chlamys flindersi (Tate), *Chlamys peroni* (Tate), *Dimya sigillata* (Tate), shark teeth. *Hantkenina primitiva* horizon. First appearance of fish bones.

S. 4

Chlamys flindersi (Tate), *Phygraea tarda* (Hutton) *Spirocolpus aldingae* (Tate), *Arca* sp. juv., fish bones.

S. 5

Chlamys flindersi (Tate), *Chlamys peroni* (Tate).

SECTION V
(between S.2 and S.3)

GASTROPODA	no. specs.	%
<i>Cirsotrema mariae</i> (Tate)	1	0.33

BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>Tot.no.</u>	<u>%</u>
					<u>specs</u>	
<i>C. (Chlamys) flindersi</i> (Tate)	7	22	1	110	140	45.90
<i>C. (Chlamys) peroni</i> (Tate)				8	8	2.62
<i>Pycnodonte tatei</i> (Suter)	91	47	3	6	147	48.20
<i>Dimya sigillata</i> Tate		4			4	1.31
<i>Chlamys</i> sp. ind.				2	2	0.66
<i>Spondylus</i> cf. <i>tortachillensis</i> sp. nov.				3	3	0.98

ALIA

Bryozoa.

DIVERSITY INDEXES						no	%	
N = 305	N _s = 7					Gastropoda	1	0.33
S _s = 7	S _g = 5					Bivalvia	304	99.67
D _s = .5578	D _g = .8571							
VALVES	LV	RV	BV	vv	total	BIVALVIA		
Epifaunal	98	73	4	129	304	Epifaunal	100%	
(%)	(32.24)	(24.01)	(1.30)	(42.43)				
ARTICULATION RATIO : 0.0001						DIFFERENTIAL TRANSPORT INDEX: 0.0918 (uncertainty index: 0.1347)		

GASTROPODA	no specs	%
<i>Cirsotrema mariae</i> (Tate)	1	0.94
<i>Gastropoda</i> spp. ind.	11	10.34

BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>C. (Chlamys) flindersi</i> (Tate)				21	21	19.81
<i>C. (Chlamys) peroni</i> (Tate)				25	25	23.58
<i>Lentipeecten</i> sp.aff. <i>hochstetteri</i> (Zittel)				1	1	0.94
<i>Lima maslinensis</i> Buonaiuto				2	2	1.89
<i>Dimya sigillata</i> Tate	12	25			37	34.91
<i>Bivalvia</i> spp. ind.				4	4	3.77
<i>Pycnodonte tatei</i> (Suter)	3	1		4	4	3.77

ALIA

Brachiopoda, Echinoids, Vermes, Bryozoa, Cirripedia (tall form), Fish bones, Sharks' teeth.

OBSERVATIONS. Sample from a highly dissolved interval

DIVERSITY INDEXES							
N = 106	N _s = 9					<u>no.</u>	<u>%</u>
S _s = 9	S _g = 8					Gastropoda	12 11.32
D _s = .7765	D _g = .9722					Bivalvia	94 88.67
VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	BIVALVIA	%
Epifaunal	15	26	-	49	90	Epifaunal	95.74
Uncertain	-	-	-	4	4	Uncertain	4.26
Total	15	26	-	53	94		
(%)	(15.96)	(27.66)	-	(56.38)			
ARTICULATION RATIO: 0.0000				DIFFERENTIAL TRANSPORT INDEX: 0.0158 (Uncertainty index: 0.1183)			

GASTROPODA	<u>tot.no.specs</u>	<u>%</u>
<i>Spirocolpus aldingae</i> (Tate) (260 ads + 1.709 juvs)	1.969	77.61
<i>Pterynotus</i> (<i>Pterochelus</i>) <i>adelaidensis</i> (Tate)	1	0.04
<i>Cylichna angustata</i> (Tate & Cossmann)	6	0.24
<i>Acteon subscalatus</i> (Tate & Cossmann)	38	1.50
<i>Siliquaria altispira</i> sp. nov.	8	0.32
<i>Lunatia aldingensis</i> (Tate)	4	0.16
<i>Cylichna</i> (<i>Cylichnania</i>) <i>callosa</i> (Tate & Cossmann)	7	0.28
<i>Cassoginella palla</i> (Cotton)	1	0.04
<i>Aerocoelum margaritatum</i> sp. nov.	1	0.04
<i>Kosugeia</i> gen.nov. <i>costatosulcata</i> sp. nov.	2	0.08
<i>Kaurmaginella</i> gen.nov. <i>tutgka</i> sp.nov.	12	0.47
<i>Alaginella submicula</i> (Tate)	1	0.04
<i>Cirsotrema mariae</i> (Tate)	1	0.04
<i>Margarites</i> (<i>Periaulax</i>) <i>rhysus</i> sp.nov. (ex Tate)	7	0.28
<i>Basilissa</i> (<i>Basilissa</i>) <i>cossmanni</i> Tate	1	0.04
<i>Calliostoma</i> (<i>Fautor</i>) <i>allasinazi</i> sp. nov.	1	0.04
<i>Pterynotus</i> (<i>Pterochelus</i>) <i>bifrons</i> (Tate)	1	0.04
<i>Argobuccinum</i> (<i>Cymatiella</i>) <i>oligostira</i> (Tate)	3	0.12
<i>Austromitra pumila</i> (Tate)	12	0.47
<i>Vexithara citharelloides</i> (Tate)	1	0.04
<i>Perynotus</i> (<i>Pterochelus</i>) <i>manubriatus</i> (Tate)	1	0.04
<i>Typhis</i> (<i>Talityphis</i>) <i>tetraphyllos</i> sp.nov. (exTate)	3	0.12
<i>Baryspira</i> (<i>Gracilispira</i>) <i>ligata</i> (Tate)	4	0.16
<i>Waimatea complanata</i> (Tate)	3	0.12
<i>Sigmesalia stylacris</i> (Tate)	3	0.12
<i>Rugobela</i> sp. nov.	2	0.08
<i>Trophon</i> (<i>Enantimene</i>) <i>monotropis</i> (Tate)	11	0.43
<i>Brocchitas altior</i> sp. nov.	4	0.16
<i>Notopeplum protorhysum</i> (Tate)	1	0.04
<i>Emarginula</i> (<i>Emarginula</i>) <i>imbricata</i> sp. nov.	1	0.04

	<u>tot.no.specs</u>	<u>%</u>
<i>Leucorhynchia bifuniculata</i> sp. nov.	3	0.12
<i>Austrofuscus</i> sp.	8	0.32
<i>Trophon</i> (<i>Trophonopsis</i>) <i>hypsellus</i> (Tate)	72	2.84
<i>Praehyalocylis annulata</i> (Tate)	4	0.16
? <i>Eumetula</i> (<i>Eumetula</i>) sp. nov. A	3	0.12
? <i>Euseila</i> sp. nov.	2	0.08
? <i>Cerithiella</i> sp. nov. A	4	0.16
? <i>Cerithiella</i> sp. nov. B	1	0.04
? <i>Cerithiella</i> sp. nov. C	1	0.04
? <i>Cerithiopsis</i> (<i>Socienna</i>) sp. nov.	4	0.16
? <i>Notoseila</i> sp. nov.	3	0.12
? <i>Cerithiopsis</i> (<i>Miopila</i>) sp. nov.	2	0.08
? <i>Pilaflexis</i> sp. nov.	3	0.16
? <i>Cerithopsis</i> sp.	3	0.16
? <i>Eumetula</i> (<i>Eumetula</i>) sp. nov. B	3	0.16
? <i>Cerithidea</i> (<i>Cerithidea</i>) sp. nov.	1	0.04
<i>Sirius fenestratus</i> (Tate)	1	0.04
? <i>Cerithioderma</i> sp.	1	0.04
? <i>Cerithiopsis</i> (<i>Cerithiopsilla</i>) sp. nov.	1	0.04
<i>Charonia</i> (<i>Austrosassia</i>) <i>cribrosa</i> (Tate)	1	0.04
<i>Distorsio</i> (<i>personella</i>) <i>maslinensis</i> sp. nov.	3	0.12
<i>Fusinus</i> (<i>Fusinus</i>) <i>sculptilis</i> (Tate)	1	0.04
<i>Comitas</i> (<i>Comitas</i>) sp. nov.	2	0.08
<i>Comitas</i> (<i>Comitas</i>) <i>aldingensis</i> Powell	3	0.12
<i>Tanea falsa</i> sp. nov.	26	1.02
<i>Marginella</i> s.l. sp. nov.	1	0.04
<i>Triphora</i> (<i>Ogivia</i>) <i>trirostrata</i> sp. nov.	2	0.08
<i>Knefastia</i> sp.	18	0.71
<i>Eulima</i> (<i>Balcis</i>) sp. nov. A	4	0.16
<i>Eulima</i> (<i>Balcis</i>) sp. nov. B	1	0.04

	<u>no. specs.</u>	<u>%</u>
<i>Eulima</i> sp. nov. A	7	0.28
<i>Eulima</i> sp. nov. B	3	0.12
<i>Chemnitzia</i> sp. nov. A	1	0.04
<i>Syrnola</i> sp. nov.	1	0.04
<i>Eulima</i> (<i>Balcis</i>) sp. nov. C	1	0.04
Gastropoda spp. ind. & juvs.	56	2.21

SCAPHOPODA

<i>Gadilina tatei</i> Sharp & Pilsbry	8	0.32
<i>Fissidentalium</i> sp. nov.	6	0.24

BIVALVIA

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Pinctada</i> sp. nov.			1		1	0.04
<i>Pinna</i> sp.				1	1	0.04
<i>Dimya sigillata</i> Tate	18	17			35	1.38
<i>Limea</i> (<i>Isolimea</i>) <i>alticosta</i> (Tate)		1			1	0.04
<i>Nuculana</i> (<i>Poroleda</i>) sp. nov.		1			1	0.04
<i>N.</i> (<i>Ledella</i>) <i>leptorhyncha</i> (Tate)	13	5	6		24	0.95
<i>Pronucula tatei</i> (Finlay)	3	2	3	1	9	0.35
<i>Nuculana</i> (<i>Saccella</i>) <i>chapmani</i> (Finlay)	1	1			2	0.08
<i>Allasinazella</i> gen.nov <i>equidens</i> (Tate)	2	1	1		4	0.16
<i>Arcopsis dissimilis</i> (Tate)	4	2	3		9	0.35
<i>Barbatia</i> (<i>Barbatia</i>) <i>limatella</i> Tate	1				1	0.04
? <i>Pythina</i> sp.	1				1	0.04
<i>Dosina</i> (<i>Dosina</i>) <i>multilamellata</i> (Tate)		3	2	9	14	0.55
<i>Vulsella</i> cf. <i>laevigata</i> Tate	2			1	3	0.12
? <i>Lissarca</i> sp. nov.	7	8	2		17	0.67
<i>Pectunculina cancellata</i> sp.nov.	2	3			5	0.20
<i>Musculus</i> (<i>Musculus</i>) <i>semigranosus</i> (Tate)	1	1	2	2	6	0.24
<i>Septifer</i> (<i>Septifer</i>) sp. nov.	1	2			3	0.12
<i>Paraglans latissima</i> (Tate) juv.		1			1	0.04

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs.</u>	<u>%</u>
<i>Lima maslinensis</i> Buonaiuto				2	2	0.08
<i>C. (Chlamys) peroni</i> (Tate)				1	1	0.04
<i>Chlamys</i> sp.				5	5	0.20
<i>Cuspidaria</i> cf. <i>adelaidensis</i> (Tate)	1				1	0.04
<i>Bivalvia</i> spp. ind. & juvs.	2	2	4	7	15	0.59

ALIA

Bryozoa, Sharks' teeth, fish bones, otoliths (first occurrence), siliceous sponges (first occurrence), Echinoids, Crinoidea, Decapoda (crabs' claws), Cirripedia, Anellida, Brachiopoda.

DIVERSITY INDEXES		<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Total</u>	
N = 2537	N _s = 92	<u>no</u>	<u>%</u>	<u>no</u>	<u>%</u>	<u>no</u>	<u>%</u>		
S _s = 92	S _g = 76	Specimens	2.361	93.12	14	0.56	162	6.40	2.537
D _s = .3955	D _g = .9933	Species	66	71.74	2	2.17	24	26.09	92
		Genera	53	69.74	2	2.63	21	27.63	76

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	BIVALVIA	
							<u>%</u>
Epifaunal	29	24	7	11	71	Epifaunal	43.83
Infaunal	28	24	13	11	76	Infaunal	46.91
Uncertain	2	2	4	7	15	Uncertain	9.26
<u>Total</u>	<u>59</u>	<u>50</u>	<u>24</u>	<u>29</u>	<u>162</u>		
(%)	(36.42)	(30.86)	(18.52)	(17.90)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs.</u>	<u>index</u>		<u>specs.</u>	<u>index</u>	<u>uncertainty index</u>
Infaunal	63	0.0070	Infaunal	63	0.0528	0.0060
Epifaunal	64	0.0102	Epifaunal	64	0.0799	0.0184
Aggregate	138	0.0032	Aggregate	138	0.0280	0.0073

GULL ROCK MEMBER

(Section V)

Phygraea tarda horizon

BIVALVIA	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>
<i>Phygraea tarda</i> (Hutton)	129	48	2	--	179
(%)	(72.07	(26.82)	(1.12)	--	

ALIA

Brachiopoda, sponge spicules.

DIVERSITY INDEX $D_s = D_g = 0.0$

ARTICULATION RATIO: 0.0001

DIFFERENTIAL TRANSPORT INDEX: 0.5300

EPIFAUNAL BIVALVIA: 100%

(Section Y)

GASTROPODA	<u>no. specs</u>	<u>%</u>
<i>Marginella (Plicaginella) aldingae</i> Tate	8	0.12
<i>Marginella</i> sp. ind	63	0.98
<i>Marginella</i> sp. juvs.	53	0.83
<i>Cottonella</i> gen.nov.mala (Cotton)	3	0.05
<i>Siliquaria kaurna</i> sp. nov.	3	0.05
<i>Leucorhynchia bifuniculata</i> sp. nov.	22	0.34
<i>Praehyalocylis annulata</i> (Tate)	20	0.31
<i>Kleinacteon dubius</i> sp. nov.	65	1.01
<i>Kaurnaacteon</i> gen.nov.elevatus sp.nov.	30	0.47
<i>Acteocina scalarum</i> sp. nov.	4	0.06
<i>Tenuiacteon acicularis</i> sp. nov.	1	0.02
<i>Baryspira (Gracilispira) ligata</i> (Tate)	41	0.64
<i>Jetwoodsia nullarborica</i> (Chapman & Crespin)	3	0.05
<i>Calliostoma (Fautor) cf. allasinazi</i> sp. nov.	7	0.11
<i>Basilissa (Basilissa) cossmanni</i> Tate juv.	13	0.20
<i>Pseudomalaxis (Pseudomalaxis) asculpturatus</i> Maxwell	3	0.05
<i>Turbonilla kaurna</i> sp. nov.	2	0.03
<i>Chemnitzia</i> sp. nov. B.	2	0.03
<i>Chemnitzia</i> sp. nov. C.	1	0.02
<i>Syrnola (Pachysyrnola) habei</i> sp. nov.	2	0.03
<i>Tiberia (Cossmannica) maxwelli</i> sp. nov.	8	0.16
<i>Turbonilla rossiae</i> sp. nov.	1	0.02
<i>Turbonilla</i> sp. nov.	1	0.02
<i>Nobolira costata</i> sp. nov.	1	0.02
<i>Merelina kaurna</i> sp. nov.	6	0.09
<i>Retusa (Decorifer) crassa</i> sp. nov.	80	1.25
<i>R. (Decorifer) gracilis</i> sp. nov.	1	0.02
<i>Kosugeia</i> gen. nov. sp. nov.	1	0.02
<i>Inella maxwelli</i> sp. nov.	3	0.05

	<u>no. specs</u>	<u>%</u>
<i>Triphora</i> s.l. <i>muna</i> sp. nov.	2	0.03
<i>Triphora</i> (<i>Isotriphora</i>) sp. nov.	1	0.02
<i>Triphora</i> s.l. sp. nov. A	1	0.02
<i>Aclis</i> (<i>Graphis</i>) <i>costata</i> sp. nov.	14	0.22
<i>Aclis</i> (<i>Graphis</i>) <i>laevigata</i> sp. nov.	2	0.03
<i>Notovoluta</i> <i>pagodooides</i> (Tate)	1	0.02
<i>Calyptraea</i> sp.	1	0.02
<i>Sigmesalia</i> <i>stylacris</i> (Tate)	1	0.02
<i>Vexinia</i> <i>callosa</i> sp. nov.	11	0.17
<i>Matilda</i> (<i>Opimilda</i>) sp. nov. A	3	0.05
<i>Cerithiopsis</i> (<i>Socienna</i>) sp. nov. A	1	0.02
<i>Seila</i> s.l. sp. nov. A	11	0.17
<i>Cerithiopsis</i> s.l. gen A sp. A	4	0.06
? <i>Eumetula</i> s.l. sp.	1	0.02
<i>Cerithiopsis</i> s.l. gen B sp. A	2	0.03
<i>Cerithiopsis</i> s.l. gen. C sp.	3	0.05
<i>Seila</i> s. str. sp.	6	0.09
<i>Cerithiopsis</i> s.l. gen. D sp.	7	0.11
<i>Cerithiopsis</i> s.l. gen. E sp.	5	0.08
<i>Cerithiopsis</i> s.l. gen. F sp.	12	0.19
<i>Cerithiopsis</i> s.l. gen. G sp.	1	0.02
<i>Cerithiopsis</i> s.l. gen. H sp.	6	0.09
<i>Cerithiopsis</i> s.l. gen. I sp.	2	0.03
<i>Cerithiopsis</i> s.l. gen. K sp.	1	0.02
<i>Cerithiopsis</i> s.l. gen. J. sp.	2	0.03
<i>Cerithiopsis</i> s.l. gen. L sp.	3	0.05
<i>Cerithiopsis</i> s.l. gen. M sp.	5	0.08
<i>Cerithiopsis</i> s.l. gen. N sp.	6	0.09
<i>Cerithiopsis</i> s.l. gen. O sp.	2	0.03
<i>Cerithiopsis</i> s.l. gen. P sp.	1	0.02

<i>Cerithiopsis</i> s.l. gen. Q sp.	3	0.05
<i>Cerithiopsis</i> s.l. gen. R sp.	2	0.03
<i>Cerithiopsis</i> ss.ll. gen. & sp. ind.	36	0.56
? <i>Cerithiopsilla</i> sp. nov.	5	0.08
<i>Cerithiopsis</i> s.l. gen. S sp.	3	0.05
<i>Scala</i> s.l. gen A sp.	1	0.02
<i>Scala</i> s.l. gen. B sp.	4	0.06
<i>Gastropoda opercula</i> gen & sp. ind.	2	0.03
<i>Margarites (Periaulax) rhysus</i> sp. nov. (ex Tate)	8	0.12
<i>Cerithium</i> s.l. sp.	1	0.02
<i>Acteon</i> ss. ll. sp. & gen. ind.	1	0.02
<i>Vexithara citharelloides</i> (Tate)	3	0.05
<i>Austromitra pumila</i> (Tate)	3	0.05
<i>Narona (Inglisella) Turriculata</i> (Tate)	3	0.05
<i>Comitas (Comitas) aldingensis</i> Powell	5	0.08
<i>Charonia (Austrosassia) cribrosa</i> (Tate)	1	0.02
<i>Comitas (Comitas) sp. nov.</i>	5	0.08
<i>Fusinus (Acrocolus) apiciliratus</i> (Tate)	3	0.05
<i>Trophon (Trophonopsis) hypsellus</i> (Tate) juv.	96	1.50
<i>Comitas</i> s.l. sp.	1	0.02
<i>Distorsio (Personella) maslinensis</i> sp. nov.	5	0.08
? <i>Tectifusus</i> sp.	1	0.02
? <i>Columbarium</i> sp. juv.	1	0.02
<i>Zeacolpus</i> sp. (53 ads + 60 juvs.)	113	1.76
<i>Gastropoda</i> sp. ind.	114	1.78
<i>Leiostraca</i> sp. ind.	10	0.16
<i>Eulima</i> s.l. sp. ind.	8	0.12
<i>Eulima</i> s.l. sp. nov.	16	0.25
<i>Odostomia (Auristomia) sulcata</i> sp. nov.	2	0.03
<i>Odostomia</i> s.l. spp. ind.	5	0.08
<i>Balcis</i> sp. nov. A	23	0.36

<i>Balcis</i> sp. nov. B	14	0.22
? <i>Leiostraca</i> sp. nov.	1	0.02
? <i>Chileutomia</i> sp. nov.	10	0.16
<i>Spirocolpus aldingae</i> (Tate) (2.630 juvs + 871 ads)	3.503	54.61
<i>Polinices (Polinices) nothos</i> sp. nov.	20	0.31
<i>Lunatia aldingensis</i> (Tate)	66	1.40
<i>Natica</i> ss.ll. sp.ind.	7	0.15
<i>Tanea falsa</i> sp. nov.	36	0.56

SCAPHOPODA

<i>Gadilina tatei</i> Sharp & Pilsbry	6	0.09
<i>Siphonodentalium</i> sp. nov.	183	2.85
<i>Cadulus</i> sp. nov.	164	2.56
<i>Fissidentalium</i> sp.	1	0.02

BIVALVIA

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs</u>	<u>%</u>
<i>C. (Chlamys) peroni</i> (Tate)				24	24	0.37
<i>C. (Chlamys) flindersi</i> (Tate)				30	30	0.47
<i>Chlamys</i> sp.				4	4	0.06
<i>Lentipecten</i> sp.				1	1	0.02
<i>Poroleda</i> sp.	12	3	13		28	0.44
<i>Dimya asseretoi</i> sp. nov.	276	90	3	1	370	5.77
<i>Corbula (Caryocorbula) pyxidiata</i> (Tate)	1				1	0.02
<i>Hiatella (Hiatella) ?vera</i> (Deshayes)	1				1	0.02
<i>Cuspidaria</i> sp.	1				1	0.02
<i>Cyclopecten</i> s.l. sp. nov.	17	7	1		25	0.39
<i>Propeamussium (Parvamussium) sp. nov.</i>	8	4			12	0.19
<i>Allasinazella gen.nov. equidens</i> (Tate)	3				3	0.05
<i>Limopsis zitteli</i> Ihering	10	4		3	17	0.27
<i>Notogrammatodon inexpectatus</i> Maxwell	4	6		2	12	0.19
<i>Arcopsis (Arcopsis) dissimilis</i> (Tate)	17	4		1	22	0.34
<i>Modiolus</i> ss.ll. sp. juv.	1				1	0.02
? <i>Lissarca</i> sp.				25	25	0.39

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs</u>	<u>%</u>
<i>Salaputium aldingensis</i> Finlay	8	7		14	29	0.45
<i>Salaputium lamellatum</i> (Tate)	6	4			10	0.16
<i>Condylocardia radiata</i> (Tate)			5	45	50	0.78
<i>Cardita</i> s.l. sp.	13	23	7	11	54	0.84
<i>Pectunculina cancellata</i> sp. nov.	134	98	8	16	256	3.99
<i>Pronucula</i> (<i>Pronucula</i>) <i>tatei</i> (Finlay)	5	2	4	1	12	0.19
<i>Nuculana</i> (<i>Ledella</i>) <i>leptorhyncha</i> (Tate)			132	193	325	5.07
<i>Dosina</i> (<i>Dosina</i>) <i>multilamellata</i> (Tate)			12	7	19	0.30

ALIA

Bryozoa, shark teeth, fish bones, otoliths, Echinoids, Brachiopoda, ahermatypic corals, Anellida, Decapoda (Crabs' claws), sponge spicules.

DIVERSITY INDEXES		<u>Gastropoda</u>		<u>Scaphopoda</u>		<u>Bivalvia</u>		<u>Total</u>	
		no.	%	no.	%	no.	%		
N = 6414	N _s = 127	Specimens	4.728	73.53	354	5.52	1.332	20.81	6414
S _s = 127	S _g = 107	Species	98	77.16	4	3.15	25	19.69	127
D _s = .6916	D _g = .9964	Genera	82	75.93	4	3.70	22	20.37	108

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	BIVALVIA	%
Epifaunal	327	111	4	63	505	Epifaunal	37.91
Infaunal	190	141	181	315	827	Infaunal	62.09
Uncertain	-	-	-	-	-	Uncertain	-

Total	517	252	185	378	1.332		
(%)	(38.81)	(18.92)	(13.89)	(28.38)			

ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES			
	<u>specs</u>	<u>index</u>		<u>specs</u>	<u>index</u>	<u>uncertainty index</u>
Epifaunal	501	0.00002	Epifaunal	501	0.3055	0.0057
Infaunal	646	0.04250	Infaunal	646	0.0439	0.0965
Aggregate	1.147	0.01350	Aggregate	1.147	0.0721	0.0317

'SOFT MARL' MEMBER

(Section Y)

Nautilus Horizon

GASTROPODA	<u>no specs.</u>	<u>%</u>
<i>Praehyalocylis annulata</i> (Tate)	15	0.59
<i>Styliola</i> sp. nov.	1	0.04
<i>Natica</i> sp. (opercula)	1	0.04
<i>Spirocolpus aldingae</i> (Tate)	122	4.83
<i>Zeacolpus</i> sp.	169	6.69
<i>Lunatia aldingensis</i> (Tate)	11	0.44
<i>Jetwoodsia nullarborica</i> (Chapman & Crespin)	25	0.99
<i>Baryspira</i> (<i>Gracilispira</i>) <i>ligata</i> (Tate)	1	0.04
<i>Tanea falsa</i> sp. nov.	22	0.87
<i>Polinices</i> (<i>Polinices</i>) <i>nothos</i> sp. nov.	38	1.50
<i>Retusa</i> (<i>Decorifer</i>) <i>crassa</i> sp. nov.	31	1.23
<i>Margarites</i> (<i>Periaulax</i>) <i>rhysus</i> sp. nov. (ex Tate)	42	1.66
<i>Paradilhia</i> (<i>Micropyrgos</i>) sp. nov.	3	0.12
<i>Marginella</i> (<i>Plicaginella</i>) <i>aldingae</i> (Tate)	26	1.03
<i>Pterynotus</i> (<i>Pterochelus</i>) <i>manubriatus</i> (Tate)	1	0.04
<i>Limacina</i> sp. nov. A	42	1.66
<i>Aclis</i> (<i>Graphis</i>) <i>costata</i> sp. nov.	2	0.08
<i>Austromitra pumila</i> (Tate)	18	0.71
<i>Ataxocerithium concatenatum</i> (Tate)	1	0.04
<i>Vexithara citharelloides</i> (Tate)	7	0.28
<i>Trophon</i> (<i>Trophonopsis</i>) <i>hypsellus</i> (Tate)	40	1.58
<i>Knefastia</i> sp. nov. B	40	1.58
<i>Kleinacteon dubius</i>	34	1.35
? <i>Miralda</i> s.l. sp. nov.	1	0.04
<i>Acteon</i> ss.ll. sp. ind.	7	0.28
<i>Cylichna</i> (<i>Cylichnania</i>) <i>callosa</i> (Tate)	6	0.24
<i>Kaurnacteon</i> gen.nov. <i>elevatus</i> sp. nov.	1	0.04

<i>Acteocina scalarum</i> sp. nov.	1	0.04
<i>Chrysallida</i> sp. nov.	17	0.67
<i>Turbonilla kaurma</i> sp. nov.	3	0.02
<i>Chemnitzia</i> sp. nov. B	1	0.04
<i>Turbonilla rossiae</i> sp. nov.	8	0.32
<i>Tiberia (Cossmannica) maxwelli</i> sp. nov.	2	0.08
<i>Eulima</i> s.l. sp. nov.	23	0.91
<i>Odostomia (Auristomia) sulcata</i> sp. nov.	11	0.44
<i>Leiostraca</i> sp. nov.	8	0.32
? <i>Leiostraca</i> sp.	4	0.16
? <i>Nototrivia</i> sp.	1	0.04
<i>Marginella</i> s.l. spp. ind. juvs	16	0.63
<i>Cassoginella palla</i> (Cotton)	3	0.12
<i>Kaurmaginella</i> gen. nov. <i>tutugka</i> sp. nov.	5	0.20
<i>Alaginella submicula</i> sp. nov.	2	0.08
<i>Archierato pyrulata</i> (Tate)	8	0.32
<i>Marginella</i> s.l. spp. ind.	23	0.91
<i>Cottonella</i> cf. <i>mala</i> (Cotton)	2	0.08
<i>Leucorhynchia bifuniculata</i> sp. nov.	1	0.04
<i>Turbo</i> ss.ll. sp. ind.	1	0.04
<i>Cirsotrema mariae</i> (Tate)	4	0.16
<i>Columbarium</i> sp.	1	0.04
<i>Matilda (Opimilda)</i> sp. nov. A	4	0.16
<i>Seila</i> s.l. sp. nov.	7	0.28
<i>Murexul prionotus</i> (Tate)	3	0.12
<i>Fusinus (Microcolus) apiciliratus</i> (Tate)	1	0.04
<i>Latirus (Brocchitas) altior</i> sp. nov.	1	0.04
<i>Trophon</i> s.l. sp.	1	0.04
<i>Comitas (Comitas) aldingensis</i> Powell	17	0.67
<i>Comitas</i> s.l. sp. nov. B	12	0.48
<i>Fusinus (Fusinus) sculptilis</i> (Tate)	3	0.12

(48)

<i>Inella maxwelli</i> sp. nov.	1	0.04
<i>Triphora</i> s.l. <i>muna</i> sp. nov.	2	0.08
<i>Argobuccinum</i> (<i>Cymatiella</i>) <i>oligostira</i> (Tate)	9	0.36
<i>Narona</i> (<i>Inglisella</i>) <i>turriculata</i> (Tate)	2	0.08
<i>Calliostoma</i> (<i>Fautor</i>) <i>allasinazi</i> sp. nov.	2	0.08
<i>Sirius</i> cf. <i>tabulatus</i> (Tate)	13	0.51
<i>Trophon</i> (<i>Enantimene</i>) <i>monotropis</i> (Tate)	3	0.12
<i>Fusus</i> s.l. sp. A juv.	10	0.40
<i>Fusus</i> s.l. sp. B juv.	3	0.12
Gastropoda spp. ind. & juvs.	75	2.97
<i>Cerithiopsis</i> s.l. gen. Q sp. nov.	3	0.12
<i>Cerithiopsis</i> s.l. gen. A sp. nov. A	9	0.36
<i>Cerithiopsis</i> s.l. gen. C sp. nov.	1	0.04
<i>Cerithiopsis</i> s.l. gen. H sp. nov.	10	0.40
<i>Cerithiopsis</i> s.l. gen. J sp. nov.	1	0.04
<i>Cerithiopsis</i> s.l. gen. M sp. nov.	14	0.55
<i>Cerithiopsis</i> s.l. gen. D sp. nov.	3	0.12

SCAPHOPODA

<i>Siphonodentalium</i> sp. nov.	66	2.61
<i>Laevidentalium</i> sp. nov.	22	0.87
<i>Plagioglypta</i> sp. nov.	4	0.16
<i>Gadilina tatei</i> Pilsbry & Sharp	138	5.46

BIVALVIA

	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>tot.no.specs</u>	<u>%</u>
<i>Notogrammatodon inexpectatus</i> Maxwell	7	1			8	0.32
<i>Bivalvia</i> spp. ind.				6	6	0.24
<i>Divarilima</i> cf. <i>polyactina</i> (Tate)	5	1			6	0.24
<i>Corbula</i> sp.				1	1	0.04
<i>Lissarca</i> sp.	3	4			7	0.28
<i>Hiatella</i> (<i>Hiatella</i>) ? <i>vera</i> (Deshayes)	2	1			3	0.12
<i>Arcopsis</i> (<i>Arcopsis</i>) <i>dissimilis</i> (Tate)	7	7	1		15	0.59

<i>Salaputium lamellatum</i> (Tate)	16	27	1	4	48	1.90
<i>Condylocardia radiata</i> (Tate)	20	17	1		38	1.50
<i>Glycymeris lenticularis</i> (Tate)	2	2		5	9	0.36
<i>Dosina (Dosina) multilamellata</i> (Tate)	2	2		13	17	0.67
<i>Propeamussium (Parvamussium) sp. nov.</i>	63	84			147	5.82
<i>Dimya asseretoi sp. nov.</i>	88	55			143	5.66
<i>Salaputium aldingensis</i> Finlay	16	16		2	34	1.35
<i>Pectunculina cancellata sp. nov.</i>	98	80	7	11	196	7.76
<i>Nuculana (Ledella) leptorhyncha</i> (Tate)	116	103	44	5	268	10.61
<i>Pronucula (Pronucula) tatei</i> (Finlay)	5	9		3	17	0.67
<i>Nuculana (Poroleda) sp. nov.</i>	2	4	3		9	0.36
<i>Solamen (Exosiperma) cf. globularis</i> (Tate)				1	1	0.04
<i>Sarepta planiuscula</i> (Tate)	2	1		4	7	0.28
<i>Nuculana (Saccella) chapmani</i> Finlay	15	17	3	11	46	1.82
<i>Limopsis zitteli</i> Ihering juv.	9	4	1	2	16	0.63
<i>Cardium</i> ss. 11. sp.	38	23	1	2	64	2.53
? <i>Eotrigonia sp. ind. juv.</i>			1		1	0.04
<i>Nemocardium sp. ind.</i>	2	7		89	98	3.88

ALIA

Ahermatypic corals (very diversified), shark teeth, otoliths, fish bones and vertebrae. Echinoids, Anellida, sponge spicules, Brachiopoda.

DIVERSITY INDEXES		Gastropoda		Scaphopoda		Bivalvia		Total	
N	N _s	no.	%	no.	%	no.	%		
N = 2526	N _s = 104	Specimens	1.901	43.19	230	9.30	1.205	47.71	2.526
S _s = 104	S _g = 93	Species	75	72.12	4	3.85	25	24.04	104
D _s = .9591	D _g = .9976	Genera	67	72.04	4	4.30	22	23.66	93

VALVES	<u>LV</u>	<u>RV</u>	<u>BV</u>	<u>vv</u>	<u>total</u>	BIVALVIA	<u>%</u>
Epifaunal	170	148	1	1	320	Epifaunal	26.56
Infaunal	348	318	61	152	879	Infaunal	72.95
Uncertain	-	-	-	6	6	uncertain	.50
<hr/>							
Total	518	466	62	159	1.205		
(%)	(42.99)	(38.67)	(5.15)	(13.20)			
ARTICULATION RATIOS			DIFFERENTIAL TRANSPORT INDEXES				
	<u>specs</u>	<u>index</u>		<u>specs</u>	<u>index</u>	<u>uncertainty</u>	<u>index</u>
Epifaunal	319	0.0000	Epifaunal	319	0.1150	0.0000	
Infaunal	818	0.0029	Infaunal	818	0.0381	0.0124	
Aggregate	1.143	0.0015	Aggregate	1.143	0.0284	0.0064	

A P P E N D I X D

1. - Late discovered species of stratigraphic significance
2. - Notes on the Australian Eocene Nautiloidea

D - 1. LATE DISCOVERED SPECIES OF STRATIGRAPHIC SIGNIFICANCE

An ongoing project on the Late Eocene Molluscan faunas from the Buccleuch Beds (Padthaway Ridge, Murray Basin), begun in the first half of 1978 on behalf of the S.A. Department of Mines & Energy, resulted in the discovery of a number of micromolluscan genera which have different representatives in the Blanche Point Formation under the Adelaide City area (St. Vincent Basin) and in the Buccleuch Beds of the Padthaway Ridge. Since this discovery has an obvious and vital significance in the construction of a biostratigraphical framework for the Late Eocene of southern Australia, it was felt that the information derived from these forms ought to be included in this study. However, at that time, writing up of this thesis was already in its final stages. It was therefore considered more appropriate to deal with these meaningful species in an appendix, informally, and to limit their discussion to the morphological differences between the older Blanche Point and the younger Buccleuch forms and to their stratigraphic occurrences.

FAMILY SCISSURELLIDAE Gray, 1847
GENUS Scissurella d'Orbigny, 1824
SUBGENUS Scissurella s. str.

OBSERVATIONS. Scissurella d'Orbigny was recorded in the Blanche Point Formation under the Adelaide City area (Adelaide Plains SubBasin) with Scissurella sp. nov. and in the lower Port Willunga Formation, Aldinga Member, and in the Buccleuch Beds with S.lamellularum sp. nov.

Scissurella sp. nov. is represented by a few juveniles, but differences in protoconch coiling and ornament suggest a different species from S. lamellularum.

DISTRIBUTION. Scissurella sp. nov. St. Vincent Basin, Adelaide Plains SubBasin, South Parklands CH-1A, 99-97 m, 85-83 m, 48.3-48 m.
Scissurella lamellularum See description of the species.

FAMILY CYCLOSTREMATIDAE Fischer, 1885

SUBFAMILY CYCLOSTREMATINAE Fischer, 1885

GENUS Circulus Jeffreys, 1865

OBSERVATIONS. This genus is represented in the Late Eocene of South Australia by three forms: the early-middle P16 Circulus sp. nov. A, the P17 Circulus sp. nov. B and the late P17 Circulus sp. nov. C. Circulus sp. nov. A and sp. nov. B are very close to each other. The former differs from the latter only in smaller protoconch and size (in relation to the whorl number), in rounder abapical-adaxial connection and in even spiral costellae, present also in the abapical margin; whereas, Circulus sp. nov. B has subangular adaxial-abapical connection, weaker abapical costellae, and two well interspaced and marked costae at the periphery.

Circulus sp. nov. C, co-occurring in the upper Buccleuch 'B'

Beds with Circulus sp. nov. B is distinguished by the heavier spiral pattern (abaxial-adapical carina, abaxial spiral costae, weak abapical costellae) and by a rounded abapical-adaxial connection.

Circulus spp. nov. A and B appear to be related to the Palaeocene Anglo-Paris Basin Circulus tenuiliratus (Cossmann) (Glibert, 1973, p.29, pl.4, fig.7) because of their similar ornament pattern. On the other hand, Circulus sp. nov. C, although closer in umbilical morphology to the other two forms, shows greater affinity with the Palaeocene Anglo-Paris Basin C. montensis (Rutot, in Cossmann) (Glibert, loc.cit., fig.4), which however displays a very angular abapical-adaxial connection.

The Pliocene Partubiola depressispira Ludbrook displays a congeneric similarity in protoconch and in ornament pattern with Circulus sp. nov. C, and because of its umbilical morphology, a closer affinity with C. montensis. Therefore, it should be referred to Circulus Jeffreys. The shell similarity to the above Paleogene species of the extant Partubiola blancha Iredale, type of Partubiola Iredale, and of the other Pliocene species, P. varilirata Ludbrook, strongly suggests a possible congenericity

of Partubiola with Circulus.

DISTRIBUTION.

Circulus sp. nov. A: Adelaide Plains SubBasin, South Parkland Bore CH-1A.
48.30-48.00 (15 specimens) (Transitional Marls).

Circulus sp. nov. B: Tintinara Area School Bore 12, 88-86 m (1 spec.),
86-84 m (4 specs.), 84-82.50 (2 specs.), 81.50-
80.00 m (4 specs.) depths.

Circulus sp. nov. C: Tintinara Area School Bore 12, 81.50-80.00 m depth
(1 spec.)

GENUS Brookula Iredale, 1912

SUBGENUS Brookula s. str.

OBSERVATIONS. Brookula is represented in the Late Eocene by two species: Brookula sp. nov. A from the Blanche Point Formation and Brookula sp. nov. B from the Buccleuch Beds.

Brookula sp. nov. A differs from Brookula sp. nov. B in broader umbilicus, shorter spire, and more interspaced axial costae. These two species represent the oldest occurrence of the genus in Australasia. The oldest forms hitherto known are the New Zealand Early-Middle Miocene B. endodonta Finlay and the late Early Miocene B. pukeuriensis Finlay.

DISTRIBUTION.

Brookula sp. nov. A: St. Vincent Basin, Adelaide Plains SubBasin,
South Parkland Bore CH-1 A, 35-35.3 m.

Brookula sp. nov. B: Tintinara Area School Bore, 12, 82.50-84.00 m,
80.00-81.50 m.

FAMILY CAECIDAE Gray, 1847

GENUS Strebloceras Carpenter, 1858

OBSERVATIONS. This genus is represented in the Late Eocene by two

forms: Strebloceras sp. nov. from the Blanche Point Formation ('Transitional Marls') (Adelaide Plains SubBasin, St. Vincent Basin), and the already described Strebloceras darraghi sp. nov. from the Buccleuch Beds (Murray Basin) and the Aldinga Member, Port Willunga Formation (St. Vincent Basin).

S. darraghi differs from the older form in higher spire and deeper and narrower umbilicus. These two species represent the oldest record of the genus in Australasia. The only other fossil form known is the New Zealand Early Miocene S. hinemoa Finlay.

DISTRIBUTION.

Strebloceras sp. nov.: St. Vincent Basin, Adelaide Plains SubBasin,
West End Brewery CH-2 50.3-50.00 m (2 specs.),
South Parkland CH-1A, sludges, 44-42 m (12 specs.);
62-60 m, (1 spec.).

Strebloceras darraghi sp. nov.: See description.

FAMILY VITRINELLIDAE
GENUS Vitrinella C.B. Adams, 1850
Vitrinella ss. 11.

OBSERVATIONS. Two Late Eocene forms are referable to Vitrinella ss. 11. the Blanche Point Formation Vitrinella sp. nov. A and the Buccleuch Beds Vitrinella sp. nov. B.

Vitrinella sp. nov. A displays very high lamellose axial costae with narrow interspaces. Vitrinella sp. nov. B has prominent but short axial costae, subdued on the abaxial and, perhaps, larger in size. Both these forms display similar protoconch: one-whorled, heterostrophic-hyperstrophic, separated from the teleoconch by a suture.

The general shell morphology recollects those of: Munditiella Kuroda & Habe (Skeneinae), though the latter has a higher spire; Pondorbis Bartsch (Skeneinae), close also in flexuous costae; and Liotella Iredale (Cyclostrematinae), particularly the extant Liotella annulata (T. Woods) (Cotton, 1959, p.243, fig. 167). The hyperstrophic protoconch, however,

is quite similar to those of Orbitestella Iredale and of the ?Elachorbis plicatella group. This would place these two forms in either Vitrinellidae or Orbitestellidae. At present, they are tentatively located in Vitrinella s.l.

DISTRIBUTION.

Vitrinella s.l. sp. nov. A: Adelaide Plains SubBasin, South Parklands Bore Bore CH-1A, sludges, 89-87 m and 77-75 m depths.

Vitrinella s.l. sp. nov. B: Murray Basin, B.Q. Butler Bore 4, Hd Kirkpatrick, Sect.8, 125-122.05 m (1 spec.), 125-118.77 m (1 spec.); Kiki Town Bore, Buccleuch Co., 103, 63-116.14 m (6 specs); Coonalpyn E&W Bore 2, Hd Coneybeer, Sect.56, 77.10-76.12 m (1 spec), 72.83-71.52 m (1 spec). Tintinara Area School Bore 12, Co Cardwell, Hd Coombe, 88-86 m (9 spec.), 86-84 m (6 spec.) 84-82.50 m (12 specs), 81.50-80 m (3 specs), 80-78 m (1 spec).

FAMILY

ARCHITECTONICIDAE

GENUS

Pseudomalaxis Fischer, 1885

SUBGENUS

Pseudomalaxis s. str.

Pseudomalaxis (Pseudomalaxis) sp. nov. aff. asculpturatus Maxwell.

FIGS. 372-375

MATERIAL. 6 specimens, partly damaged.

OBSERVATIONS. In comparison with the P15/16 P. ludbrookae and with P. asculpturatus Maxwell, this form appears to be closer to the latter.

It is similar to P. asculpturatus in plain and very concave abapical region and flat abaxial margin. On the other hand, it differs in more marked and faintly crenulated carinae, in convex adapical margin and presence of a faint abaxial adapical spiral groove. This combination of crenulated carinae, convex adapical margin, and abaxial adapical spiral groove may indicate a development of characters precursory to the Neogene forms of Pseudomalaxis (see Buonaiuto, 1975, p.25, figs. 4a,b).

DISTRIBUTION. Murray Basin, Tintinara Area School Bore 12, 80-81.50 m and 78-80 m depths.

D - 2 NOTES ON THE AUSTRALIAN EOCENE NAUTILOIDEA

Although they are not dealt with in this study, the Australian Eocene Nautiloidea cannot be totally ignored because of their significance as palaeoclimatic indicators. However, since the possibility of post-mortem transport of Nautiloid shells over very long distances by tides and oceanic currents has been proven (Furnish & Glenister, 1964; Teichert, 1964; Stenzel, 1964), the validity of their use in the reconstruction of palaeoclimates in southern Australia rests entirely upon the demonstrability of their autochthony to this region. Yet this problem cannot properly be discussed without considering the known record of the Nautiloidea in the Australian Eocene.

Biostratigraphical observations

Only four genera and species have hitherto been recorded in the Australian Middle and Late Eocene: Deltoidonautilus prora (Glenister, Miller & Furnish), Aturoidea brunnschweileri Glennister, Miller & Furnish, Aturia clarkei Teichert, and Cimomia felix (Chapman). All these four genera have a similar Tethyan-Atlantic-Eastern Pacific-Australian distribution (Sastry & Mathur, 1968; Kummel, Furnish & Glenister, 1964; Ruzhentsev et al., 1962). The Late Jurassic-Oligocene Cimomia Conrad seems to have initially appeared in the Australian record in the Late Cretaceous (Glenister, Miller & Furnish, 1956) and disappeared by the end of the Late Eocene (McGowran, 1959; this study). The Late Cretaceous-Oligocene Deltoidonautilus Spath and Aturoidea Vredenburg initially appeared in the Middle Eocene and Paleocene, respectively, and both disappeared sometime during the Middle/Late Eocene (loc. cit.; Teichert, 1943). The Paleocene-Miocene Aturia Brönn represents the most recent 'immigrant' of the four, since its record is in the Middle to Late Eocene and its last record is from the Middle Miocene locality of Muddy Creek (Cockbain, 1968a; Glaessner, 1955).

The stratigraphic and geographic distributions of the nautiloid species present in the Australian Eocene are shown in the table below.

BASIN	STRATIGRAPHIC LOCATION	AGE	AUTHORS
	<i>Deltoïdonautilus prora</i>		
Carnarvon	lower Giralia Calcarenite	Middle to early Late Eocene	Glenister, Miller & Furnish, 1956
	Jubilee Calcarenite	"	"
Norseman-South Coast	Plantagenet Group	early Late Eocene	Glenister & Glover, 1958
	<i>Aturoidea brunnschweileri</i>		
Carnarvon	lower Giralia Calcarenite	Middle to early Late Eocene	" "
	Jubilee Calcarenite	"	" "
	<i>Aturia clarkei</i>		
Carnarvon	lower Giralia Calcarenite	Middle to early Late Eocene	as above; Cockbain, 1968a.
	Merlinleigh Sandstone	Middle to Late Eocene ⁽⁰⁾	Cockbain, 1968a; ⁽⁰⁾ Quilty, 1974
Norseman-South Coast	Pallinup Siltstone	early Late Eocene	Cockbain, 1968a
St. Vincent Basin	lower and middle Blanche Point Formation	early-mid. Late Eocene	Glaessner, 1955; Cockbain, 1968a
	Tortachilla Limestone	early Late Eocene	as above
Otway	Browns Creek Formation	early-mid. Late Eocene	O.P. Singleton, 1967
	Clifton Formation (reworked specimen)	Lower Miocene	Glaessner, 1955; Cockbain, 1968a.
	<i>Cimomia felix</i>		
Norseman-South Coast	Plantagenet Group	early Late Eocene	Cockbain, 1968b
St. Vincent (Yorke Peninsula)	Muloowurtie Formation (?)	Late Eocene	McGowran, 1959
St. Vincent (Eastern part)	Blanche Point Formation	early-mid. Late Eocene	McGowran, 1959; this study
	Tortachilla Limestone	early Late Eocene	as above
Otway	Browns Creek Formation (above, <i>Phygraea</i> horizon and below it within <i>Hantkenina</i> zone)	middle Late Eocene	McGowran, 1959; this study

A fifth nautiloid genus, the Upper Jurassic-Miocene Eutrephoceras Hyatt, is known in the Australian Early Cretaceous-Tertiary (Ludbrook, 1966; McGowran, 1959) but its Tertiary occurrence is restricted to a Paleocene (1 species) and a Late Oligocene-Middle Miocene incursion (3 species) (Teichert, 1943; McGowran, 1959; Darragh, 1970).

In particular, a more detailed nautiloid biostratigraphy can be drawn for the Eastern St. Vincent Basin. Field observations and the examination of the matrix of the specimens which are kept in collections of the S.A. Museum and of the Geology Department, University of Adelaide, revealed that: -

- Aturia clarkei and Cimomia felix are frequent and co-occur in the Tortachilla Limestone s. str., in the 'Transitional Marl' Member where they show an acme and in the middle and upper Gull Rock Member.
- they are not recorded in the Phygraea tarda and Bryozoa horizons (lower Gull Rock Member);
- in the 'Soft Marl' Member A. clarkei is not found, but C. felix still occurs in patches of individuals clustered together.

Paleoclimatological observations

The paleobiogeographic distribution of the four genera, their biostratigraphical occurrences linked with the Australian palaeoclimatic curves, suggest that: (a) Aturia and Cimomia were able to live in both tropical and subtropical waters, with Cimomia showing a higher eurythermality; (b) Deltoidonautilus and Aturoidea restricted to tropical waters, with Aturoidea showing the higher stenothermality, since their first occurrences in Australia appears to be linked with the climatic optima of the Paleocene and the Middle Eocene, as suggested by the brief appearance of Aturoidea and Eutrephoceras in the Paleocene of the Otway Basin (Teichert, 1943) and by the restricted northern distribution of Aturoidea in the Middle-Late Eocene.

To support the hypothesis of a climatic control in the occurrences

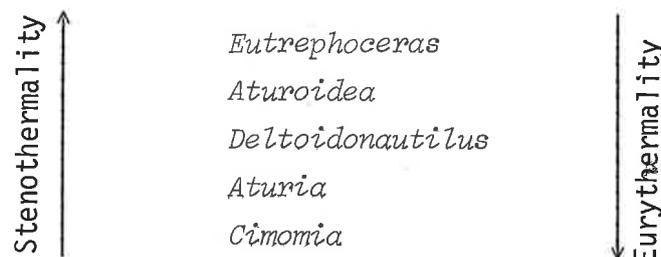
of the Australian Tertiary Nautiloidea, the analysis of the vertical and horizontal distribution of the Eocene species in the western and southern Australian margins revealed the following:

Four distinct nautiloid assemblages can be distinguished in time and place :

- the northern middle to Late Eocene Deltoidonautilus-Aturoidea-Aturia, Carnarvon Basin;
- the south-western early Late Eocene Deltoidonautilus-Aturia-Cimomia Norseman-South Coast;
- the southern early-middle Late Eocene Aturia-Cimomia, St. Vincent, and Otway Basins;
- the southern middle Late Eocene monotypic Cimomia assemblage.

These assemblages indicate a decreasing diversity related to time and latitude. This trend appears consistent with the general climatic deterioration from the middle to the Late Eocene and with the present distribution of Nautilus, the only extant genus today, which shows similar climate controlled diversity gradient. In fact, four of the five living 'species' are distributed in a NW-SE intertropical belt from the Philippines to the Samoa Archipelago; the fifth species, Nautilus repertus Iredale is the only subtropical form and occurs alive off the South Australian Coast (Stenzel, 1964).

Suggested eurythermality and stenothermality of the Australian Tertiary Nautiloidea, as inferred by their paleogeographic and stratigraphic record.



Autochthony of the Australian Eocene Nautiloidea

Reyment (1958), Stenzel (1964), Furnish & Glenister (1964), and Teichert (1964) discuss exhaustively the problems in using the Nautiloidea for palaeoecological and palaeoclimatological interpretations. The main problem in regard to autochthony is the post-mortem drift of nautiloid shell over distances which may extend up to 2.900-7.000 km (Stenzel, 1964; Furnish & Glenister, 1964). The post-mortem drift in nautiloids is controlled by the buoyancy of their shells, which depends in its turn upon several factors: soundness of the shell, wall thickness, size and shape of the body chamber, the presence/absence on the shell of encrusting organisms, the temperature and the salinity of the waters, the hydrodynamic properties of the shell, the rate of shell corrosion in water, etc. (for detailed discussion see Reyment, 1958).

Since all the four genera present in the Eocene have a long, if disjunct, record in the Australian Tertiary, and since all of them are represented by species precinctive to the Australian region, it would be very difficult to deny the possibility of nautiloid populations living at least at drift range from the Australian coasts during that time.

In regard to the possibility of nautiloid populations off the Australian coasts during the Eocene, there are a number of indications pro and against it.

The streamlined shells of Deltoidonautilus and, to a lesser degree, of Aturia favour sinking rather than long distance transport. Stenzel (1935) believed that Aturia in the Texas Eocene was deposited after having drifted, after death, from the deeper parts of the Gulf of Mexico, which does not entail great distances.

The shell shape of Cimomia and Aturoidea is, on the contrary, favourable to buoyancy, and thus to long distance transport. However, the shell drift would more than likely have been restricted to the gyre of the ProtoSouthern Ocean by the South Tasman Rise on the East, as

suggested by the late arrival in the western and southern Australian basins of Aturia, which is already present in the Late Paleocene of New Zealand (Fleming, 1945, 1966). A westward shell drift would also be limited by the Eocene analogue of the Western Australian current (Fig. 19). The Western Australian Current analogue may also have allowed a S-N drift along the Western Australian coasts. The resulting distribution might have been similar to that of the living Nautilus repertus Iredale. A N-S drift would have been possible only by shoreline and tidal currents, which would imply a rather short transport prior to the stranding or the sinking of shells. Another possibility, involving long distance floating, would be shell transport by the analogue of the West Wind Drift. This possibility would locate the possible origin of the two nautiloids off the Antarctic or the South American coasts.

However, there is some evidence against the latter hypothesis. Possible cracks in the shell of Cimomia and Aturoidea, due to faulty mantle secretion (Reyment, 1958), may expedite the decay of the shell conchyolin, thus favouring a rapid perforation of the walls. Analogously, the thinness and fragility of the Aturia shells make them predisposed to organic decay and also to damage by impact (Teichert, 1964). Both factors would cause an early sinking of the shells. The absence of encrusting organisms in most of the specimens observed suggests a short span of time between the death of the individual and the final entombment of its shell, which would again infer the possibility of a short transport only.

The frequency of Cimomia and Aturia in the horizons of Blanche Point Formation in which they are known to occur, might find an explanation in Reyment's 1958 hypothesis of a possible predation on nautiloids which would bring about the immediate sinking of the damaged shell. Although no predation marks have been noticed, several of the observed and figured specimens display a missing or damaged body chamber, prior to fossilization. This, however, may also be due to the dragging of the shell against the

sea bottom after sinking or to wave action and impacts after being stranded (Reyment, 1958).

Finally, the Middle-Late Eocene N-S diversity gradient observed in the Nautiloids would be against long distance transport as a sole explanation to their presence in the Eocene deposits, because it would rather favour a haphazard accumulation of shells of forms characteristic of different climatic belts. In this case, the co-occurrence of 'sinkers', i.e. Aturia and Deltoidonutilus, together with 'floaters', i.e. Cimomia and Aturoidea, indicates a mixed tana-thocoenosis, composed of elements living in deep waters ('floaters') and of elements selective for shallower waters, probably from areas adjacent to their place of entombment.

In conclusion, the hypothesis of autochthony for the Australian Eocene Nautiloidea appears simpler and, therefore, more probable than allochthony.

Finally, taking into account the palaeoenvironmental interpretation of the Eastern St. Vincent Basin, as suggested by the other Molluscan Phyla, the Tortachilla Limestone specimens were probably deposited as a result of shell stranding and those in Blanche Point Formation as a result of shells sinking.

A P P E N D I X E

J.M. Lindsay

B.J. Cooper

(personal communications)

J.M. Lindsay, S.A. Department of Mines & Energy

Personal communication

'One small, broken half-specimen of Hantkenina sp. has been recovered recently from a small amount of fossiliferous limestone matrix in and around a specimen of 'Notostrea lubra' from the Wilson Bluff Limestone, Eucla Basin. Details of the sample are: S.A.D.M.E. Palaeontology Catalogue No. F186/54, collected by I.R. Campbell, S.A.D.M. 1954, from coastal cliffs 12-15 miles (19-24 km) east of Eucla (i.e. 4-7 miles or 6-11 km east of Wilson Bluff, S.A.) at 83 feet (25.3 m) above sea level. Another sample, F185/54 is described as having the same locality and height above sea level, but lacks Hantkenina.

When key parameters are plotted, Hantkenina sp. shows morphological affinities both with H. primitiva from a thin zone in the Late Eocene of the St. Vincent Basin (Zone P15, top), and with H. australis, from basal Wilson Bluff Limestone, Middle Eocene (Zone P12) of the Eucla Basin. However this preliminary examination suggests that the specimen is morphologically closer to H. australis because of its relatively low apertural height ratio and its stout tubulospines.

The associated and rather limited foraminiferal faunas in F185/54, F186/54, lack any species restricted to the Middle Eocene part of the Wilson Bluff Limestone, particularly spinose acarininids such as A. primitiva; but such negative evidence should be treated with caution. Globorotalia (Turborotalia) insolita which is common in both samples might favour a Late Eocene rather than a Middle Eocene age from its presently known range. Other species present such as Globigerinatheka index, Subbotina linaperta, Chiloguembelina cubensis, and the benthonic Maslinella chapmani, do not discriminate between Middle and Late Eocene. Neither Tenuitella aculeata nor Truncorotaloides collactea were found;

which could imply either poor and unsuitable planktonic facies, or a Late Eocene stratigraphic position above the range of Tr. collectea and within the gap in the disjunct range of T. aculeata. Thus, if anything, the foraminiferal fauna associated with Hantkenina sp. favours a Late rather than Middle Eocene age, but the evidence is inconclusive and a Late Eocene age would tend to conflict with the specimen's likeness to H. australis, a species which is typically (but probably not exclusively) Middle Eocene.

Stratigraphically, at 83 feet (25.3 m) above sea level, and with Wilson Bluff Limestone extending below sea level in this area, it is almost certain that Hantkenina sp. is well above the Hantkenina event at the base of Wilson Bluff Limestone (Zone P12). On the other hand the specimen cannot be identified with any certainty with the Hantkenina event at top Zone P15 in the St. Vincent Basin. The individual may represent an intermediate level of Middle to Late Eocene age, with a poorly preserved associated microfauna. Alternatively, Hantkenina sp. could be a peripheral morphotype in a population of H. primitiva of much the same age as in the St. Vincent Basin. Detailed sampling of this section of Wilson Bluff Limestone is needed to resolve the matter.'

J.M. Lindsay
29/3/78

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Personal communication

WILLUNGA EMBAYMENT BOREHOLE WLG 40
(Project QA16)

The succession from 55 m to 74 m in QA16 correlates with the Aldingan unit of the Port Willunga Beds (Lindsay, 1967) and is Late Eocene in age. The topmost beds of the unit (55-56 m) lack Turborotalia aculeata and probably correlate with the Subbotina linaperta Zone of Lindsay (1969) and with the P17 zone of Blow (1969). The remainder of this interval correlates with the T. aculeata Zone of the local zonal scheme and with P16 on the international scale.

B.J. Cooper
8/11/76

A P P E N D I X F

PUBLISHED PAPERS

- 1975 - Notes on the genus *Pseudomalaxis* (Mollusca: Gastropoda) and its fossil species in Australia. Trans. R. Soc. S. Aust., 99 (1), pp.21-30.
- 1977a - Revision of the Australian Tertiary species ascribed to *Limatula* Wood (Mollusca: Bivalvia). Trans. R. Soc. S. Aust., 101 (1), pp. 21-33.
- 1977b - Revision of the composite species *Lima bassi* Tenison Woods (Mollusca: Bivalvia). Trans. R. Soc. S. Aust., 101 (3), pp. 75-83.

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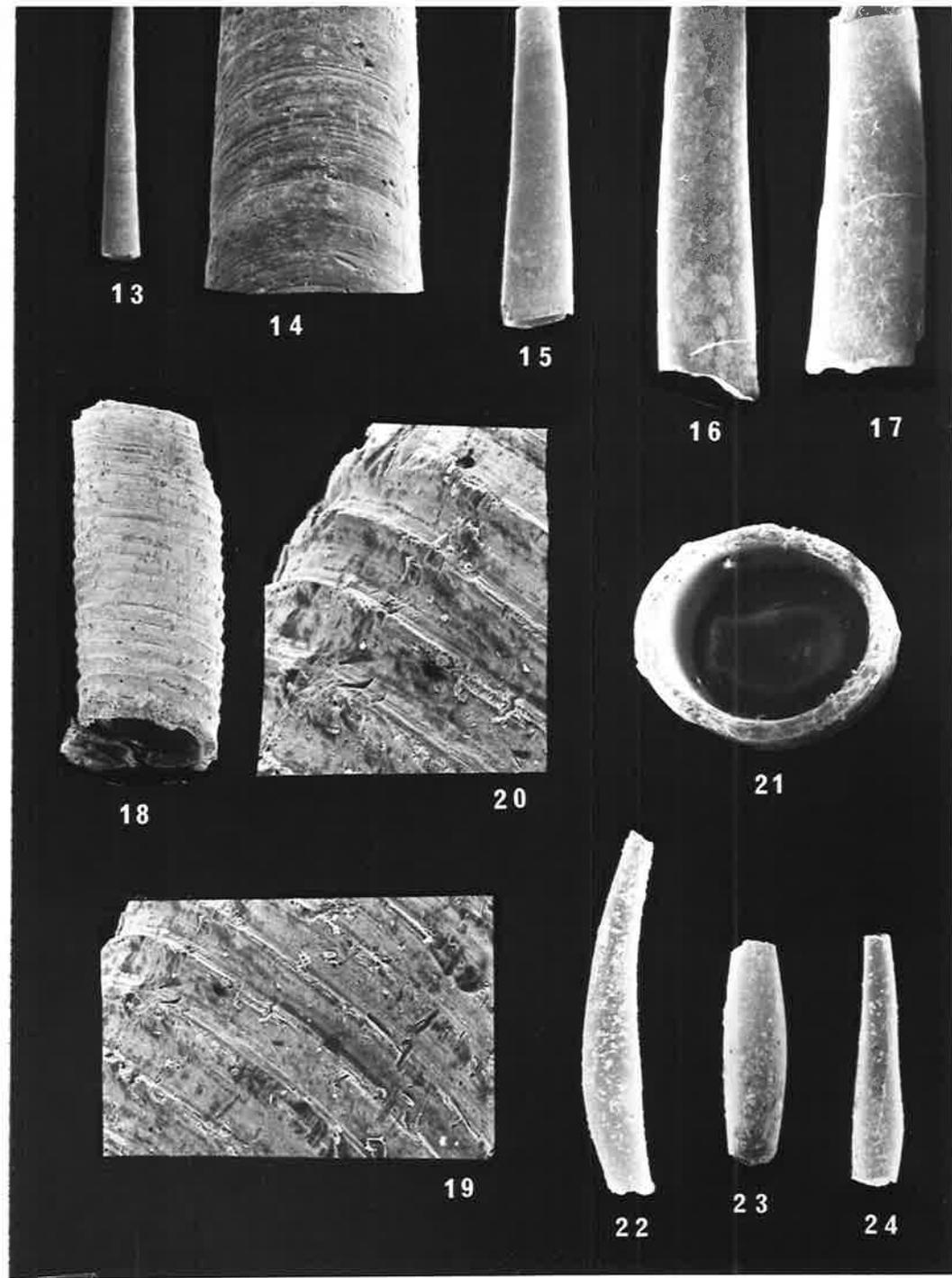
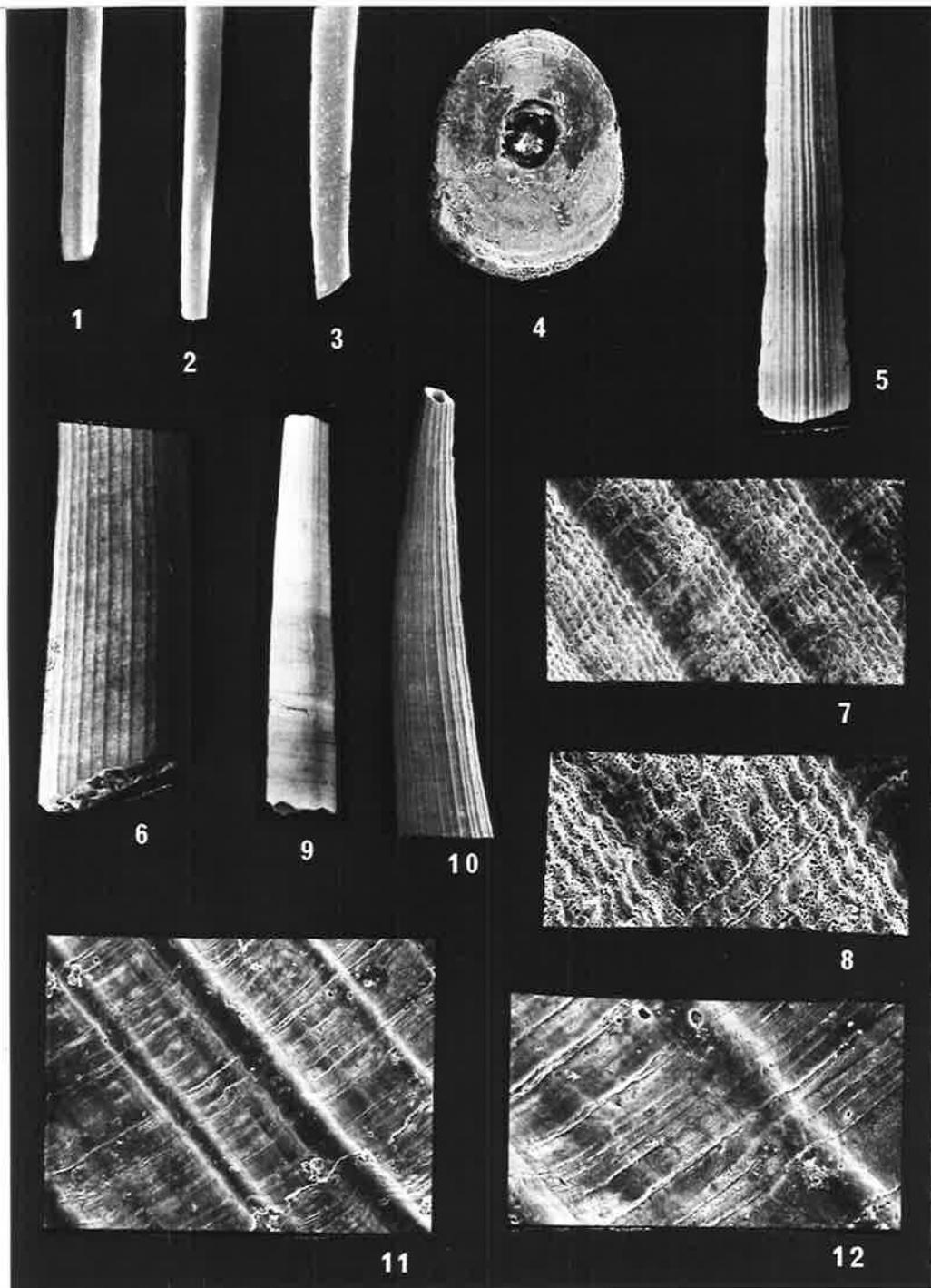
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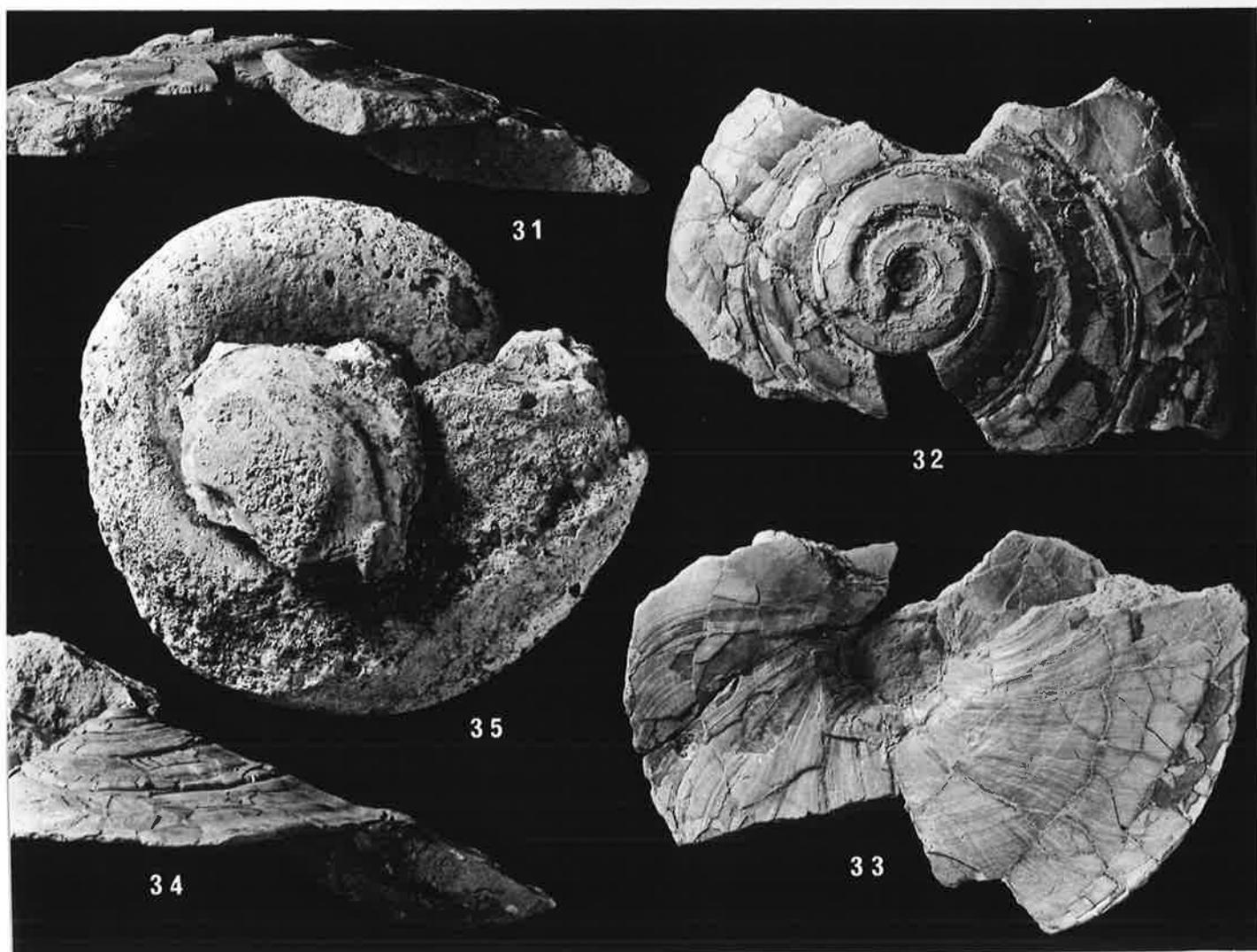
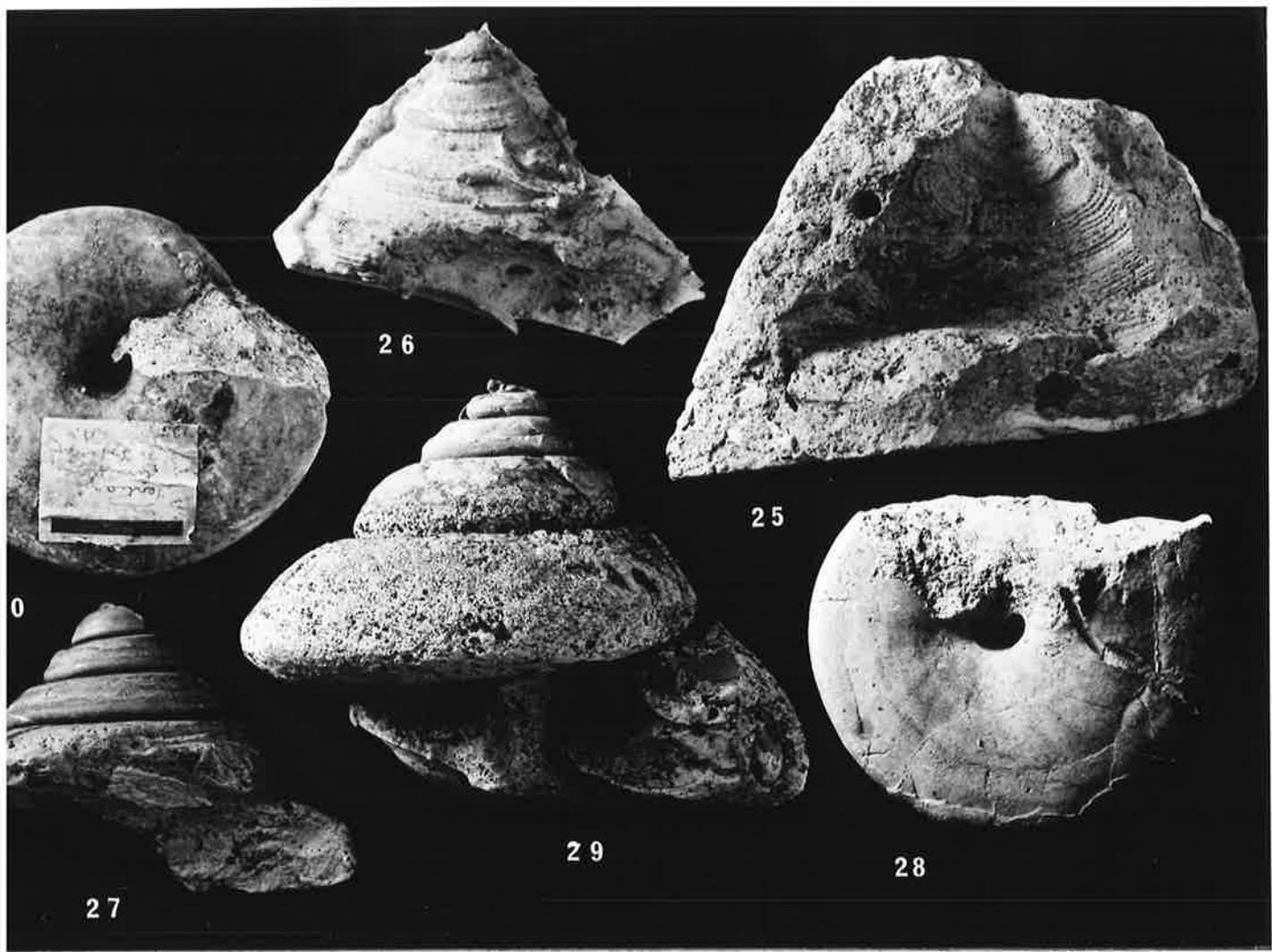
PLATES

ERRATA - CORRIGE

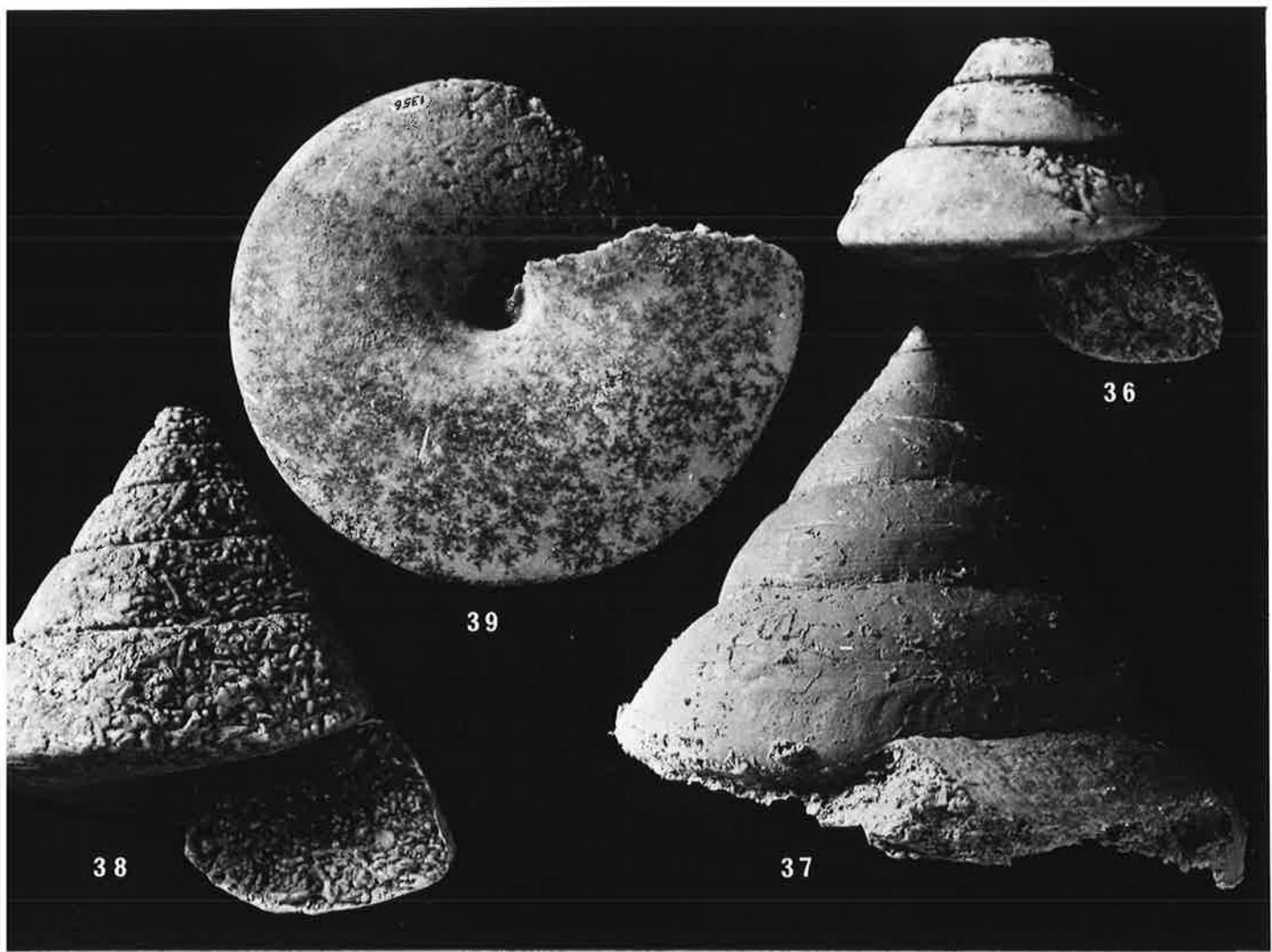
Explanation of figures

"particular" should be "detail"





- FIG. 36 Perotrochus sp. (SAM P 21270), axial view (x .75).
- FIG. 37 Perotrochus cf. tertiarius (McCoy) (SAM P 21269), Gambier Limestone, ?Oligocene, Pritchard's Quarry, axial view (x .77).
- FIGS. 38-39 Mikadotrochus sp. (AUGD 1356), Pt. Vincent Limestone, age unknown, Stansbury, Yorke Peninsula: 38) axial view (x .77); 39) abapical view (x .77).
- FIGS. 40-41 Perotrochus ?darraghi sp. nov. SAM P 13113. Views: 40) abapical (x .70); 41) axial (x .75). To note: 41) on the upper right, traces of ornament; 40) traces of umbilical features.
- FIGS. 42-44 Mikadotrochus sp. nov. NMV P 42704. Views: 42) axial (x 3); 43) adapical (x 4); 44) abapical (x 4).

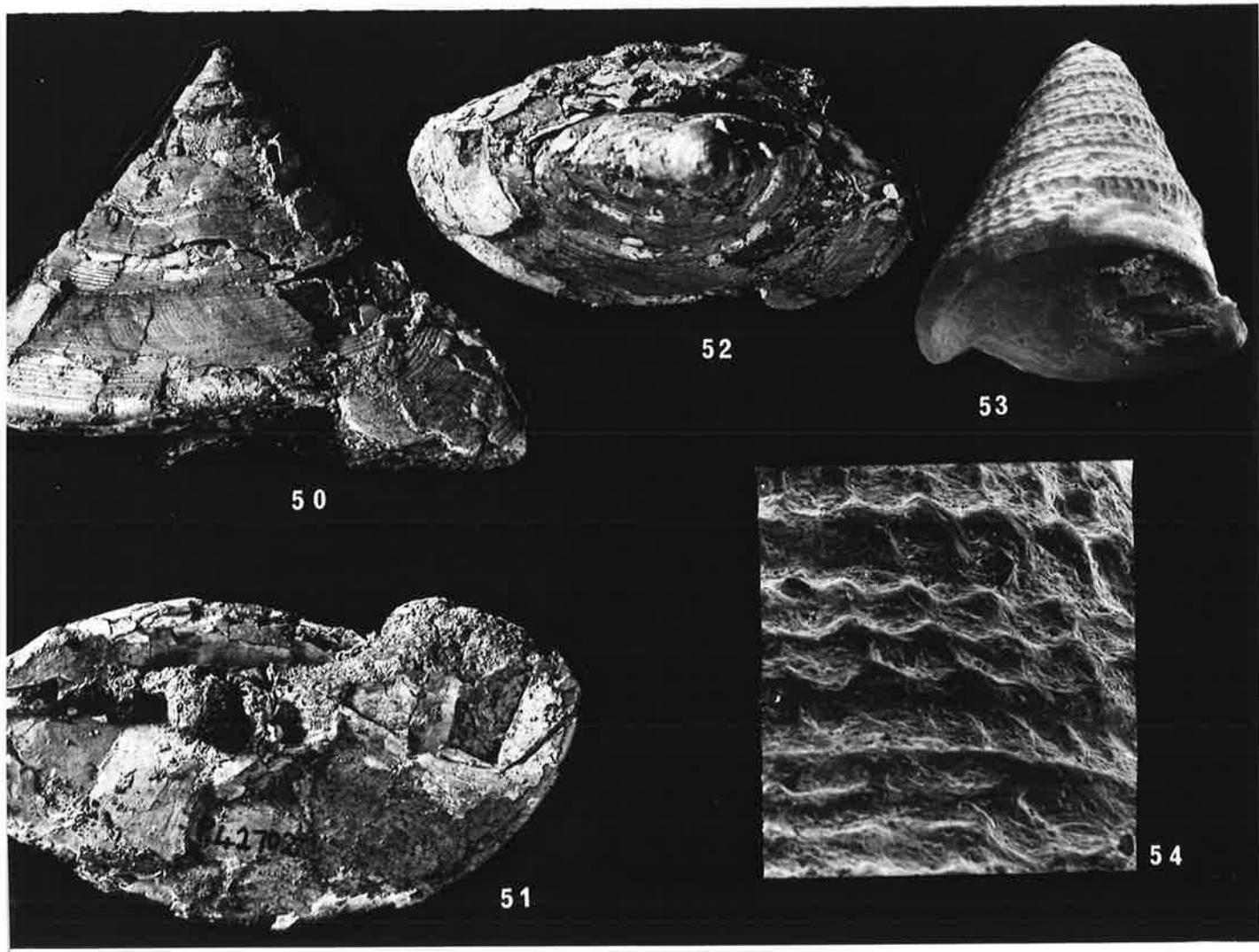


FIGS. 45-47 Perotrochus tertiarus (McCoy), NMV P 42701. Views: 45) axial oral (x .70); 46) abapical (x .75); 47) axial aboral (x .65).

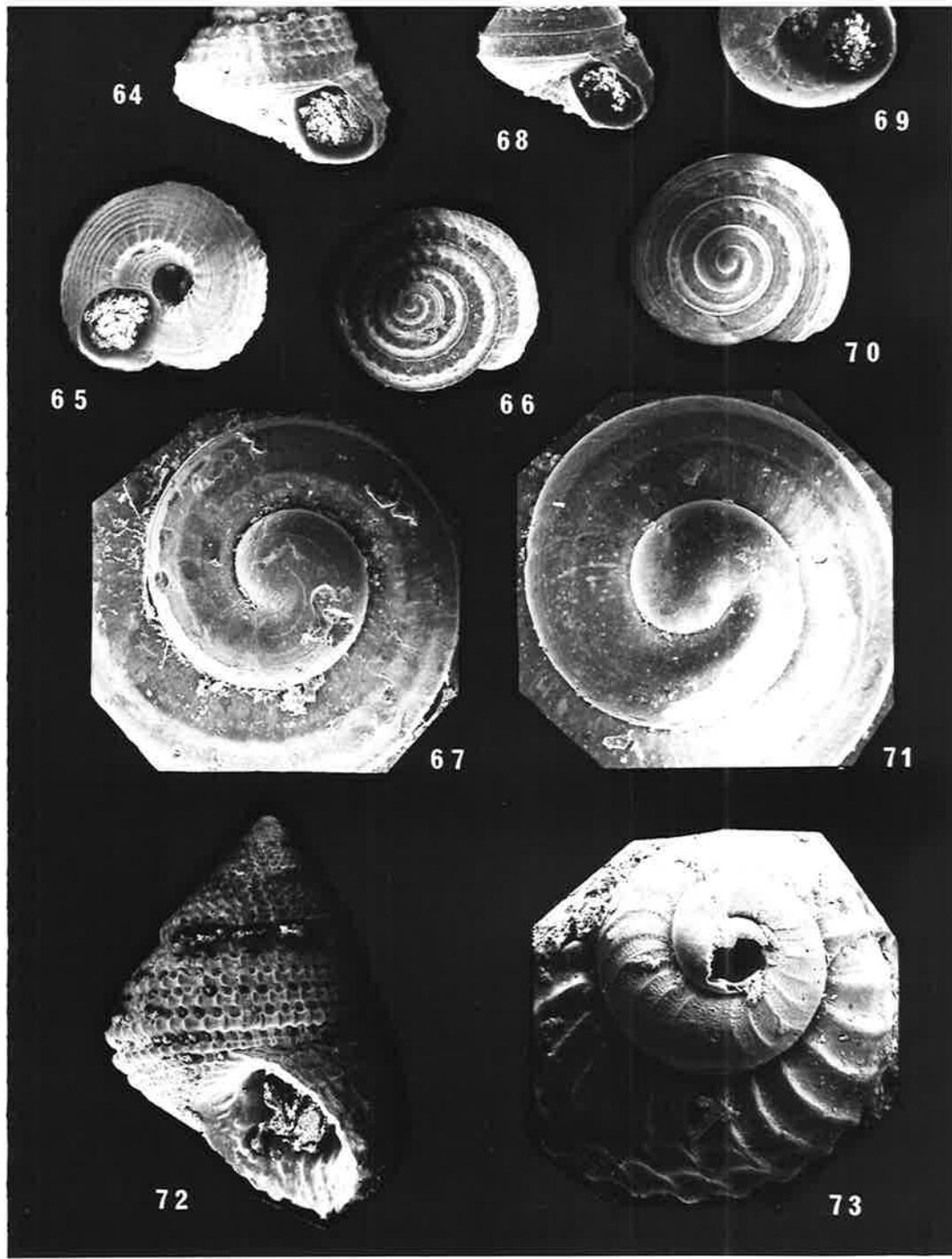
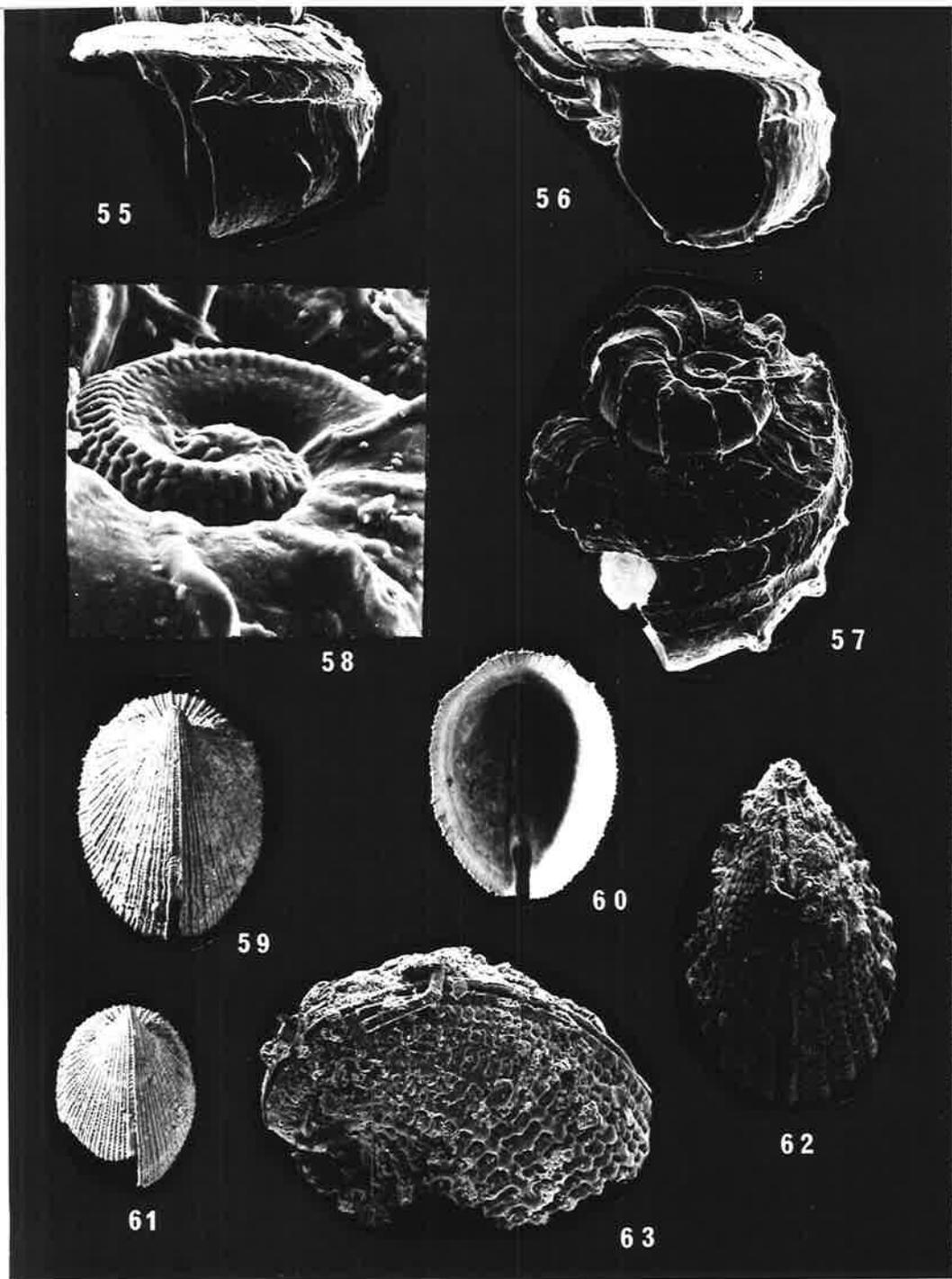
FIGS. 48-49 Mikadotrochus cf. bassi (Pritchard), NMV P 42703. Views: 48) axial (x 2); 49) abapical (x 2).

FIGS. 50-52 Perotrochus darraghi sp. nov., NMV P 42702. Views: 50) axial (x .95); 51) abapical (x 1.1); 52) adapical (x 1.0).

FIGS. 53-54 Perotrochus cf. tertiarius (McCoy) juv., NMV P 42705: 53) axial view (x 7.5); 54) particular ornament and selenizone (x 30).



- FIGS. 55-58 Scissurella (Scissurella) lamellularum sp. nov., GSSA M 3325. 55) axial, dorso-oral view (x 70); 56) oral view (x 70); 57) adapical view, tilted (x 70); 58) protoconch (x 300).
- FIGS. 59-63 Emarginula (Emarginula) imbricata sp. nov. Holotype, SAM P 21271-A: 59) dorsal view (x 2.45); 60) oral view (x 2.45). Paratype, SAM P 21271-B: 61) dorsal view (x 2.45). Paratype, SAM P 21271-C: juvenile: 62) dorsal view (x 32); 63) lateral view (x 40).
- FIGS. 64-67 Margarites (Periaulax) rhysus sp. nov. M. margaritatus Holotype. SAM P 21272-A: 64) axial view (x 10); 65) abapical view (x 10). SAM P 21272-B: 66) adapical view (x 10); 67) protoconch (x 60).
- FIGS. 68-71 Margarites (Periaulax) rhysus sp. nov., m. laevigatus. SAM P 21273-A: 68) axial view (x 10); 69) abapical view (x 10); SAM P 21273-B: 70) adapical view (x 10); 71) protoconch (x 40).
- FIGS. 72-73 Olivia sp. nov. 72) SAM P 21274-A: axial view (x 10); 73) SAM P 21274-B: protoconch and early whorls (x 60).

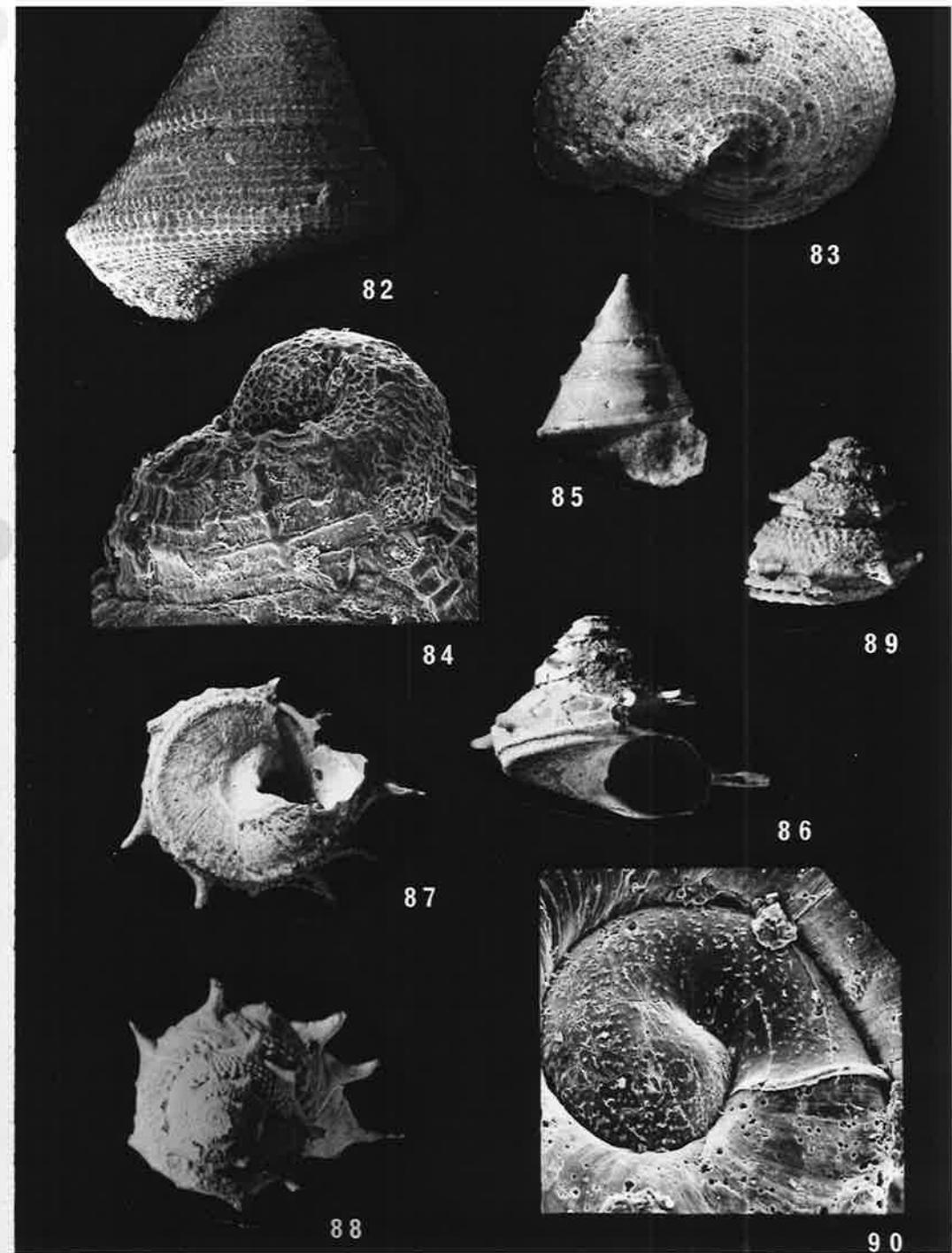
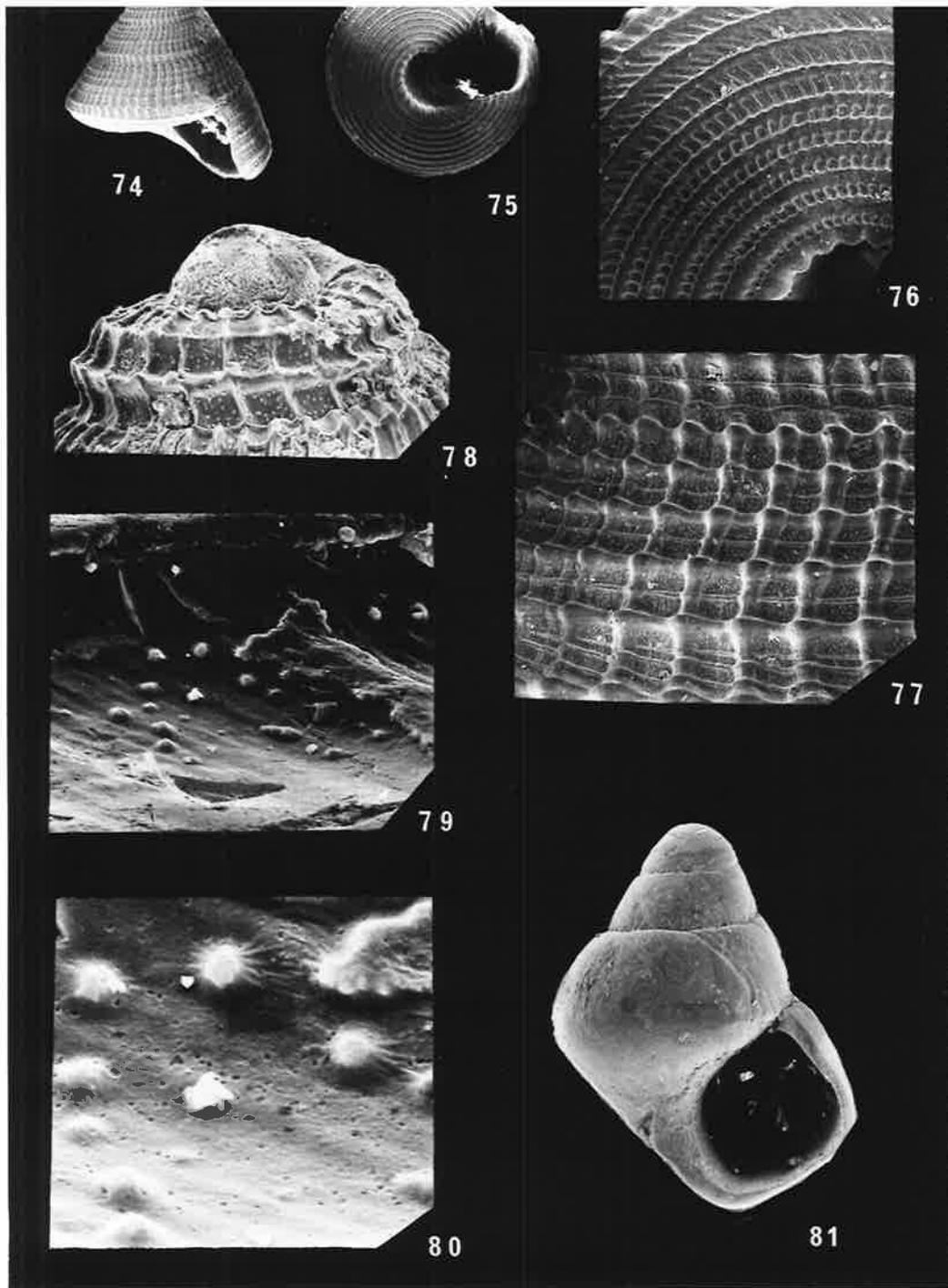


FIGS. 74-80 · Basilissa (Basilissa) cossmanni Tate, GSSA M 3446. Views:
74) axial (x 10); 75) abapical (x 10); 76) abapical
ornament, particular (x 30); 77) last whorl ornament,
particular (x 50); 78) protoconch (x 110); 79) pustulate
interspaces, particular (x 600); 80) micro-pustulae,
particular (x 2000).

FIG. 81 Gastropoda gen & sp. ind. GSSA M 3447, axial view (x 10).

FIGS. 82-85 Calliostoma (Fautor) allasinazi sp. nov., Paratype, SAM P
18345-C; 82) axial view (x 10); 83) abapical view, tilted
(x 10); 84) protoconch (x 100). SAM P 18345-A; holotype, worn
surface; 85) axial view (x 2.45).

FIGS. 86-90 Guildfordia (Pseudastralium) maslinensis sp. nov.
Holotype, SAM P 18346-B: 86) axial view (x 2.25); 87)
abapical view (x 2.25); 88) adapical view (x 2.25). Paratype
SAM P 18346-C: 89) axial view (x 2.25). Paratype M 3448,
juvenile: 90) protoconch (x 140).



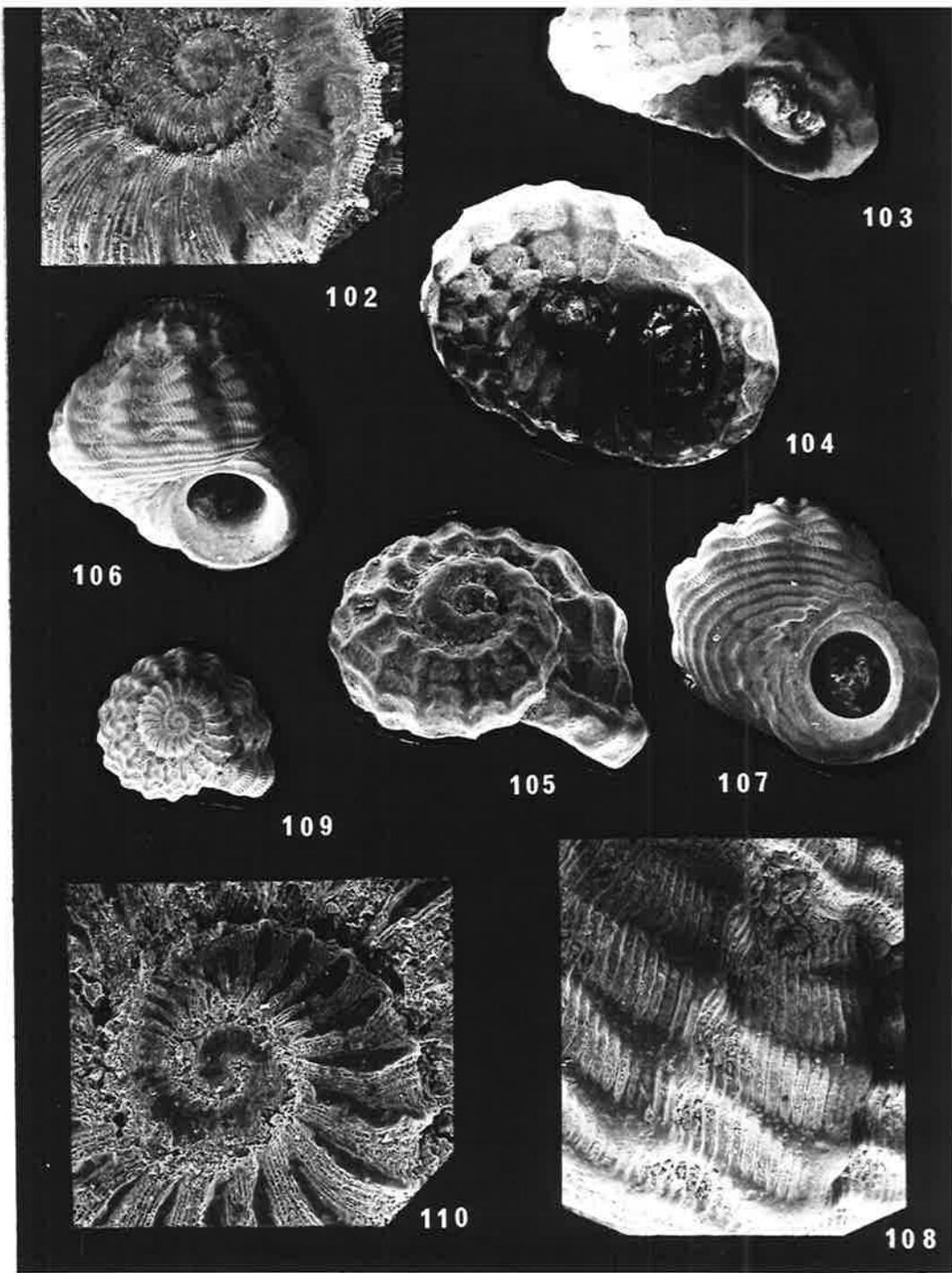
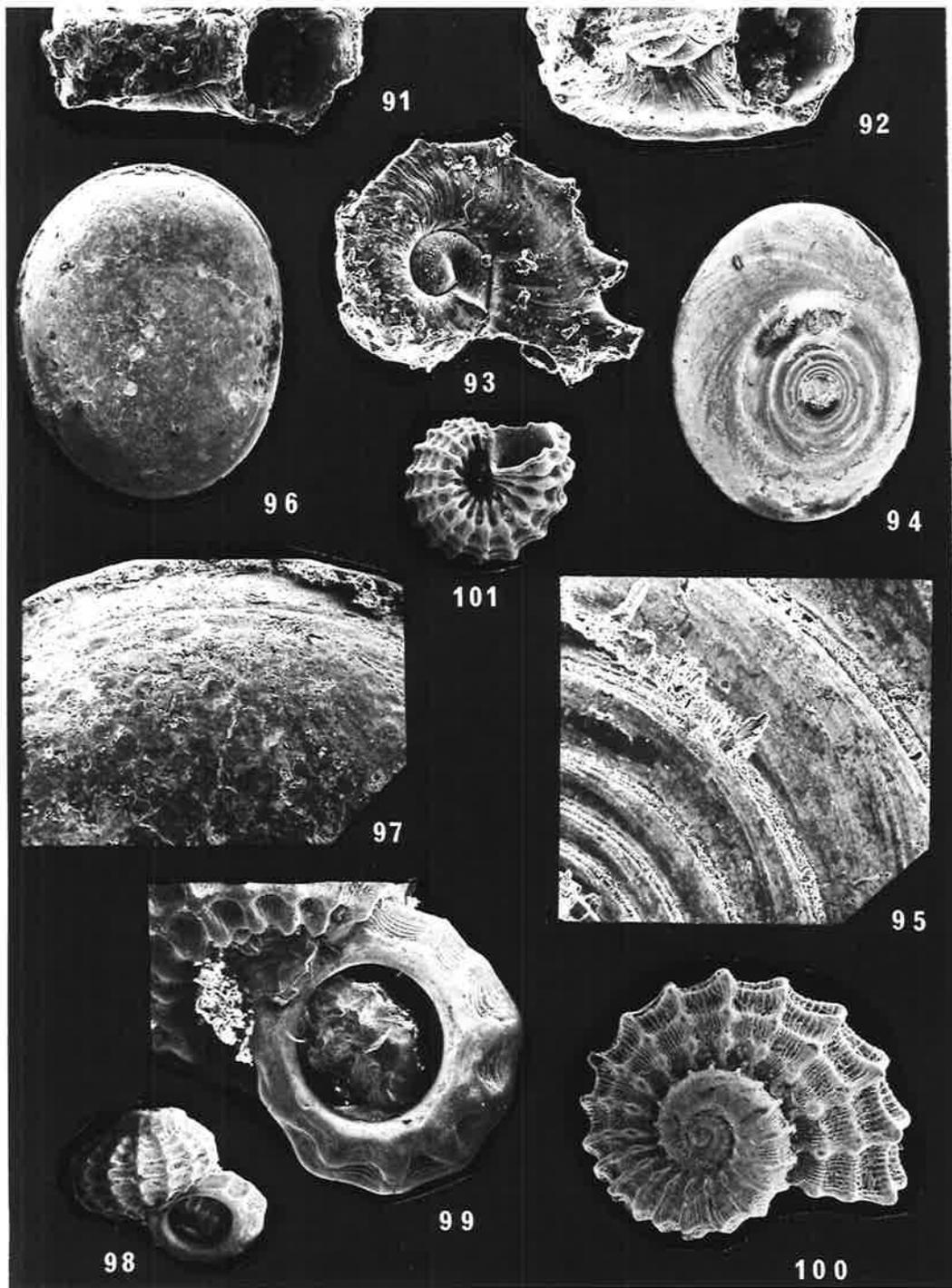
FIGS. 91-97 Guildfordia (Pseudastraliu) maslinensis sp. nov. Paratype GSSA M 3448: 91) axial view (x 40); 92) abapical view, tilted (x 40); 93) adapical view (x 40). Opercula. SAM P 18346-Q: 94) outer surface (x 10); 95) particular whorls and suture (x 60) SAM P 18346-R: 96) inner surface (x 10); 97) particular inner surface (x 30).

FIGS. 98-101 Liotina (Austroliotia) intermedia sp. nov. Holotype, GSSA M 3449; 98) axial view (x 10); 99) peristome and umbilicus (x 20). Paratypes: 100) GSSA M 3450, adapical view (x 20); 101) GSSA M 3451, abapical view (x 10).

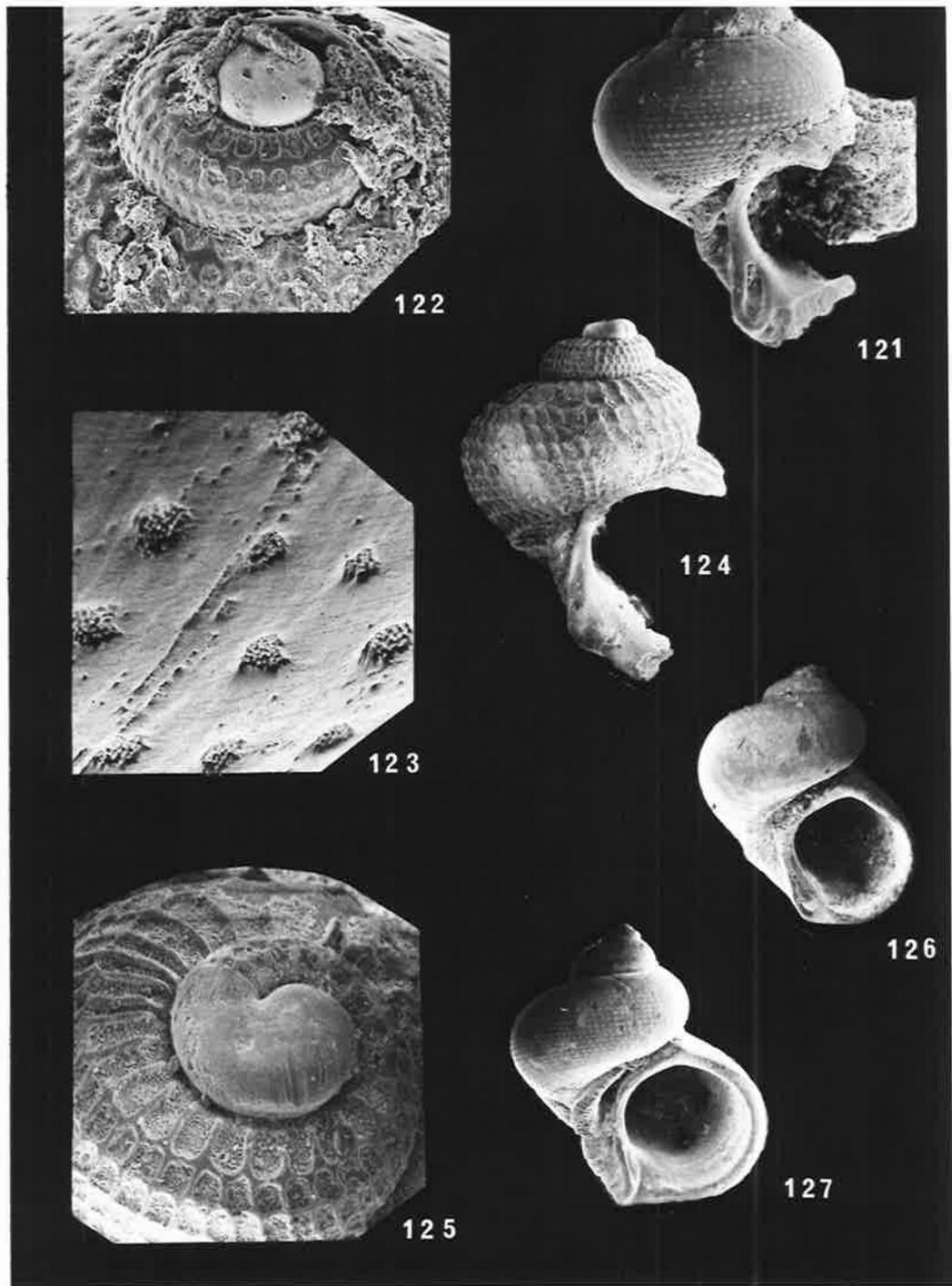
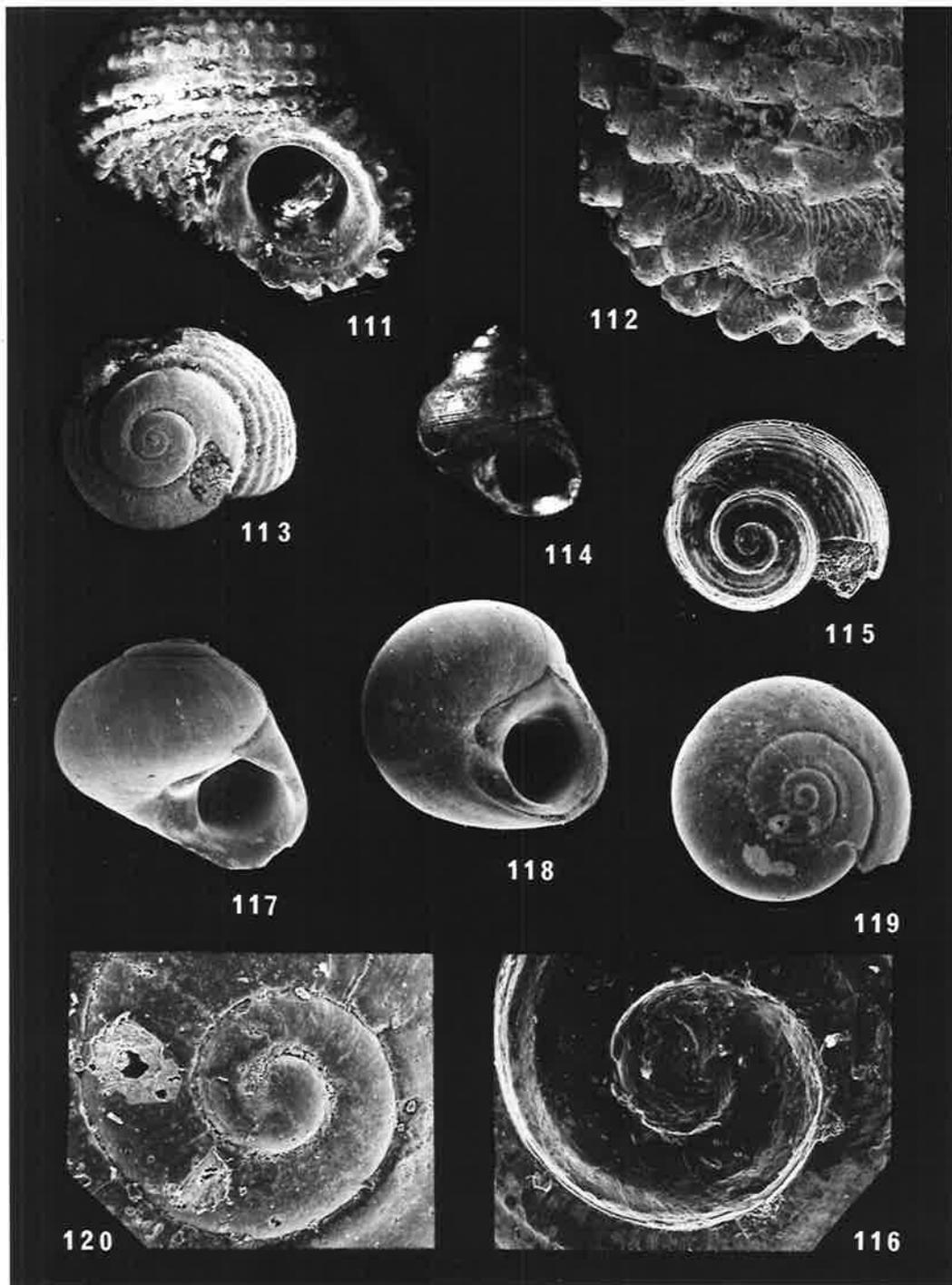
FIG. 102 Liotina (Austroliotia) intermedia sp. nov. Paratype GSSA M 3450, protoconch (x 50).

FIGS. 103-105 Liotina (Austroliotia) ampla sp. nov. Holotype, GSSA M 3454: 103) axial view (x 20); 104) abapical view, tilted (x 20); 105) adapical view, tilted (x 20).

FIGS. 106-110 Liotina (Austroliotia) stricta sp. nov. Holotype, GSSA M 3452: 106) axial view (x8.8); 107) abapical view, tilted (x8.85); 108) particular last whorl ornament (x 30). Paratype GSSA M 3453: 109) adapical view (x 10); 110) protoconch (x 60).



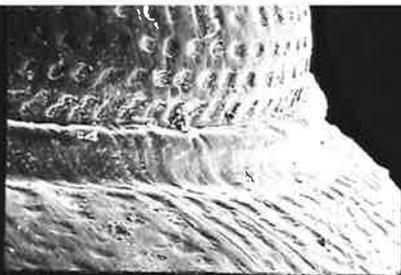
- FIGS. 111-113. Cycloliotia hyotis sp. nov. Holotype, SAM P 21274: 111) axial view (x 10); 112) particular last whorl ornament (x 30). Paratype, SAM P 21275: 113) adapical view (x 10).
- FIGS. 114-116 ?Homalopoma (?Caintrainea) ancestralis sp. nov.: 114) holotype, GSSA M 3455, axial view (x 3); 115) paratype, GSSA M 3456, adapical view (x 20); 116) paratype's protoconch (x 80).
- FIGS. 117-120 Vexinia callosa sp. nov. Holotype, GSSA M 3457: 117) axial view (x 20); 118) abapical view, tilted (x 20). Paratype, GSSA M 3458: 119) adapical view (x 20); 120) protoconch (x 70).
- FIGS. 121-123 Crossea (Crosseola) antiqua sp. nov. Holotype, SAM T 806-E: 121) axial view (x 20); 122) protoconch (x 90); 123) last whorl punctations, particular (x 500).
- FIGS. 124-125 Crossea (Crosseola) semiornata Tate. Paratype, SAM T 810-B: 124) axial view (x 30); 125) protoconch (x 100).
- FIG. 126 Crossea (Crosseola) evoluta sp. nov. Holotype, SAM T 806-G, axial view (x 10).
- FIG. 127 Crossea (Crosseola) intermedia sp. nov. Holotype, SAM T 806-B, axial view (x 10).



- FIGS. 128-129 Crossea (Crosseola intermedia sp. nov. Holotype: 128) protoconch (x 90); 129) last whorl ornament and suture, particular (x 70).
- FIGS. 130-132 Crossea (Crosseola) princeps Tate. Syntype, SAM T 806-D: 130) axial view (x 10); 131) early whorls, particular (x 40); 132) protoconch (x 70).
- FIGS. 133-136 Leucorhynchia bifuniculata sp. nov. Morpha A, paratypes: 133) GSSA M 3460-A, axial view (x 20); 134) GSSA M 3460-B adapical view (x 20). Morpha B, paratype, GSSA M 3460-B: 135) axial view (x 20); 136) protoconch (x 90).
- FIGS. 137-138 Leucorhynchia bifuniculata sp. nov. Holotype, GSSA M 3459-A, intermediate between Morphae A and B: 137) axial view (x 20); 138) protoconch (x 90).
- FIGS. 139-142 ?Parviturbo dubius sp. nov. Holotype, GSSA M 3461: 139) axial view (x 10); 140) last whorl ornament (x 40); 141) protoconch and pustular early whorls (x 80); 142) particular micropustulae (x 1000).
- FIGS. 143-145 Lironoba (Nobolira) costata sp. nov. Holotype, SAM P 21276: 143) axial view (x 20); 144) particular spiral ornament (x 140); 145) protoconch (x 80).



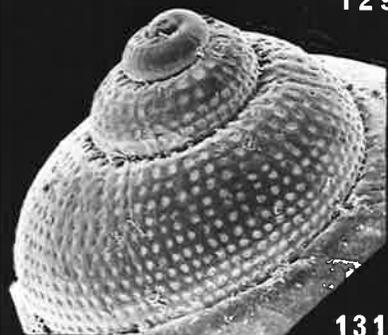
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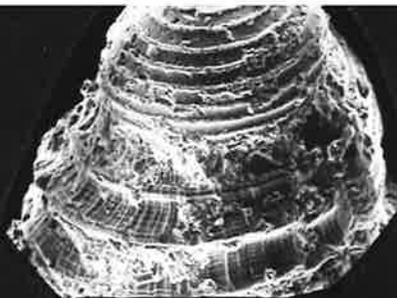
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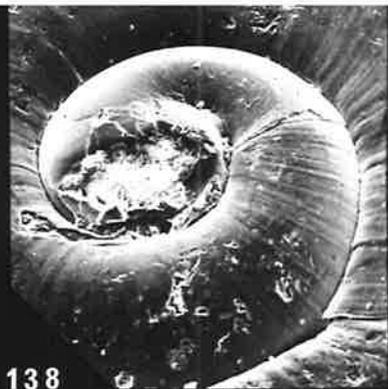
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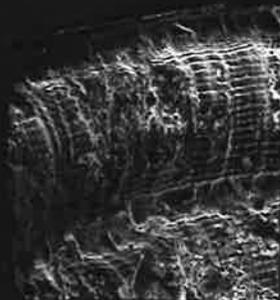
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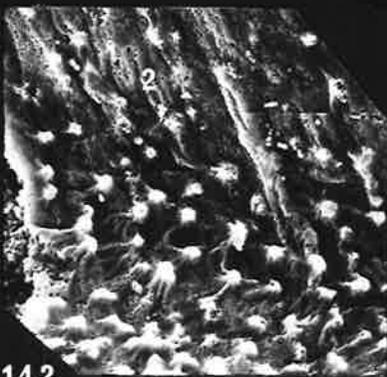
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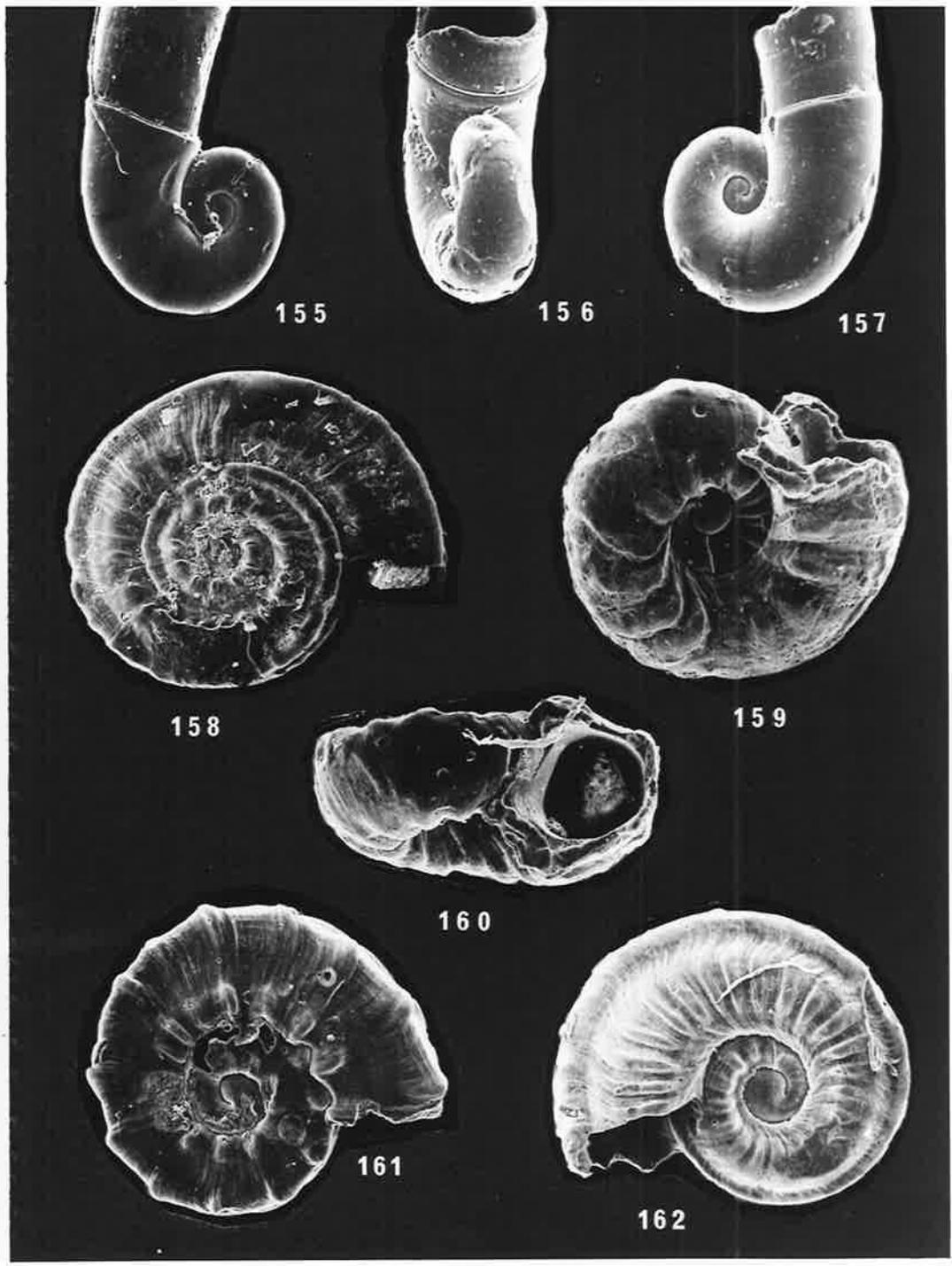
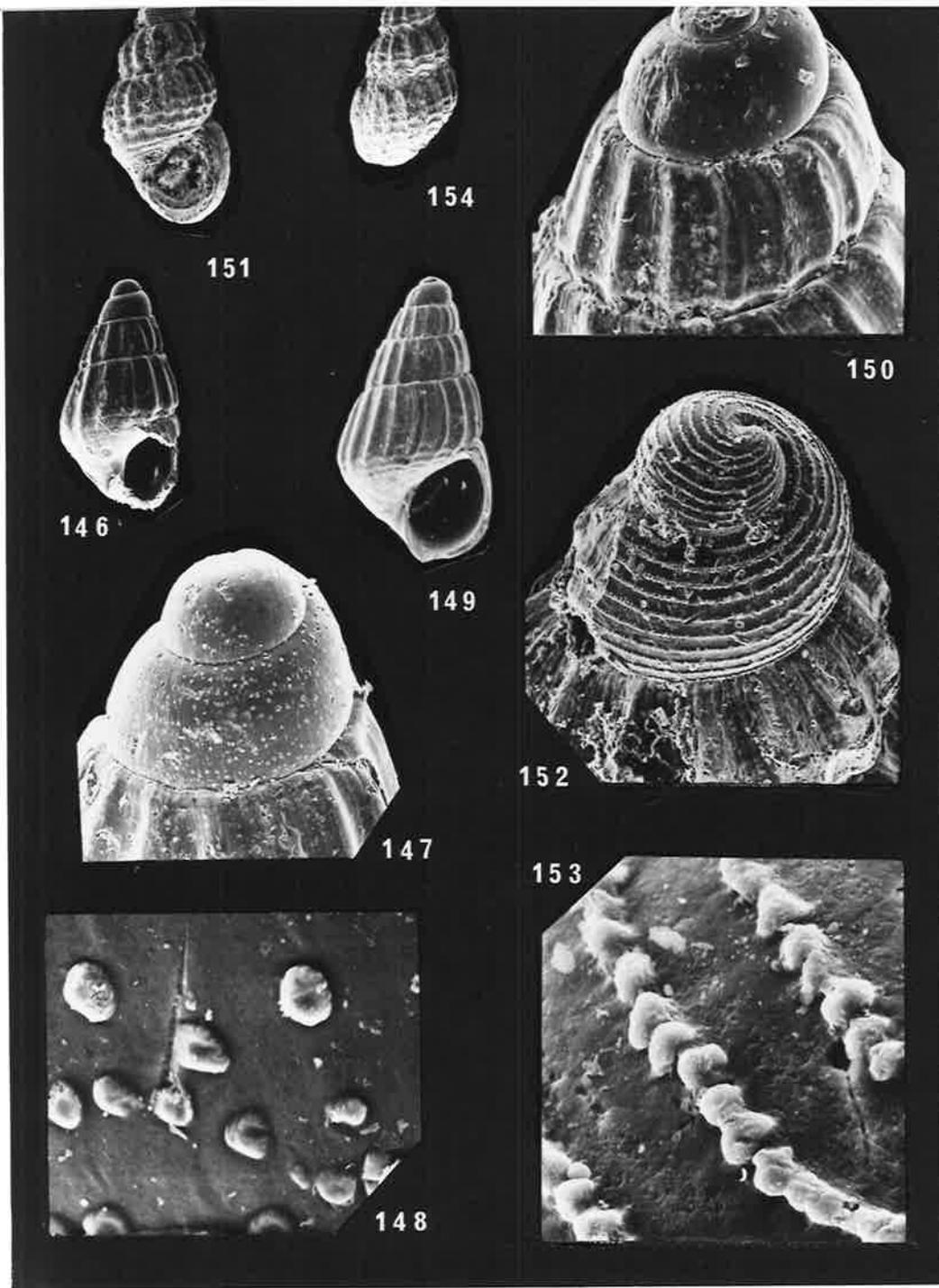


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- FIGS. 146-148 Rissoa (Haurakia) costata sp. nov. Holotype, GSSA M 3462: 146) axial view (x 20); 147) protoconch (x 80); 148) protoconch pustulae, at the protoconch/teleoconch suture (x 1200).
- FIGS. 149-150 Turboella (Turboella) flexilis sp. nov. Holotype, GSSA M 3463: 149) axial view (x 20); 150) protoconch (x 80).
- FIGS. 151-154 Merelina kaurna sp. nov. Holotype, SAM P 2177-A: 151) axial view (x 20); 152) protoconch (x 80); 153) particular protoconch cords (x 1300). Paratype, SAM P 21277-B: 154) axial view (x 20).
- FIGS. 155-157 Strebloceras darraghi sp. nov. Holotype, GSSA M 3322: 155) adapical view (x 50). Paratypes: 156) GSSA M 3322-A, axial view (x 80); 157) GSSA M 3322-B, abapical view (x 80).
- FIGS. 158-160 Orbitestella margaritata sp. nov. Holotype, GSSA M 3323-A: 158) adapical view (x 50). Paratypes: 159) GSSA M 3323-B, abapical view (x 90); 160) GSSA M 3323-C axial view (x 90).
- FIGS. 161-162 Orbitestella spinosa sp. nov. 161) Holotype, GSSA M 3321-A, adapical view (x 80); 162) Paratype, GSSA M 3321-B, abapical view (x 90).



FIGS. 163-165 Orbitestella rugosa sp. nov. Paratypes: 163) GSSA M 3319-A, adapical view, slightly tilted (x 60); 164) GSSA M 3319-B, abapical view (x 70). Holotype, GSSA M 3318: 165) axial view (x 50).

FIGS. 166-167 Orbitestella bastowi (Gatliff). SAM TD 383: 166) adapical view, slightly tilted (x 50); 167) abapical view (x 70).

FIGS. 168-169 ?Elachorbis pentagonalis sp. nov. 168) Holotype, GSSA M 3320-A, adapical view (x 50); 169) Paratype, GSSA M 3320-B, abapical view (x 50).

FIGS. 170-175 Spirocolpus aldingae (Tate) SAM P 21161-A-F. Morphae: from left to right, suture from flush to grooved. Axial views (all x 2.30).

FIGS. 176-179 Zeacolpus (Stiracolpus) sp., SAM P 21162-A-D. All axial views: 176) x 3.5; 177) x 3.35; 178) x 3.5; 179) x 3.4.



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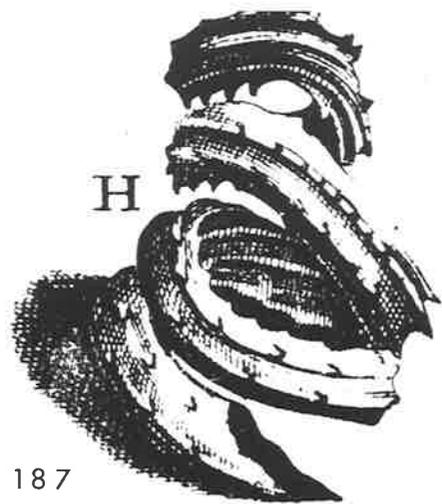
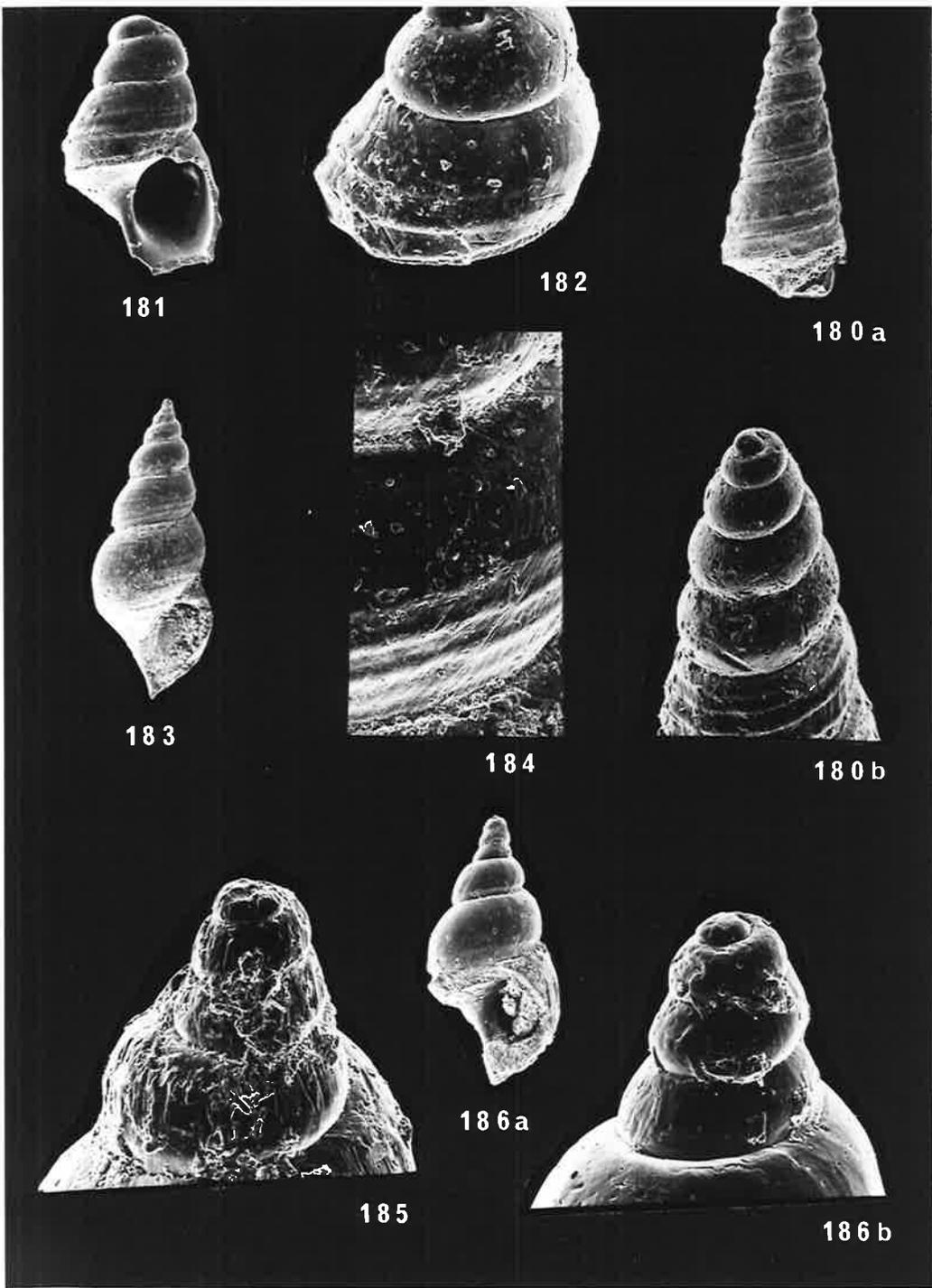
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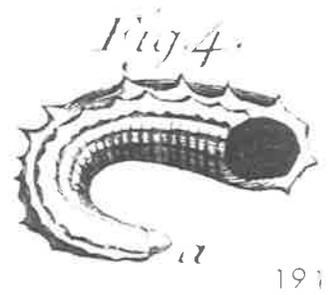
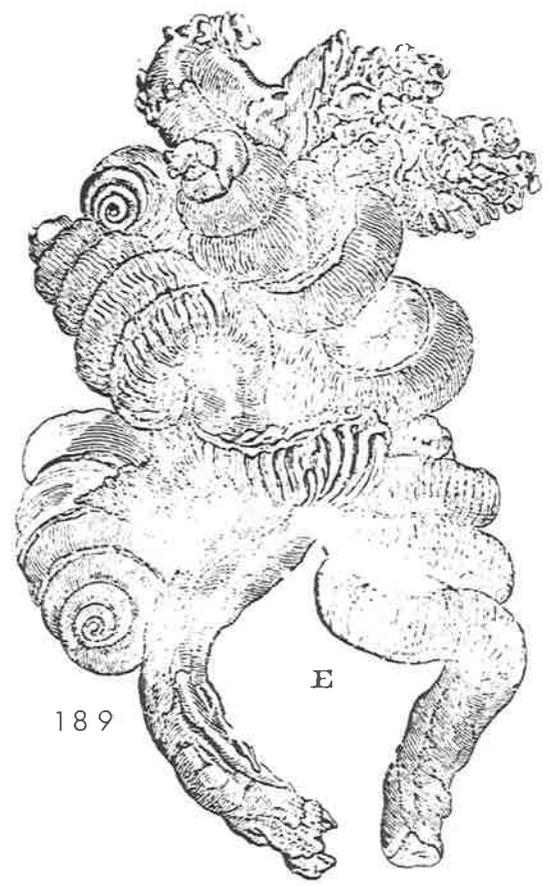
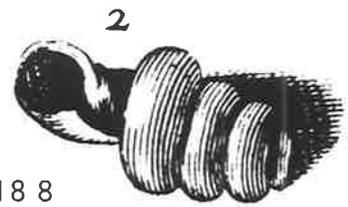
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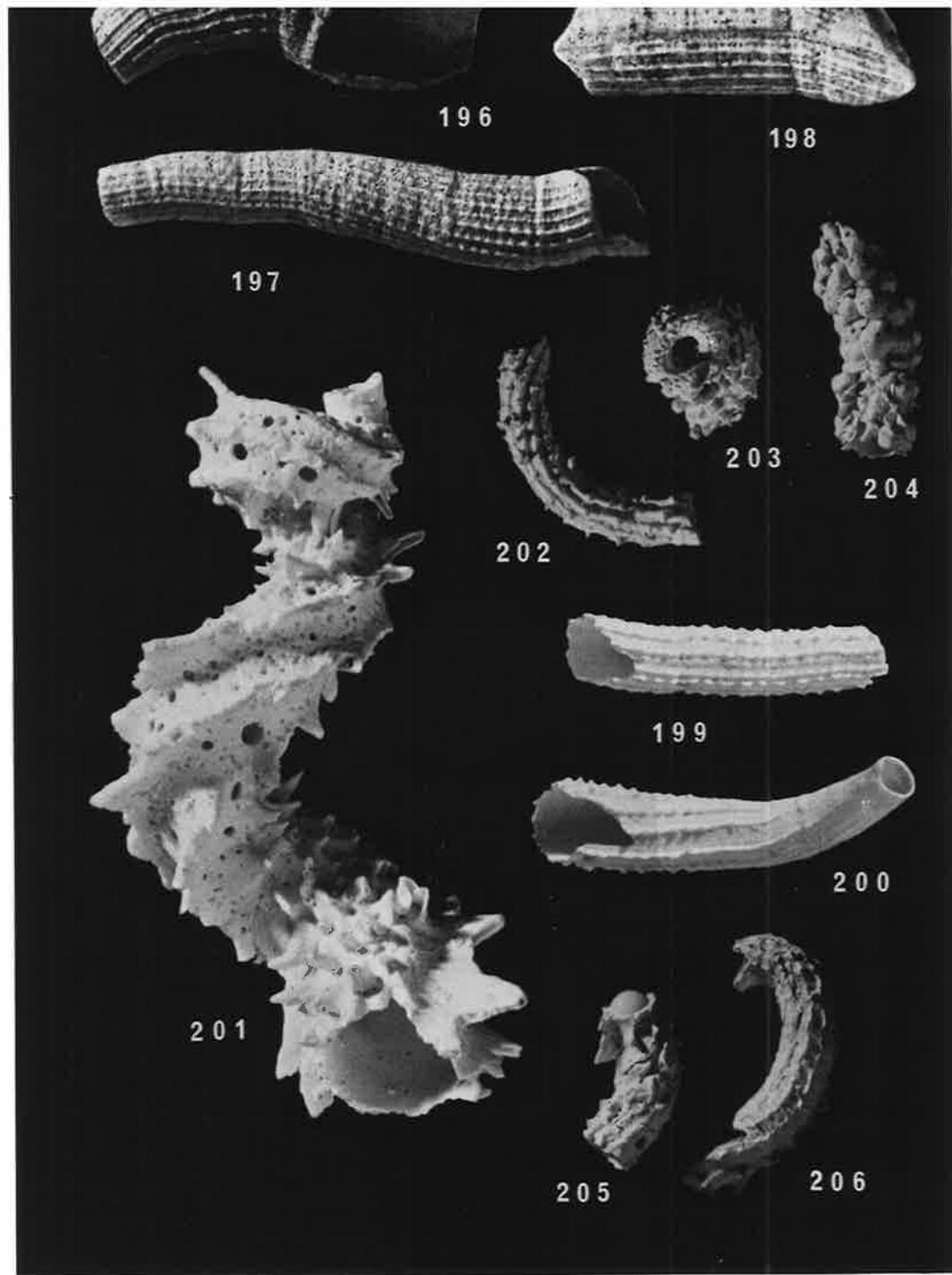
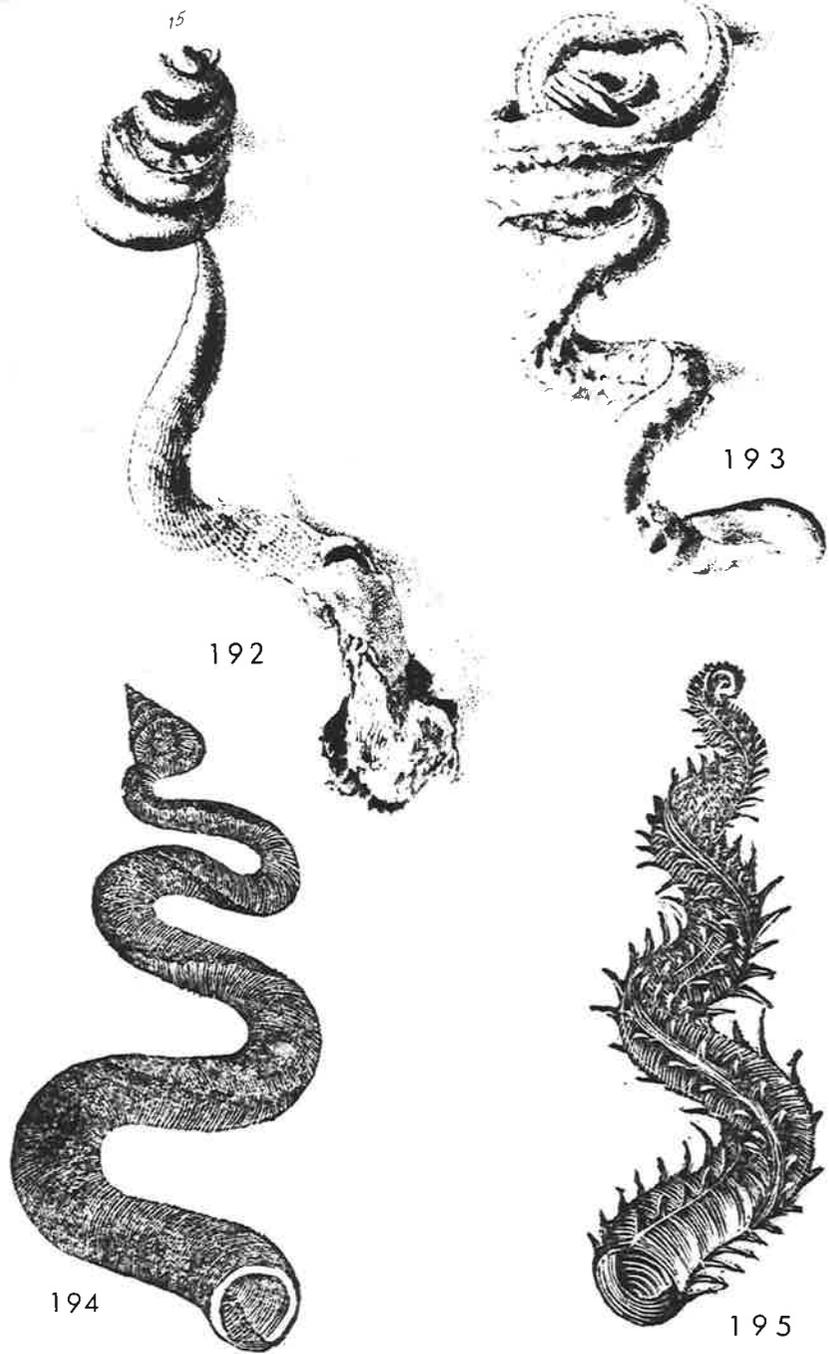
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u. verticillata Jun-
 -ura quadam Secur-
 -dium volutas insig-
 -intus.



- FIG. 192 Serpula anguina Born, Born's original illustration, pl. 18, fig. 15.
- FIG. 193 Serpula muricata Born, Born's original illustration, pl. 18, fig. 16.
- FIG. 194 Agathyrsos furcellus Montfort, Montfort's 1808 original illustration, I, p.598.
- FIG. 195 'Siliquarius anguilus' Montfort, Montfort's 1810 original illustration, II, p.38.
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- FIGS. 196-197 Campylothyrsos gen. nov. multistriatus (Defrance), SAM P 21278, Auvers (Late Eocene) (x 2.6).
- FIG. 198 Campylothyrsos gen. nov. mitis (Deshayes), SAM P 21279, Auvers (Late Eocene). To note, the slit. (x 2.6).
- FIGS. 199-200 Campylothyrsos gen. nov. brevifissuratus (Deshayes), Coll. Buonaiuto, La Ferme de L'Orme, middle Lutetian (by courtesy of J. Le Renard). 199) x 3.05; 200) x 3.15.
- FIG. 201 Agathyrsos furcellus Montfort, Coll. Le Renard, La ferme de l'Orme, middle Lutetian (x 3.6).
- FIGS. 202-204 Agathyrsos millepeda (Deshayes). Grignon, middle Lutetian (by courtesy of J. Le Renard). 203-4) Coll. Buonaiuto (x 1.9); 204) Coll. Le Renard (x 1.7).
- FIGS. 205-206 Agathyrsos lima (Lamarck), Coll. Buonaiuto, Grignon, middle Lutetian (by courtesy of Le Renard) (x 1.7). To note, the slit in Fig. 206.

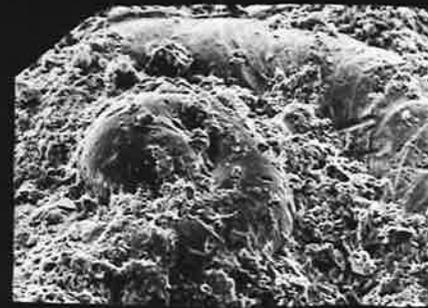
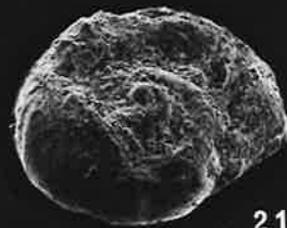
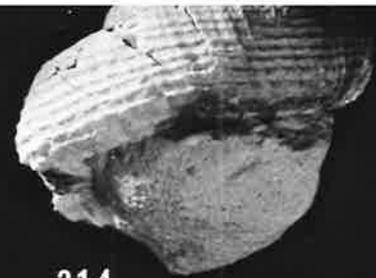
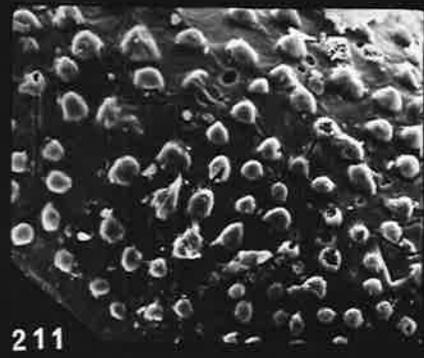


FIGS. 207-211 Pyxipoma squamigera sp.nov. Holotype, SAM P 21166: 207) axial view (x 3.30). Paratypes: 208-09) SAM P 21167-A & B, axial view (x 2.75). Paratype GSSA M 3464: 210) protoconch (x 80); 211) particular pustulae (x 400).

FIGS. 212-213 Siliquaria altispira sp. nov. Holotype, SAM P 21168: 212) axial view (x 1.85). Paratype, GSSA M 3465: 213) axial view (x 2.30).

FIGS. 214-220 Siliquaria kaurna sp. nov. Holotype, SAM P 21169: 214) axial view (x 2.00); 215) adapical view (x 2.00). Paratypes, SAM P 21170A-C, Juveniles: 216-218) adapical view (all x 2.00). Paratype SAM P 21171, juvenile: 219) adapical view (x 20); 220) protoconch (x 80).

FIG. 221 Siliquaria sp. nov. A. SAM P 21172, axial view (x 2.7)



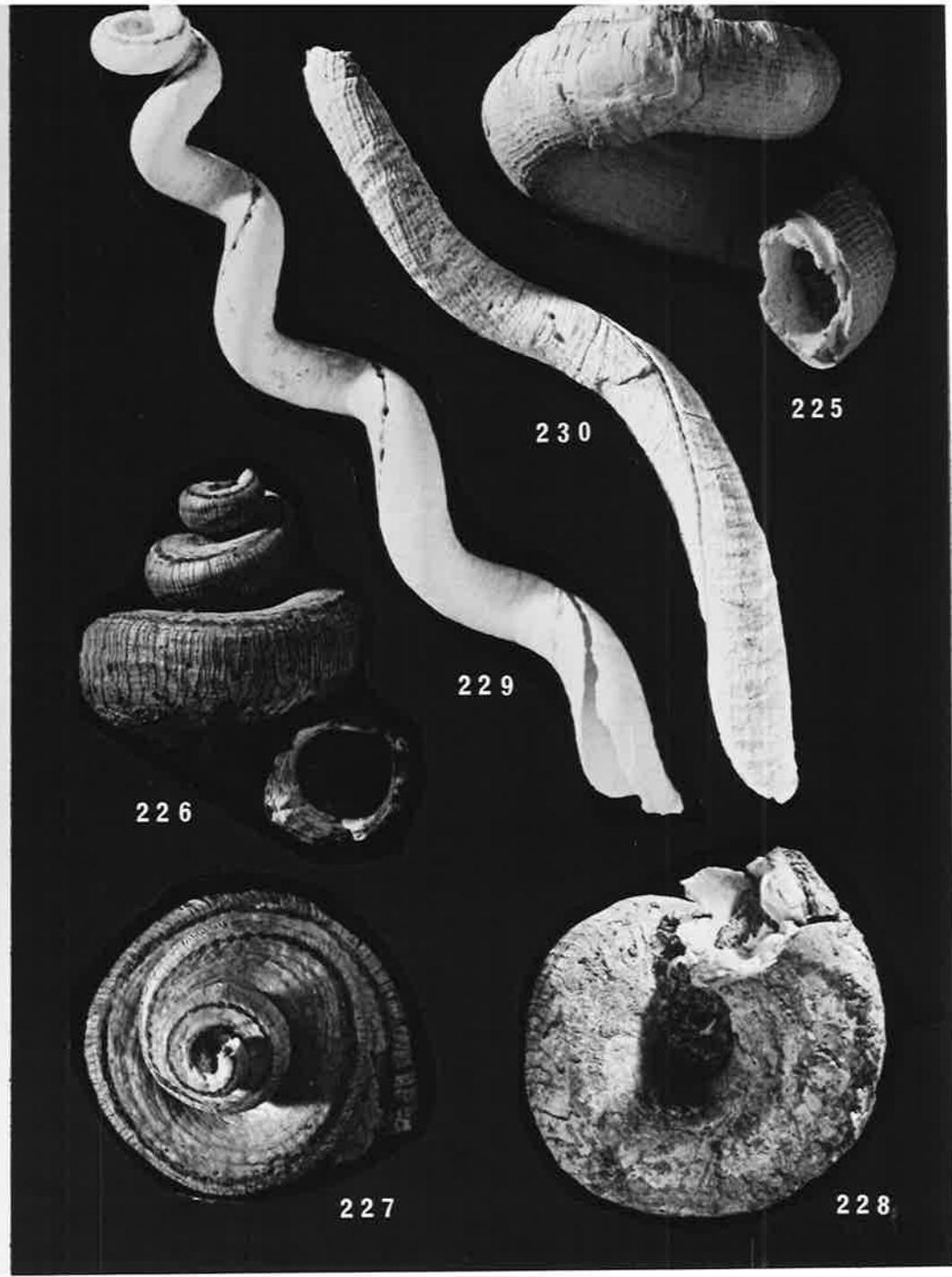
FIGS. 222-224 Siliquaria cadelli sp. nov. Holotype, SAM P 21173: 222) adapical view (x 1.5); 223) abapical view (x 1.5). Paratypes GSSA M 3316 ; 224) specimens still imbedded in matrix (x .70).

FIG. 225 Siliquaria cadelli sp. nov. Holotype, axial view (x 1.7).

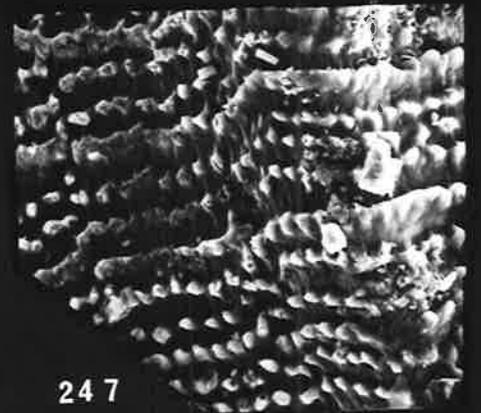
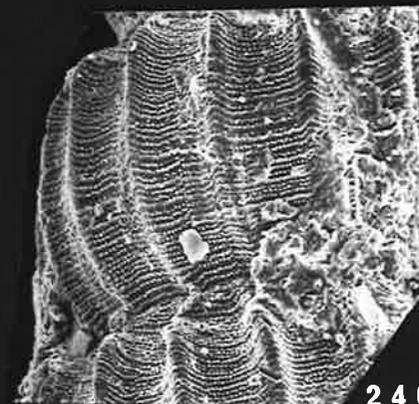
FIGS. 226-278 Siliquaria occlusa (T. Woods), Neotype, SAM P 21174: 226) axial view (x 2.4); 227) adapical view (x 2.45); 228) abapical view (x 2.7).

FIG. 229 Siliquaria sp. nov. B. SAM P 21176, axial view (x 2.85).

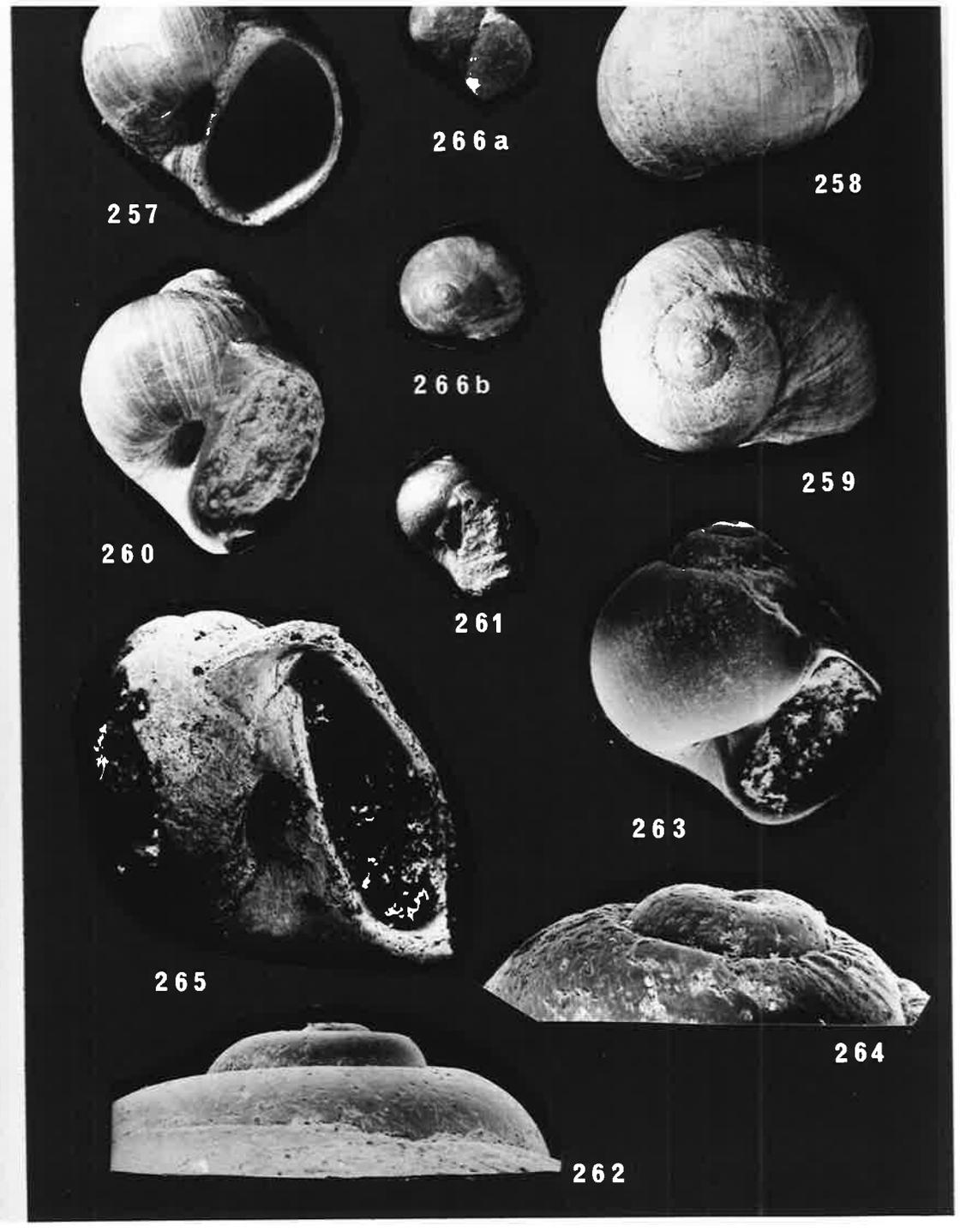
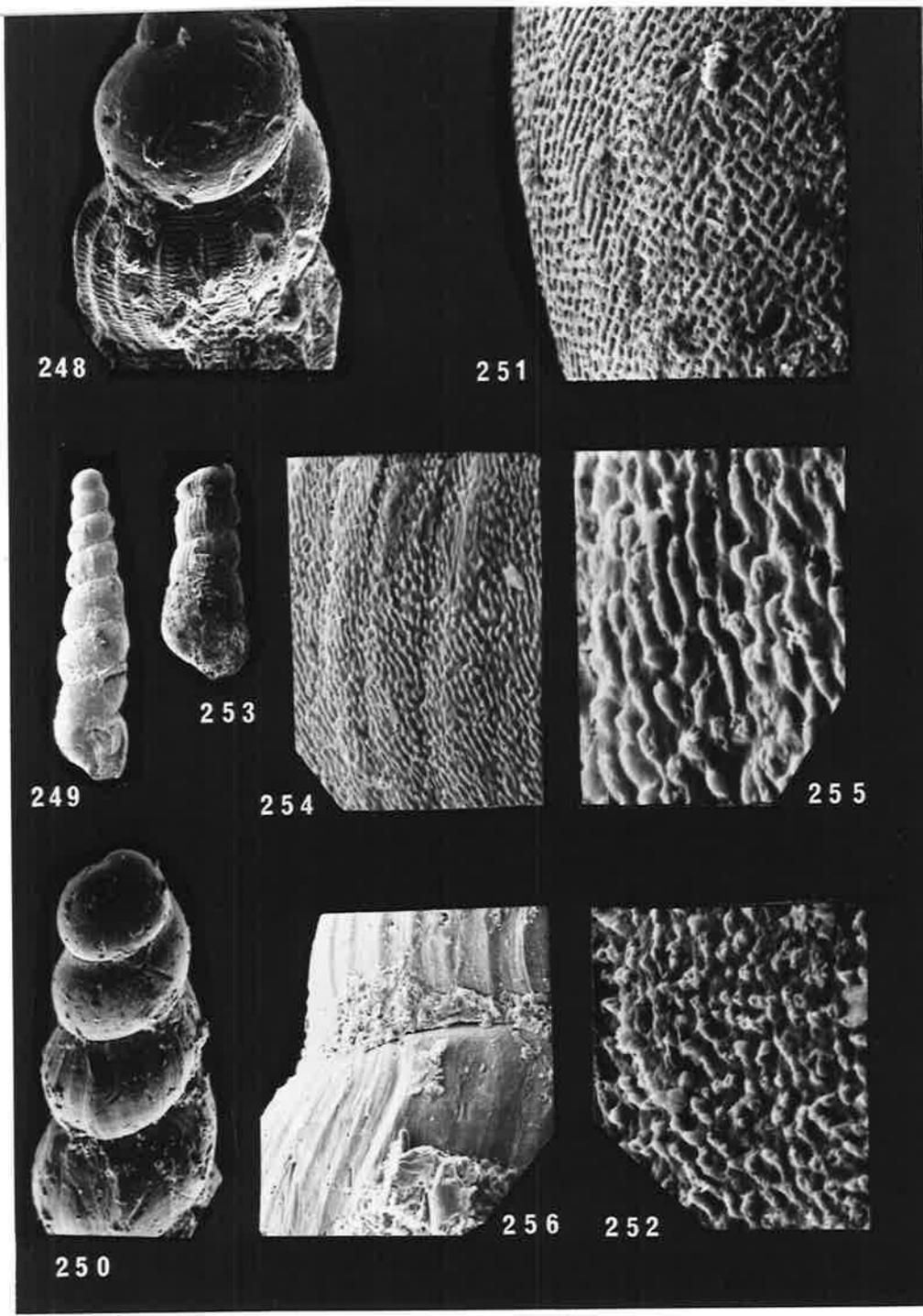
FIG. 230 Siliquaria striata Defrance, SAM P 21280, Uily St. George, Middle Eocene (x 1.25).



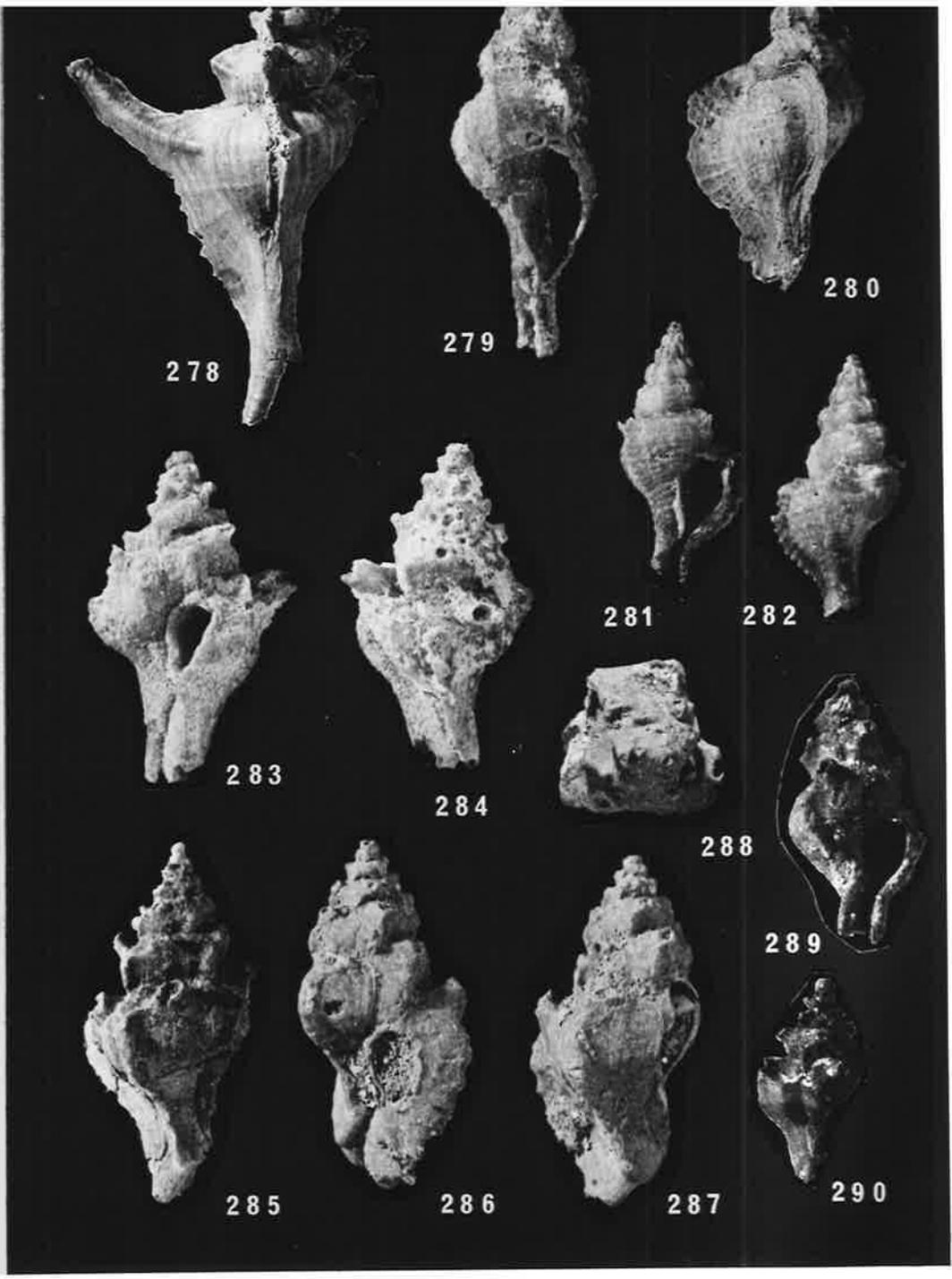
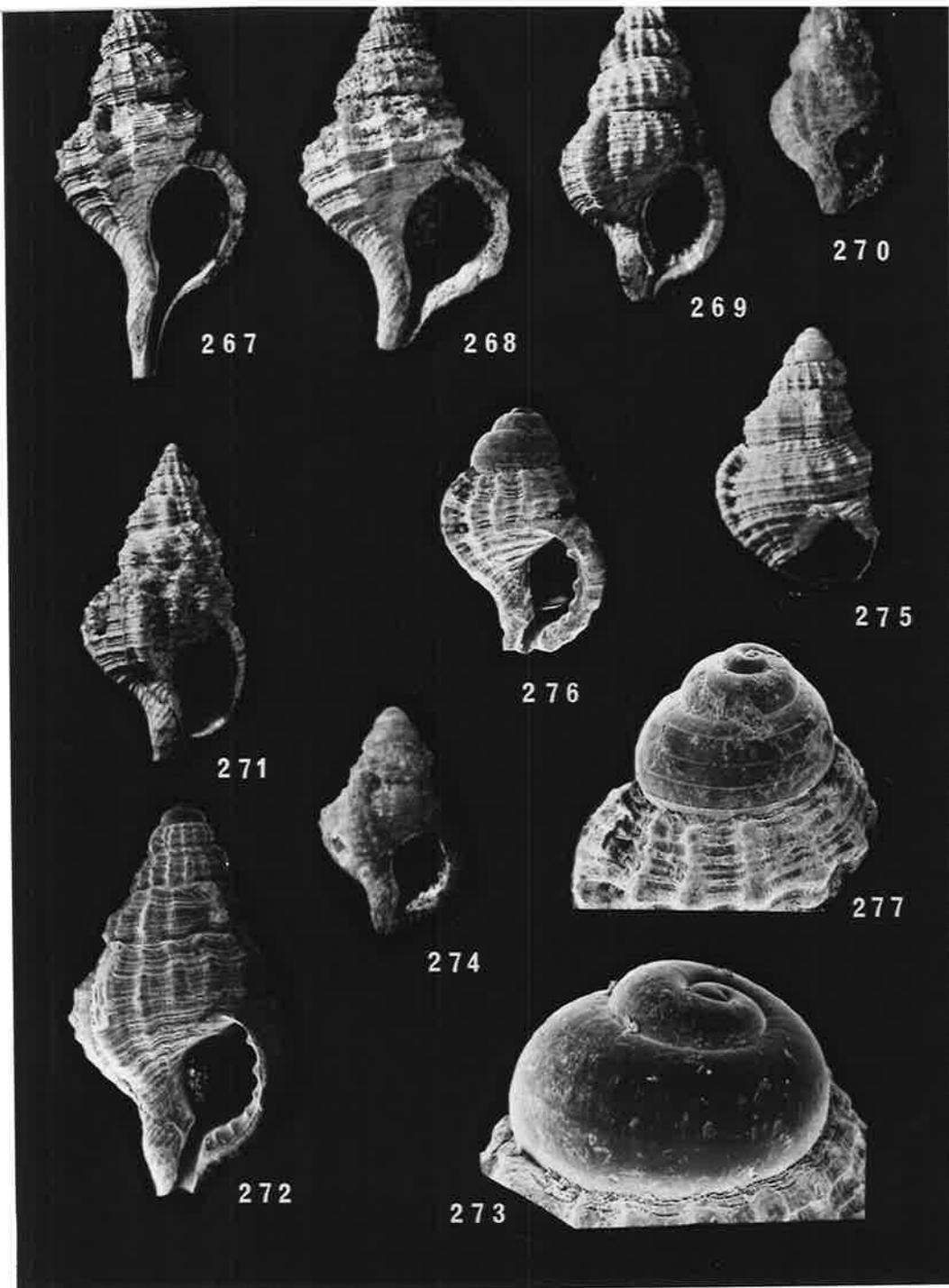
- FIGS. 231-232 Siliquaria occlusa (T. Woods). Paraneotypes, SAM P 21175A-B; axial view (x 2.70).
- FIGS. 233-236 Siliquaria rugosa sp. nov. Holotype, SAM P 21177: 233) axial view (x 2.5); 234) adapical view (x 2.8). Paratypes, SAM P 21178A-B: 235-36) axial view (x 2.4).
- FIGS. 237-241 Niso (Niso) laevigata sp. nov. Holotype, GSSA M 3467-A: 237) axial view (x 4.15). Paratypes: 238) GSSA M 3467-B, axial view (x 4.15); 239) GSSA M 3468, axial view (x 4.15); 240) GSSA M 3469, axial view (x 10); 241) the same, protoconch (x 50).
- FIG. 242 Eulima (Margineulima) striata sp. nov. Holotype, GSSA M 3470, axial view (x 3.8).
- FIGS. 243-247 Aclis (Graphis) costata sp. nov. Holotype, SAM P 21179A: 243) axial view (x 20); 244) spiral and axial ornament (x 100); 245) particular spiral ornament (x 800). Paratypes, axial view (both x 20): 246) SAM P 21179B; 247) SAM P 21179C.



- FIG. 248 Aclis (Graphis) costata sp. nov. Holotype, protoconch (x 120).
- FIGS. 249-256 Aclis (Graphis) laevigata sp. nov. Holotype, SAM P 21180A: 249) axial view (x 20); 250) protoconch (x 70); 251) protoconch ornament, particular (x 700); 252) penultimate whorl, particular pustulae (x 2000). Paratype, SAM P 21180B: 253) axial view (x 20); 254) young adult stage, vermiculations (x 200); 255) vermiculations, particular (x 2000); 256) penultimate/last whorl sutural region, particular (x 120).
- FIGS. 257-260 Lunatia aldingensis (Tate). Holotype, SAM T 1505B: 257) axial view (x 2.0); 258) axial aboral view (x 2.0); 259) adapical view (x 2.0). Paratype, SAM T 1505A: 260) axial view (x 2.0).
- FIGS. 261-262, 266 Polinices nothos sp. nov. Holotype, SAM T 1505J: 261) axial view (x 2.35); 262) protoconch (x 38). Paratype, SAM T 1505K: 266a) axial view (x 3.5); 266b) adapical view (x 3.3).
- FIGS. 263-265 Tanea falsa sp. nov. Holotype, SAM T 1505M: 263) axial view (x 10); 264) protoconch (x 40). Paratype, SAM P 21181: 265) abapical view, tilted (x 9.5).



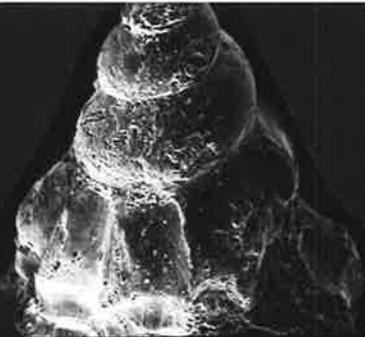
- FIGS. 267-268 Charonia (Austrosassia) cribrosa (Tate). 267) SAM T 503B, holotype (x 1.9); 268) SAM T 503C, paratype (x 2.8).
- FIGS. 269-273 Argobuccinum (Cymatiella) oligostirum (Tate). 269) SAM T 495C, holotype (x 2.55); 270) SAM T 495B, paratype (x 3.15); 271) SAM P 21187, axial view (x 2.65). GSSA M 3392: 272) axial view (x 8.67); 273) protoconch (x 50).
- FIGS. 274-277 Distorsio (Personella) maslinensis sp. nov. 274) SAM P 21188, holotype (x 3.5); 275) GSSA M 3393, paratype (x 3.5). GSSA M 3394, paratype: 276) axial view (x 10); 277) protoconch (x 40).
- FIGS. 278-279 Pterynotus (Pterochelus) manubriatus (Tate): 278) holotype, SAM T 435 B (x 2.85); 279) SAM P 21189, deformed silicified specimen (x 3.6).
- FIGS. 280 Pterynotus (Pterynotus) bifrons (Tate), holotype, SAM P 439A (x 3.0).
- FIGS. 281-282 Pterynotus (Pterochelus) adelaidensis sp. nov., SAM P 21185: 281) adoral axial view (x 3.45); 282) aboral axial view (x 3.45).
- FIGS. 283-285 Typhis (Talityphis) tetraphyllos sp. nov. Holotype, SAM P 21183: 283) adoral axial view (x 3.70); 284) aboral axial view (x 3.70); 285) paratype SAM T 459, aboral axial view (x 2.6).
- FIGS. 286-288 Typhis (Talityphis) waikeriensis sp. nov. Holotype, GSSA M 3395: 286) adoral axial view (x 3.25); 287) aboral axial view (x 3.25); 288) adapical view (x 3.35).
- FIGS. 289-290 Laevityphis (Laevityphis) ludbrookae Keen & Campbell. 289) holotype, SAM T 453A (x 3.1); 290) paratype SAM T 453B (x 3.1).



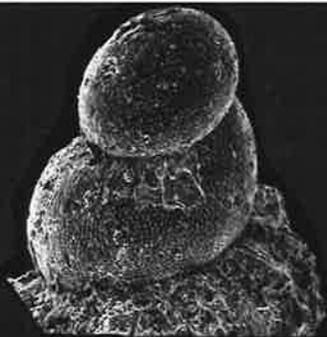
- FIGS. 291-292 Trophonopsis (Trophonopsis) hypsellus (Tate). GSSA M 3396:
291) axial view (x 10); 292) protoconch (x 50).
- FIGS. 293-294 Trophon (Enantimene) monotropis Tate, SAM P 21190:
293) axial view (x 10); 294) protoconch (x 40).
- FIG. 295 Latirus (Broccchitas) aldingensis (Tate). Holotype,
SAM T 570B, axial view (x 3.1).
- FIG. 296 Latirus (Broccchitas) altior sp. nov. Holotype, SAM
P 21191, axial view (x 3).
- FIGS. 297-299 Latirus (Broccchitas) altior m. intermedia m. nov.
297) Holotype, GSSA M 3397 (x 3.0); 298) paratype
GSSA M 3398 (x 3.3); 299) paratype, SAM T 570C (x 3.35).
- FIGS. 300-301 Fusinus (Fusinus) sculptilis (Tate). Paratype, SAM T
478H: 300) axial view (x 10); 301) protoconch (x 40).
- FIGS. 302-306 Baryspira (Gracilispira) ligata (Tate). 302) SAM P
21191, axial view, silicified slightly deformed specimen
(x 3.5); 303) GSSA M 2858, axial view (x 10); 304)
GSSA M 2855 A, juvenile (x 20). GSSA M 2855 B, juvenile:
305) axial view (x 20); 306) protoconch (x 40).
- FIGS. 307-308 Austromitra pumila (Tate) SAM P 21192: 307) axial view
(x 20); 308) protoconch (x 80).
- FIG. 309 Notopeplum protorhysum (Tate). GSSA M 2978, axial view
(x 3.3).
- FIGS. 310-311 Waimatea subcrenularis (Tate). 310) GSSA M 3400A, axial
view (x 3.45); 311) GSSA M 3400 B, axial view (x 3.45).



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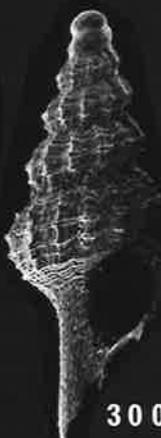
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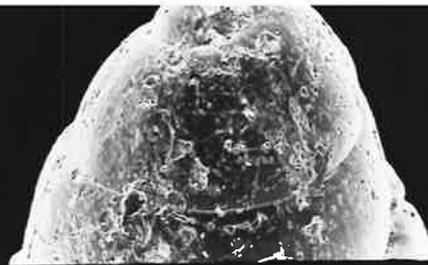
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- FIGS. 312-313 Waimatea subcrenularis (Tate). GSSA M 3399: 312) axial view (x 10); 313) protoconch (x 50).
- FIGS. 314-315 Waimatea complanata (Tate). GSSA M 3401: 314) axial view (x 10); 315) protoconch (x 50).
- FIGS. 316-319 Narona (Iglisella) turriculata (Tate). SAM P 21193: 316) axial view (x 24.35); 317) particular, ornament last whorl (x 60); 318) Protoconch (x 60); 319) particular, protoconch pustulae (x 400).

- FIGS. 320-322 Cottonella mala (Cotton), SAM P 21207A-C, axial view, all x 3.7.
- FIG. 323 Conuginella muna sp. nov., holotype, GSSA M 3417, axial view (x 3.55).
- FIGS. 324-327 Marginella sp. nov. A, GSSA M 3402: 324) axial view (x 10); 325) abaxial lip denticulations (x 30, tilted); 326) protoconch (x 70); 327) abapical vermiculations (x 80).
- FIGS. 328-330 Kaurnaginella tutugka sp. & gen. nov. GSSA M 3403 A-C, axial view (all x 20).



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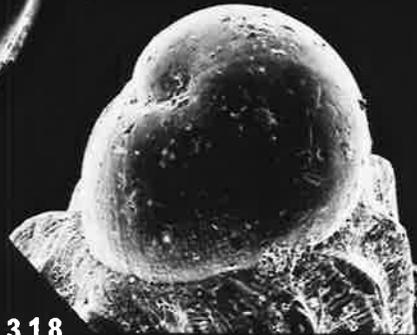
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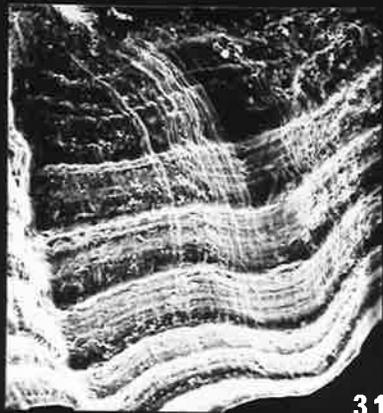
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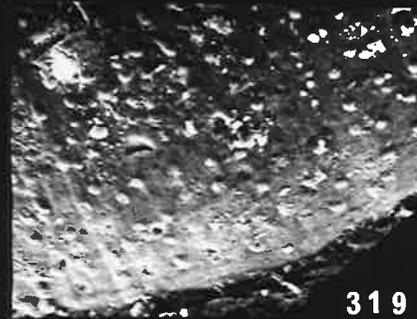
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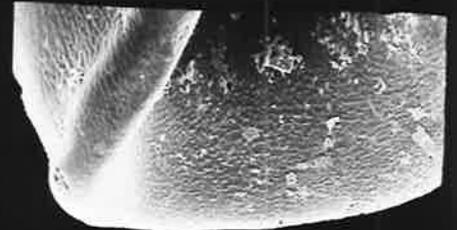
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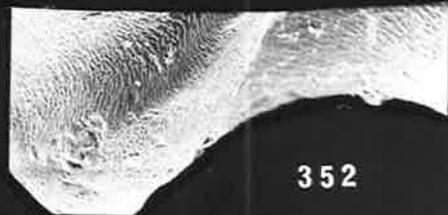
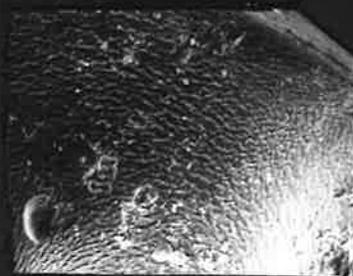
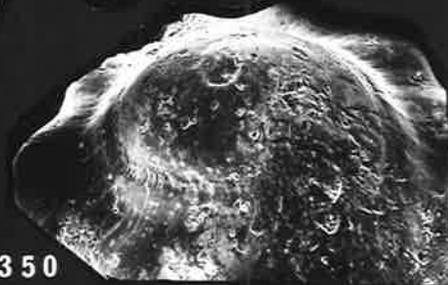
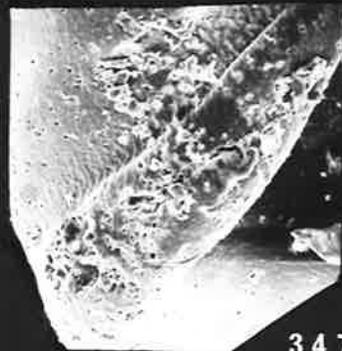
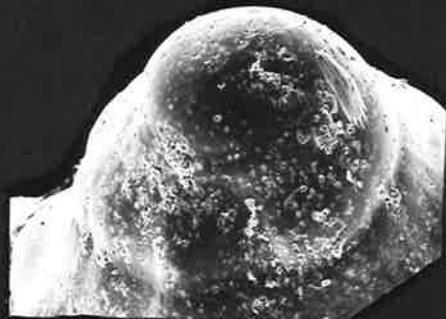
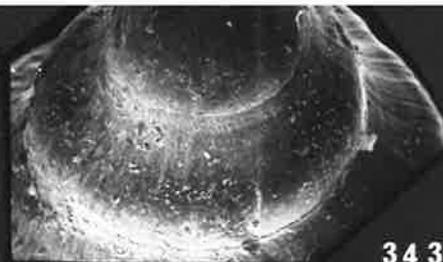
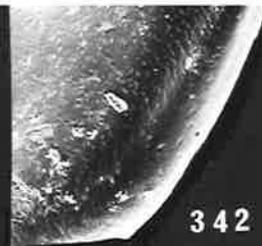
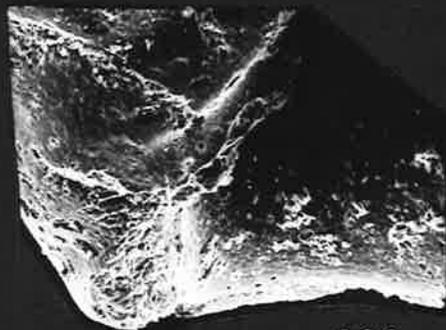
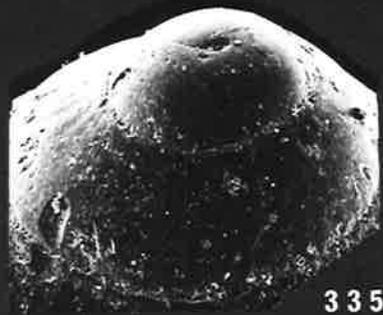
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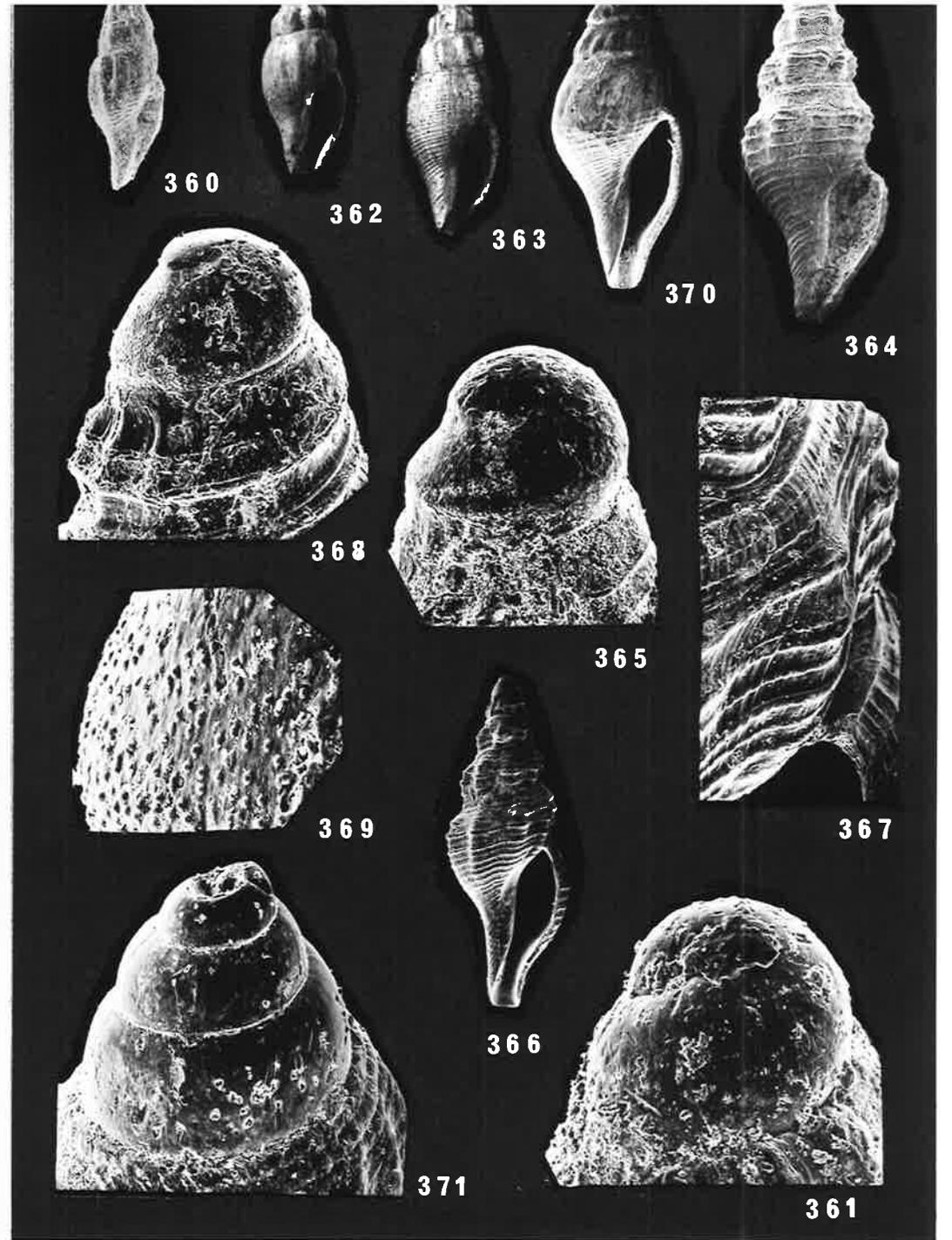
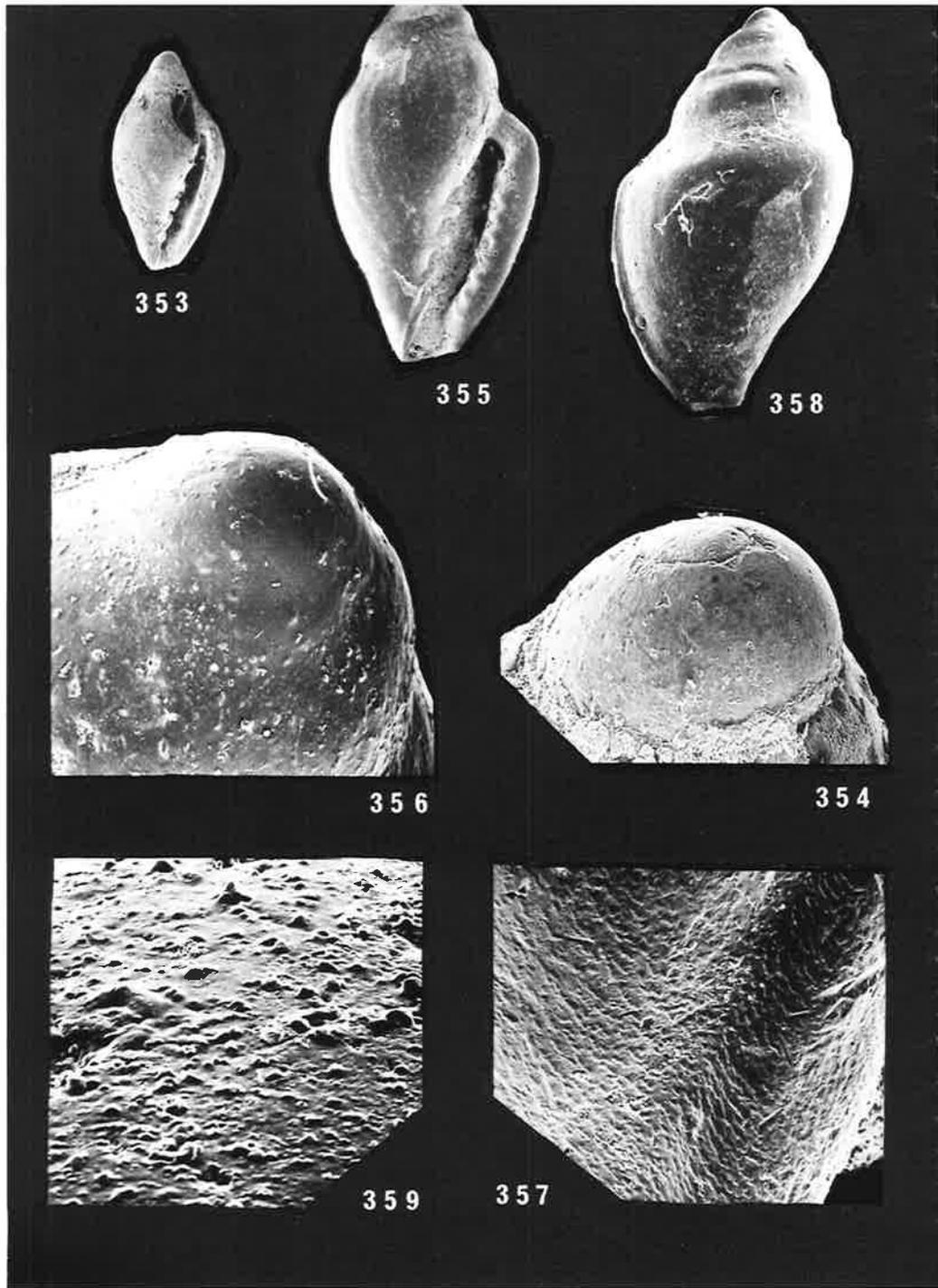
FIGS. 353-359 Marginella (Mioginella) regula (Cotton). SAM P 21194:
353) juvenile, axial view (x 10); 354) protoconch (x 60).
SAM T 656A: 355) axial view (x 10); 356) protoconch (x 50);
357) abapical vermiculations (x 140). SAM T 656B: 358)
axial aboral view (x 10); 359) protoconch pustulations.

FIGS. 360-363 Vexithara citharelloides (Tate). SAM P 21208:
360) axial view, juvenile (x 10); 361) protoconch
(x 80). SAM T 631B, holotype: 362) axial view
(x 3.25). SAM P 21182: 363) axial view (x 3.45).

FIGS. 364-365 Comitas (Comitas) aldingensis (Powell). SAM P 21195:
364) axial view (x 10); 365) protoconch (x 50).

FIGS. 366-369 Comitas sp. nov. GSSA M 3411: 366) axial view (x 10);
367) ornament, particular (x 40); 368) protoconch
(x 50); 369) protoconch pustulae (x 300).

FIGS. 370-371 Rugobela sp. nov. GSSA M 3412: 370) axial view (x 10);
371) protoconch (x 50).



- FIGS. 372-375 Pseudomalaxis (Pseudomalaxis) sp. nov. aff. asculpturatus
Maxwell. GSSA M 3415A-C: 372) GSSA M 3415-A, adapical
view (x 30); 373) GSSA M 3415-B, abapical view (x 30);
374) GSSA M 3415-C, axial view (x 30); 375) the same,
tilted adapical view (x 100).
- FIGS. 376-378 Acrocoelum margaritatum sp. nov. SAM P 21196,
holotype: 376) axial view (x 10); 377) ornament,
particular (x 50); 378) protoconch (x 50).
- FIGS. 379-381 Cirsotrema (Cirsotrema) mariae (Tate). 379) SAM T 778A,
holotype, axial view (x 1.5); 380) SAM T 778E, paratype
(x 1.8); 381) SAM T 778H, paratype (x 1.8).
- FIGS. 382-384 Chemnitzia sp. nov. A. SAM P 21197: 382) axial view
(x 30); 383) protoconch, abapical view (x 80);
384) protoconch, adapical-axial view (x 80).
- FIGS. 385-387 Chemnitzia sp. nov. B. SAM P 21198: 385) axial view
(x 30); 386) protoconch, axial-abapical view (x 80);
387) protoconch, adapical view (x 80).
- FIGS. 388-390 Turbonilla rossiae sp. nov. SAM P 21199, holotype:
388) axial view (x 30); 389) protoconch, abapical
view (x 80); 390) protoconch, axial-adapical view
(x 80).



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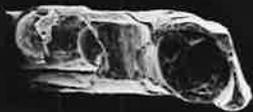
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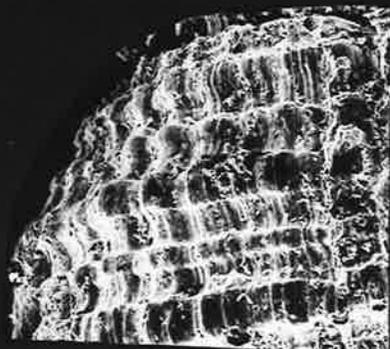
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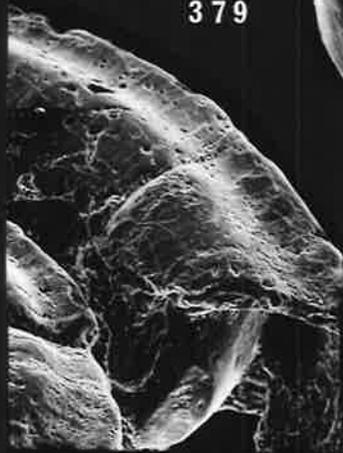
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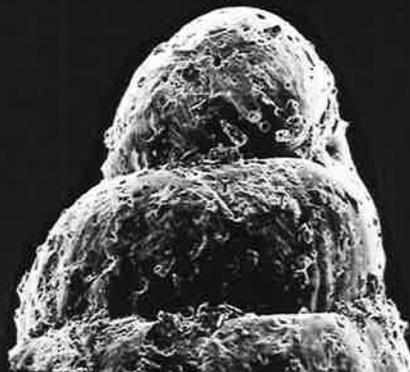
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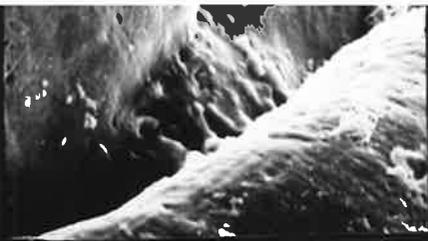
- FIG. 391 Turbonilla kaurna sp. nov. SAM P 21200, holotype, axial view (x 30).
- FIG. 392 Turbonilla sp. nov., SAM P 21201, axial view (x 20).
- FIGS. 393-397 Syrnola (Pachysyrnola) habei sp. nov., GSSA M 3413, holotype: 393) axial view (x 30); 394) protoconch, axial-adapical view (x 100); 395) protoconch, particular nucleus (x 1000). SAM P 21202B, paratype: 396) axial view (x 30); 397) corroded protoconch, abapical view (x 100).
- FIGS. 398-399 Syrnola s.l. sp., GSSA M 3414: 398) axial view (x 30); 399) protoconch (x 160).
- FIGS. 400-401 Tiberia (Cossmannica) maxwelli sp. nov. 400) SAM P 21203-A, holotype, axial view (x 20); 401) SAM P 21203-B, paratype, axial view (x 20).
- FIGS. 402-405 Tiberia (Cossmannica) maxwelli sp. nov.: 402) SAM P 21204-C, paratype, axial view (x 20); 403) SAM P 21204-B, protoconch, axial-adapical view (x 110); 404) the same, particular nucleus (x 1000); 405) SAM P 21204-A, protoconch, abapical view (x 110).
- FIGS. 406-410 Odostomia (Auristomia) sulcata sp. nov.: SAM P 21205-A, holotype: 406) axial view (x 30); 407) protoconch, abapical view (x 110); Paratypes: 408) SAM P 21205-B, axial view (x 30). To note on the right margin, predation drilling; 409) SAM P 21205-C, axial view (x 30); 410) the same, protoconch, adapical view (x 110).



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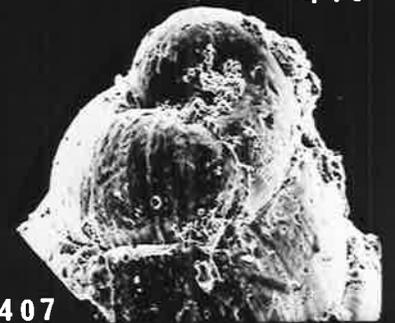
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FIGS. 411-414 Kosugeia costatosulcata sp. nov. Paratype GSSA M 3418, 411) shell (x 24); 412) last whorl (x 42); 413) protoconch (x 60, tilt 45⁰). Holotype, Maslin Bay, SAM P 21209: 414) shell (x 20).

FIGS. 415-420 Inella maxwelli sp. nov. Holotype SAM P 21210: 415) shell (x 34); 416) last whorl (x 52); 417) protoconch (x 54, tilt 45⁰). Paratype SAM P 21210-A: 418) shell (x 20). Paratype SAM P 21210B: 419) shell (x 20); 420) last whorl (x 38).

FIGS. 421-422 Viriola sp. nov. GSSA M 3419: 421) shell (x 10); 422) last whorl (x 30).

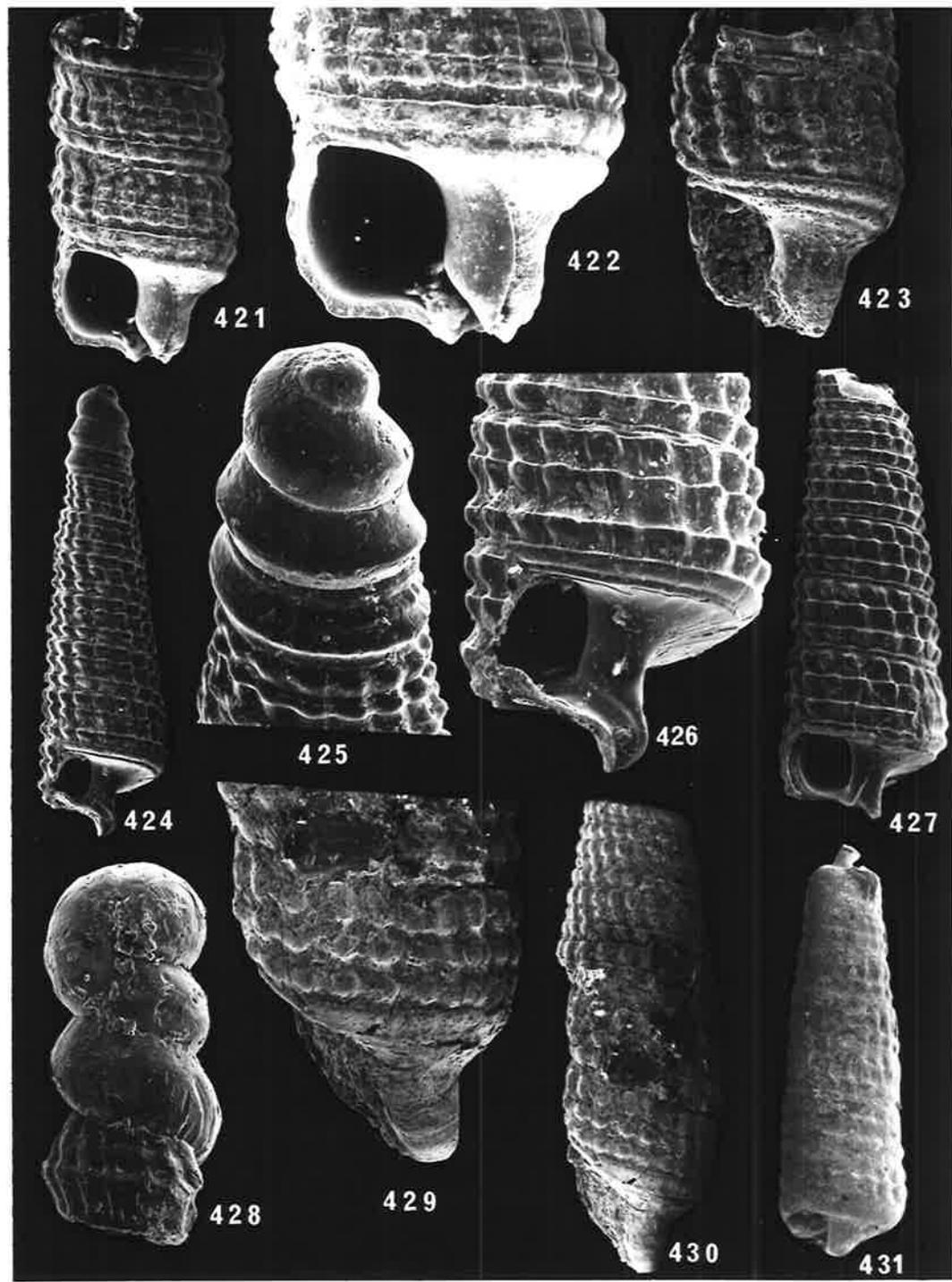
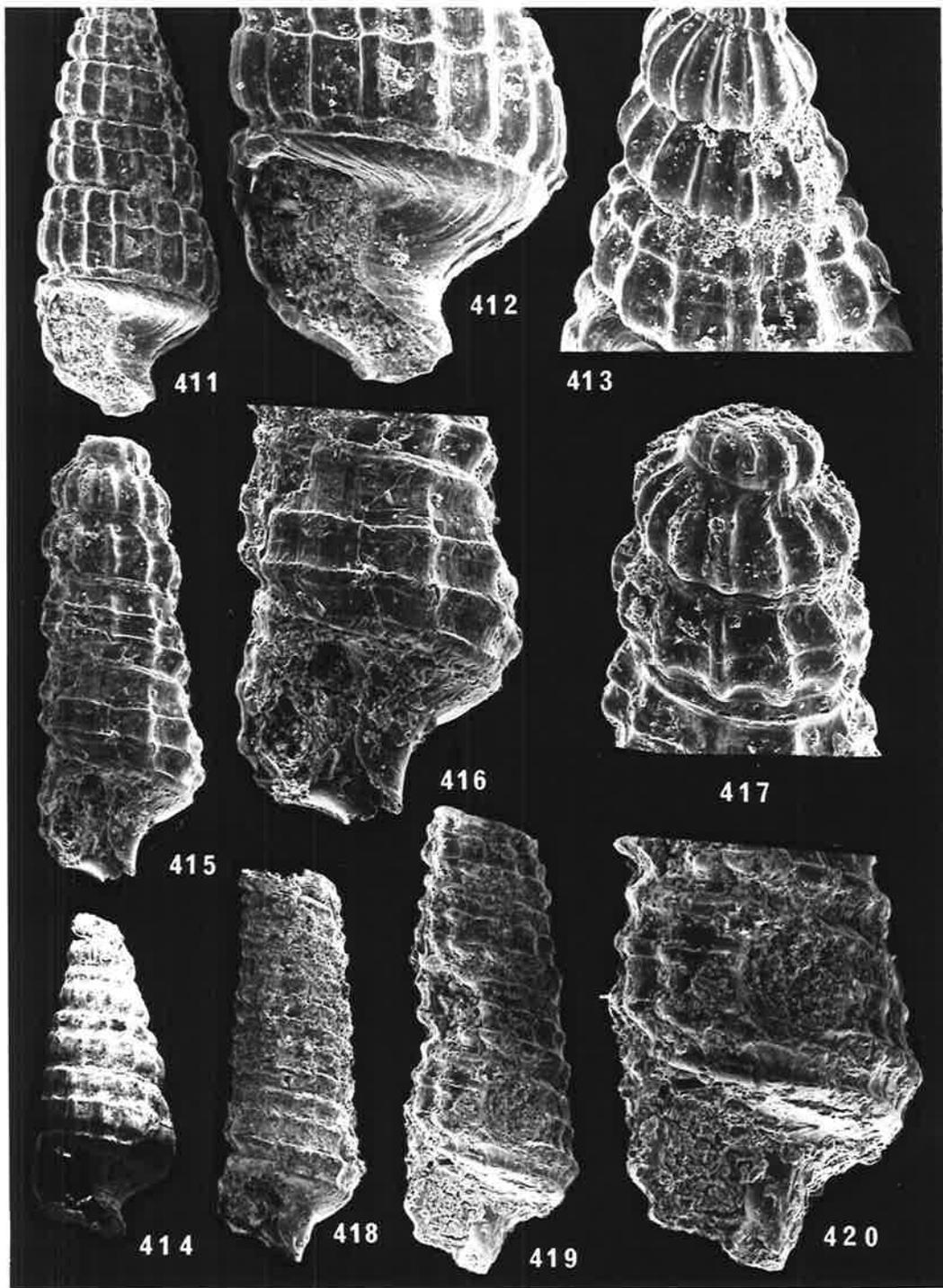
FIG. 423 Isotriphora sp. nov. SAM P 21211, shell (x 20).

FIGS. 424-427 Triphora (Ogivia) trirostrata sp. nov. Holotype GSSA M 3420: 424) shell (x 10); 425) protoconch (x 50, tilt 45⁰); 426) last whorl (x 30). Paratype GSSA M 3421: 427) shell (x 20).

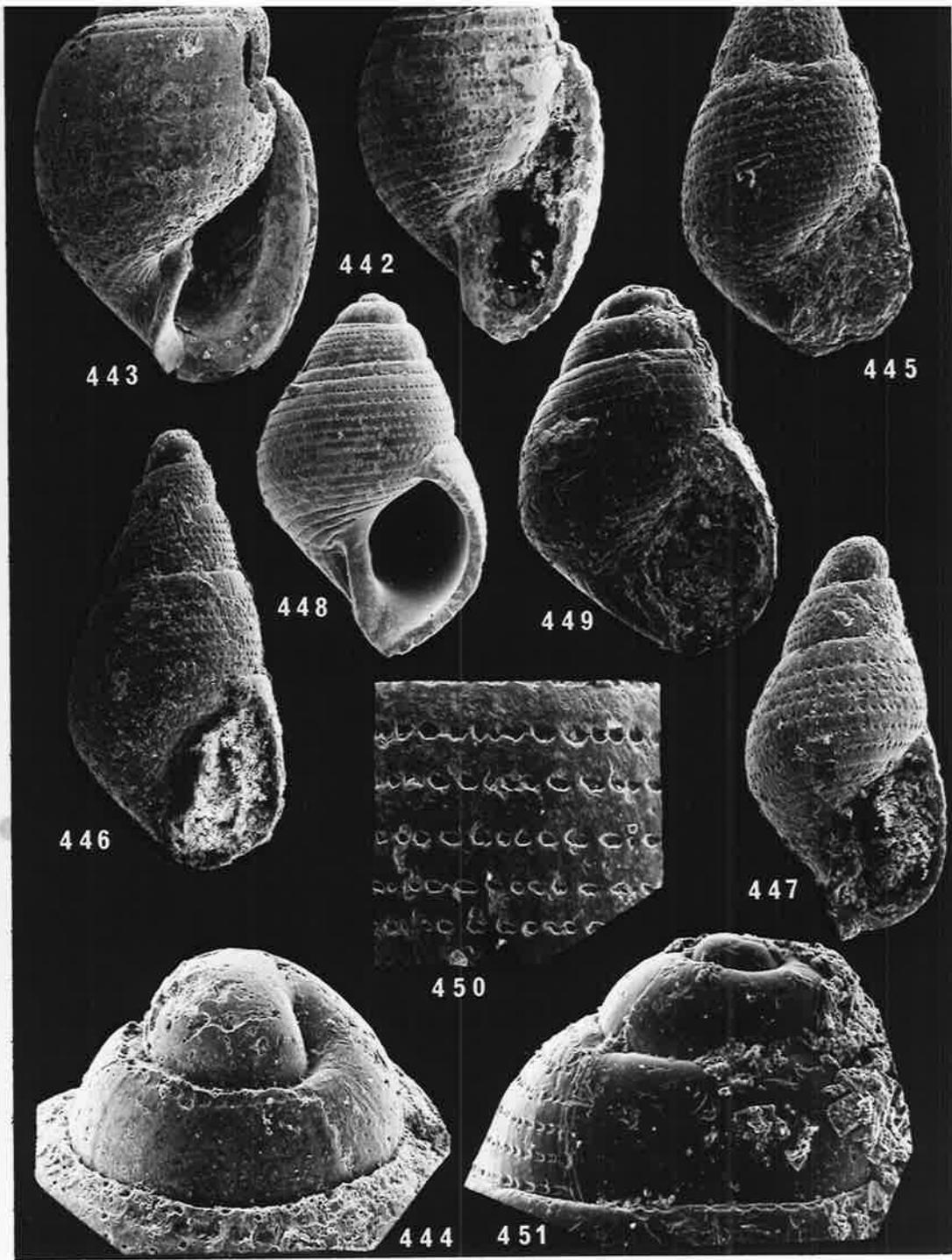
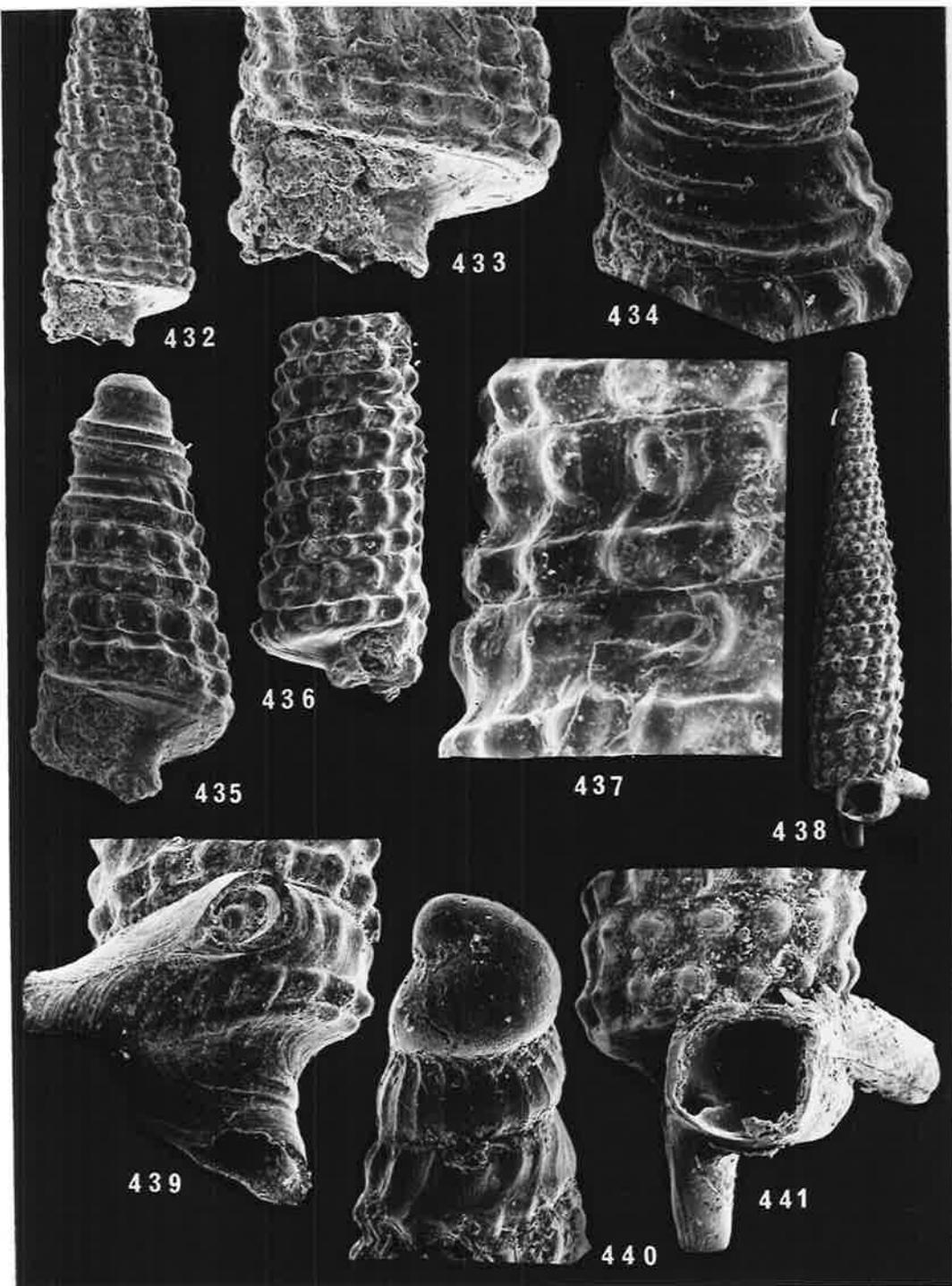
FIG. 428 Triphora s.l. sp. nov. A, SAM P 21212, protoconch (x 40).

FIGS. 429-430 Triphora s.l. muna sp. nov. Paratype SAM P 21213A: 429) deformed last whorl (x 20); 430) shell (x 10).

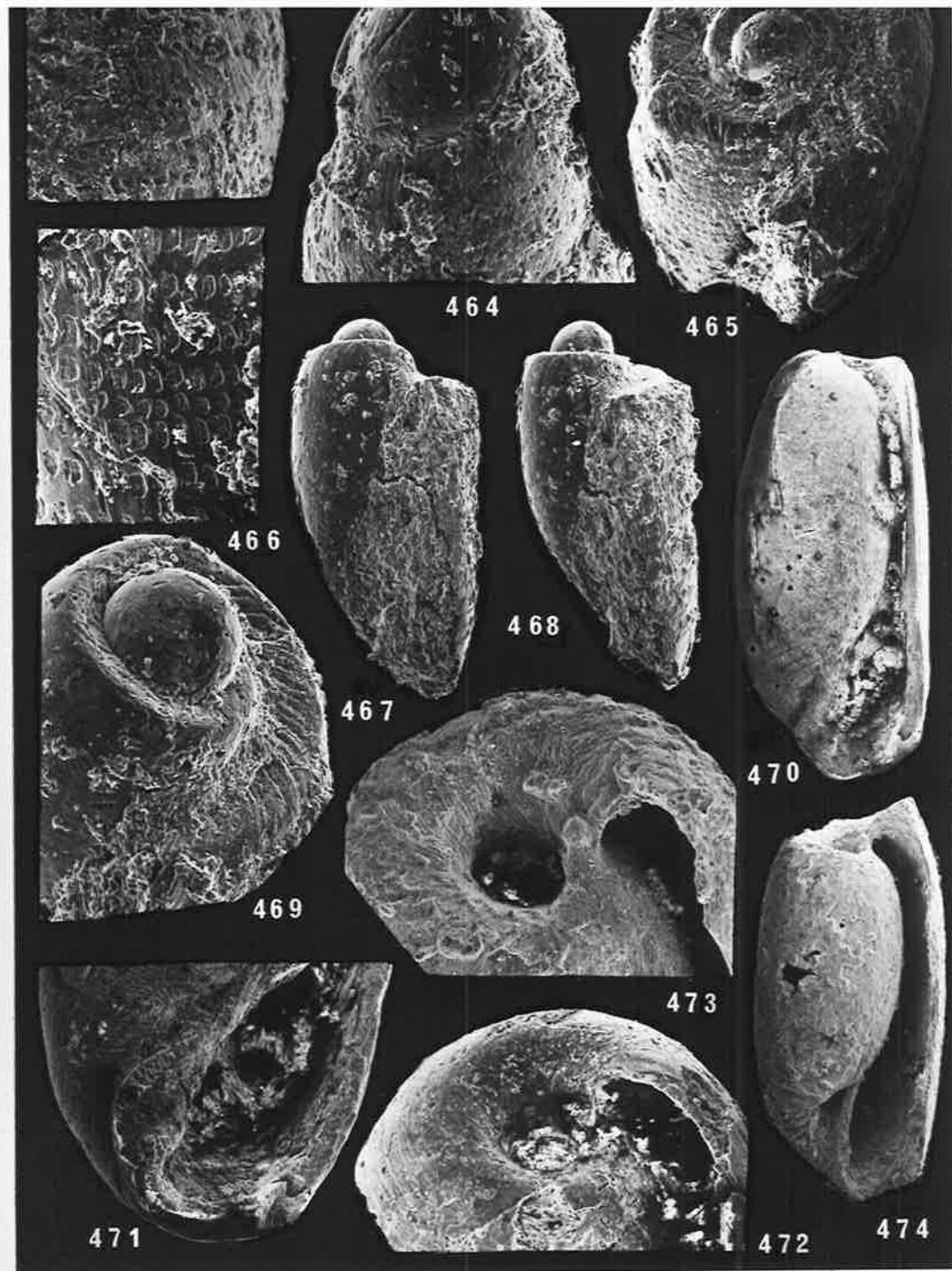
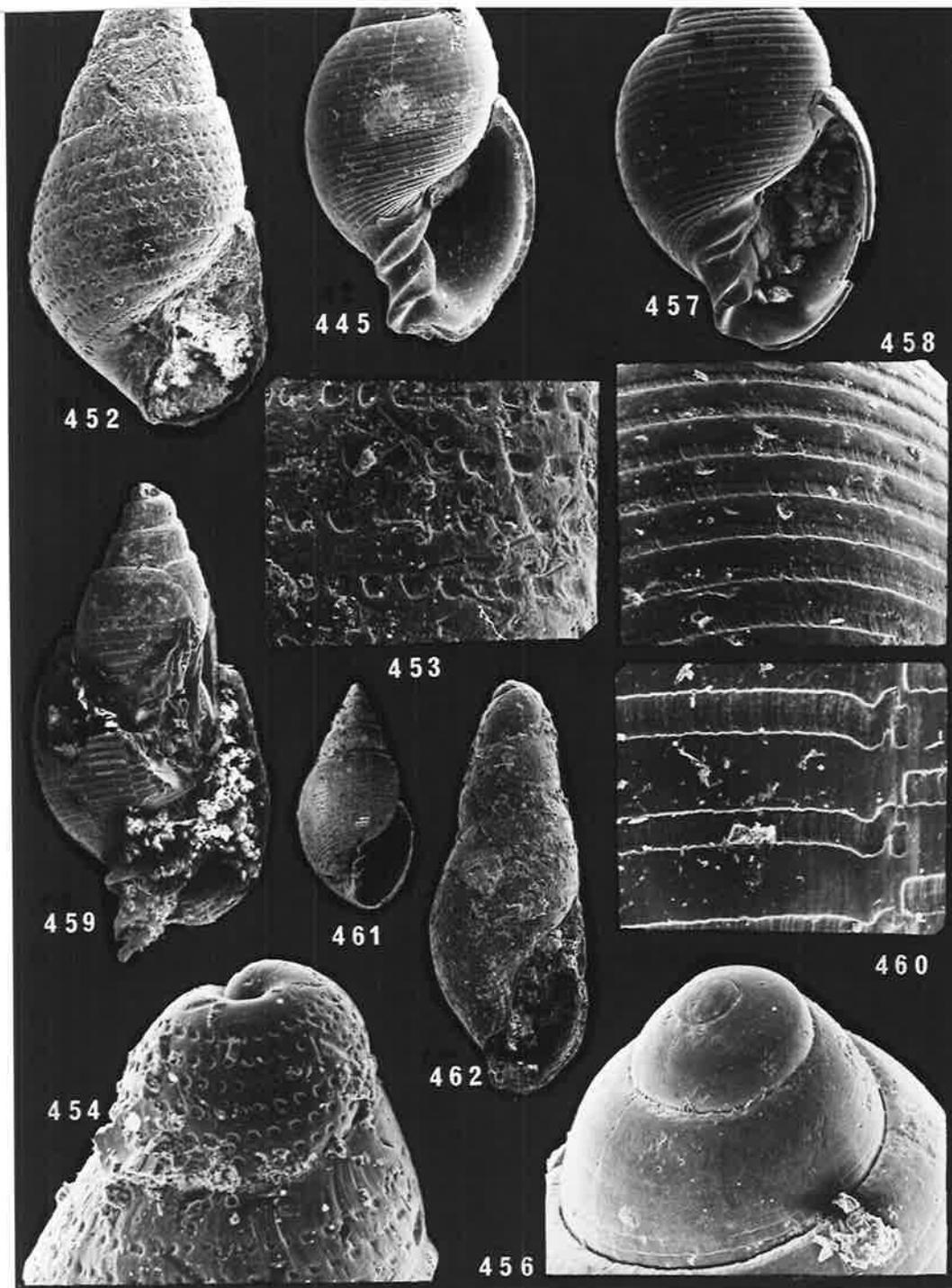
FIG. 431 Triphora s.l. ?sp. nov. B, GSSA M 3423, worn shell (x 10).



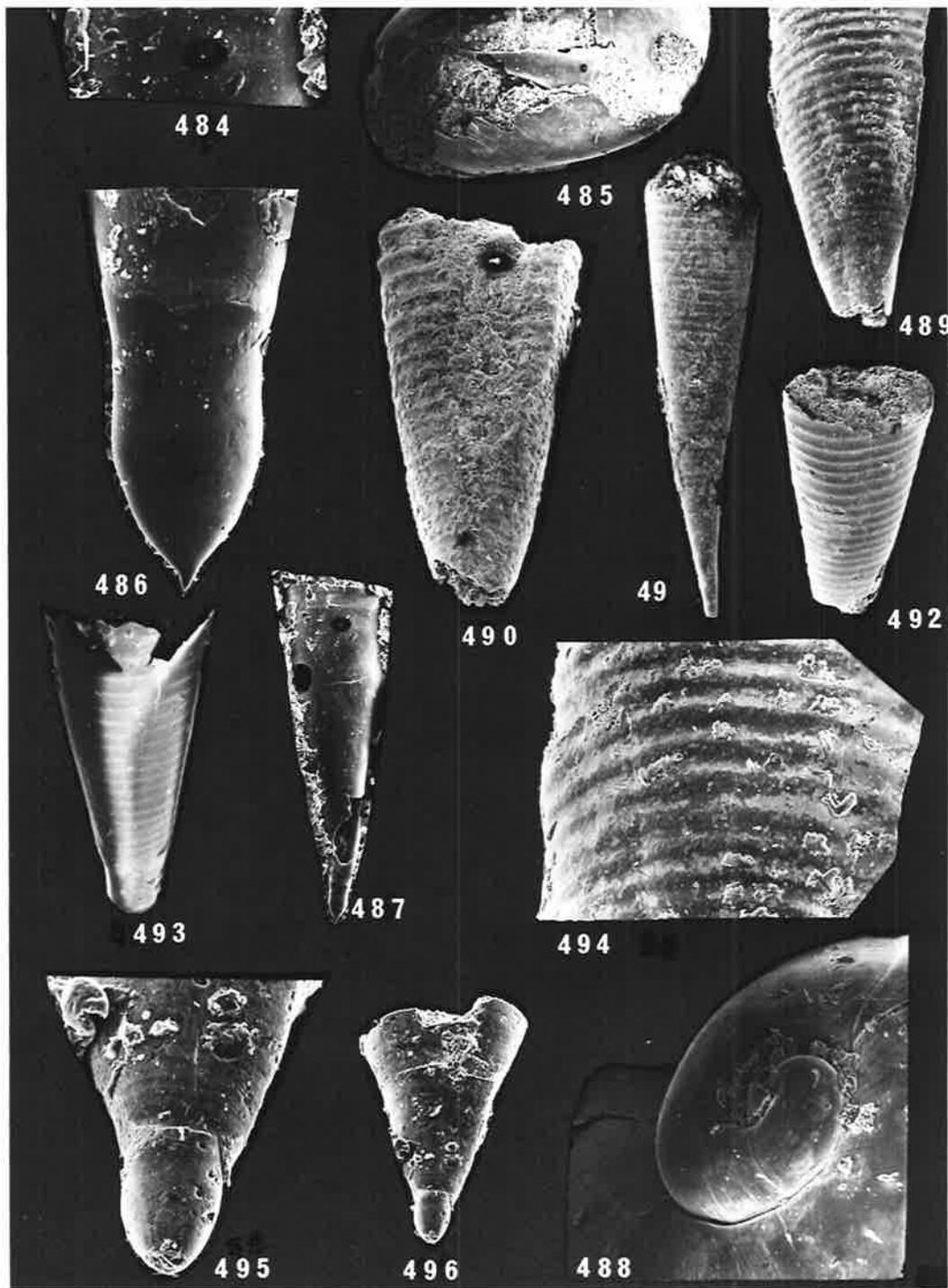
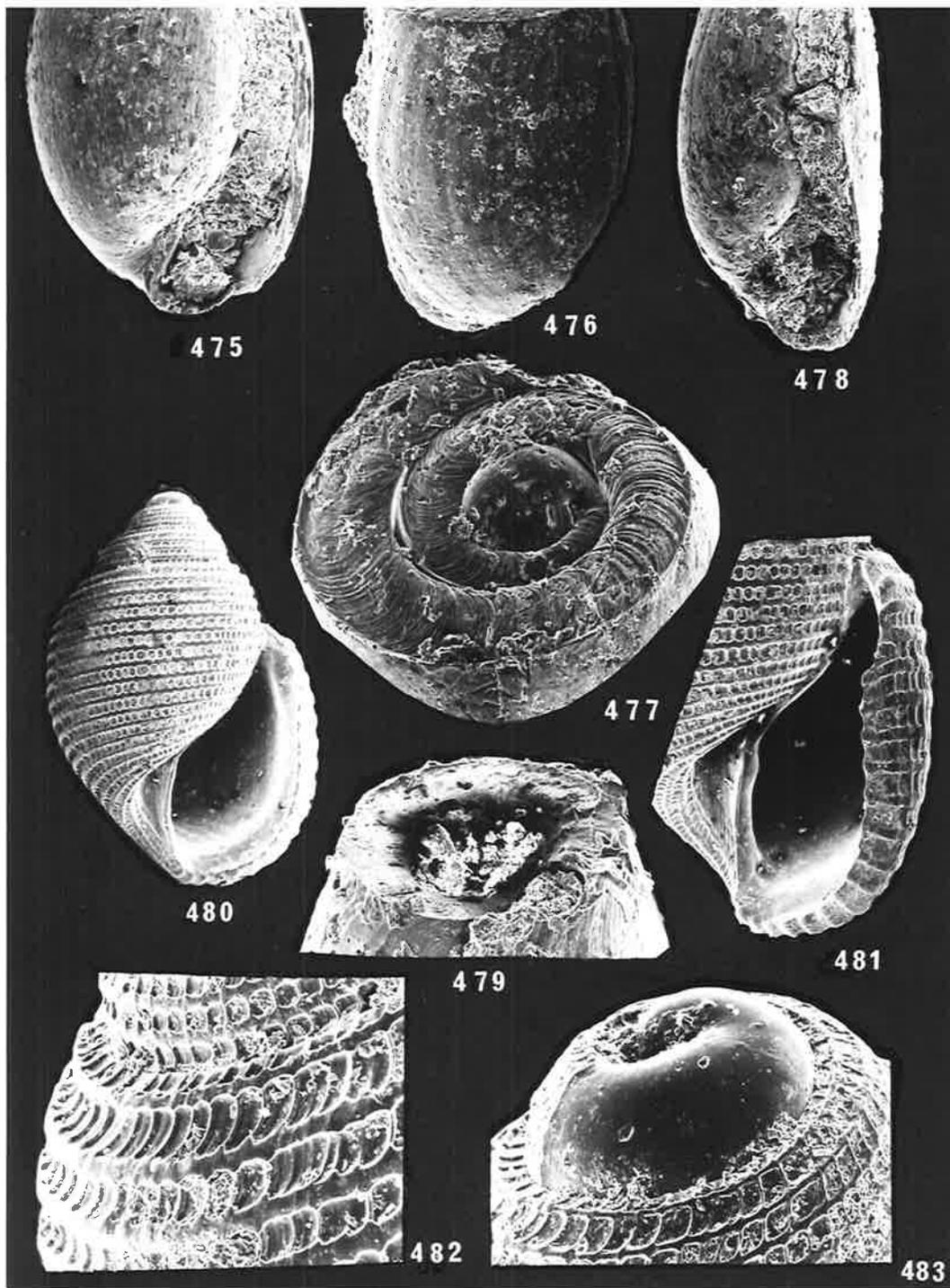
- FIGS. 432-435 Triphora s.l. muna, sp. nov. Holotype, SAM P 21213:
432) axial view (x 20); 433) last whorl (x 40); 434)
protoconch (x 80). Paratype, GSSA M 3422; 435) axial view
(x 40).
- FIGS. 436-437 Triforis (Granulotriforis) sp. nov. GSSA M 3424: 436)
shell (x 20); 437) particular ornament and whorl (x 70).
- FIGS. 438-441 Triforis (Granulotriforis) epallaxa (Verco) SAM P
21214, Great Australian Bight, 50-120 fms (Holocene):
438) shell (x 10); 440) protoconch (x 60, tilt 45⁰);
441) last whorl (x 30). Topotype SAM P 21215, Cape
Jaffa, 300 fms. 439) last whorl, particular adapical
channel (x 30).
- FIGS. 442-444 Acteon subscalatus Cossmann. SAM P 21216: 442) axial view
(x 20). SAM P 21217: 443) axial view (x 30); 444)
protoconch (x 60, tilt 45⁰).
- FIGS. 445-447 Kaurnacteon elevatus gen. & sp. nov. (all x 30): 445)
paratype SAM P 21218B; 446) paratype SAM P 21218E;
447) paratype SAM P 21218A.
- FIGS. 448-451 ?Kleinacteon dubius sp. nov.: 448) paratype GSSA M 3425,
axial view (x 30). Holotype, SAM P 21219: 449) axial
view (x 40); 450) ornament (x 100); 451) protoconch
(x 80, tilt 45⁰).



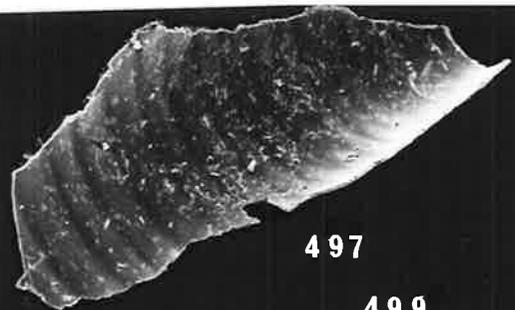
- FIGS. 452-454 Kaurnacteon elevatus gen. & sp. nov. Holotype SAM P 21220: 452) shell (x 30); 453) ornament (x 100); 456) protoconch (x 120, tilt 45⁰).
- FIGS. 455-458 Tornatellaea (Tornatellaea) minutissima sp. nov. Paratype GSSA M 3426: 455) axial view (x 30); 456) protoconch (x 80, tilt 45⁰). Holotype GSSA M 3427: 457) shell (x 30); 458) ornament (x 100).
- FIGS. 459-461 Tornatellaea (Triploca) ligata (Tate). Paralectotype SAM 1758 B: 459) axial view (x 10); 460) ornament (x 120). Lectotype SAM T 1758 A; 467) axial view (x 4.7).
- FIG. 462 Tenuiacteon acicularis sp. nov. Holotype SAM P 21221: 462) (x 30).
- FIGS. 463-464 Tenuiacteon acicularis sp. nov. Holotype: 463) ornament (x 100); 464) protoconch (x 80, tilt 45⁰).
- FIGS. 465-469 Acteocina scalarum sp. nov. Paratype SAM P 21222A: 465) apical view (x 40); 466) ornament (x 130). Holotype SAM P 21222: 467-468) shell two views (x 30); 469) protoconch (x 70, tilt 45⁰).
- FIGS. 470-472 Cylichna (Cylichnania) callosa (Tate & Cossmann). SAM P 21223: 470) axial view (x 10); 471) peristome (x 20). SAM P 21224; 472) adapical view (x 50, tilt 45⁰).
- FIGS. 473-474 Cylichna (Cylichna) cf. angustata (Tate & Cossmann). 473) SAM P 21225, adapical view (x 50, tilt 45⁰); 474) SAM P 21226, axial view (x 20).



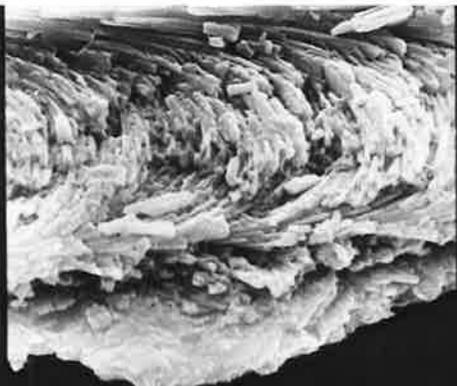
- FIGS. 475-477 Retusa (Decorifer) crassa sp. nov. 475) Holotype SAM P 21227 (x 30); 476) paratype SAM P 21227A, dorsal view (x 20); 477) Paratype SAM P 21227B, adapical view (x 70, tilt 45°).
- FIGS. 478-479 Retusa (Decorifer) gracilis sp. nov. Holotype SAM P 21228, 478) axial view (x 20); 479) adapical view (x 70, tilt 45°).
- FIGS. 480-483 Obrussenia alveolata (Tate). Holotype SAM T 713: 480) shell (x 10); 481) peristome and umbilicus (x 20, tilt 30°); 482) particular ornament (x 80, tilt 45°); 483) protoconch (x 70, tilt 45°).
- FIGS. 484-487 Bovicornu robbai sp. nov. Holotype SAM P 21229: 484) peristome (x 70); 485) the holotype into the matrix of Ectosinum sp. nov. (x 10); 486) protoconch (x 200); 487) shell (x 30);
- FIGS. 485,488 Sinum (Ectosinum) sp. nov. Adelaide Bore, SAM P 21229: 485) axial view (x 10); 488) protoconch (x 50, tilt 45°).
- FIGS. 489-496 Praehyalocylis annulata (Tate). 489) SAM P 21230A, squashed shell (x 10); 490) SAM P 21230B, squashed shell (x 10); 491) SAM P 21230C, compressed shell (x 10, tilt 45°); 492) SAM P 21230D, particular protoconch (x 100); 494) the same, juvenile (x 50); 495) SAM P21230A, particular ornament (x 30); 496) SAM P 21230E, shell (x 10). SAM T 214: 497) axial view (x 10).



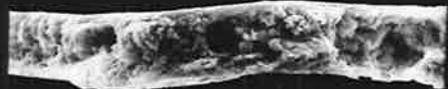
- FIGS. 497-500 Praehyalocylis annulata (Tate). SAM T 214: 497) shell fragment, inner ornament (30); 498) the same, particular, cone-in-cone structure (x 3000); 499) grouped tubules (x 800); 500) solitary tubule (x 5000).
- FIG. 501 Bovicornu robbai sp. nov. SAM P 21229, particular shell structure (x 4000).
- FIG. 502 Pronucula (Pronucula) tatei (Finlay), SAM P 21251, specimen with abraded surface (x 20).
- FIGS. 503-508 Nuculana (Saccella) chapmani (Finlay). SAM P 21231-A: 503) RV, outer (x 10); 504) particular anterior ornament (x 100); 505) particular, ventral ornament (x 100). SAM P 21231-B: 506) RV, inner (x 10); 507) particular, umbonal cardinal region; 508) particular, posterior teeth (x 90).
- FIGS. 509-513 Nuculana (Ledella) leptorhyncha (Tate). SAM P 21232-A: 509) LV, outer (x 10); 510) particular ornament (x 130). SAM P 21232-B: 511) LV, inner (x 10); 512) umbonal cardinal region (x 60); 513) particular, posterior teeth (x 100).
- FIG. 514 Nuculana (Poroleda) sp. nov. SAM P 21230, LV, bivalved specimen (x 10).



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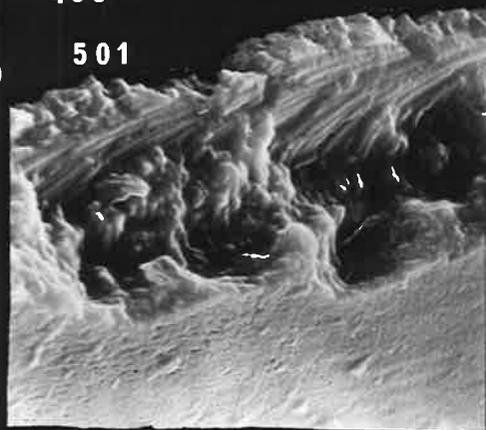
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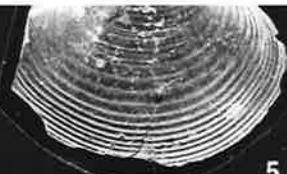
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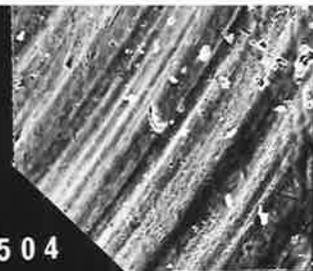
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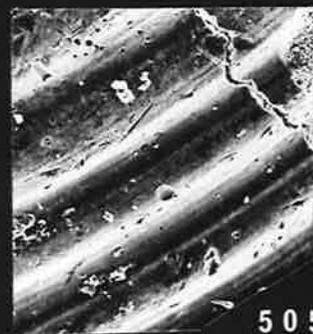
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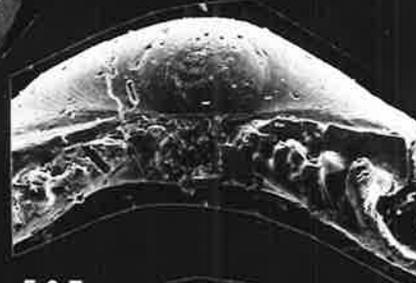
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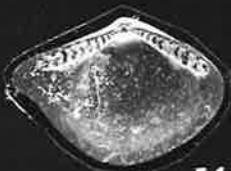
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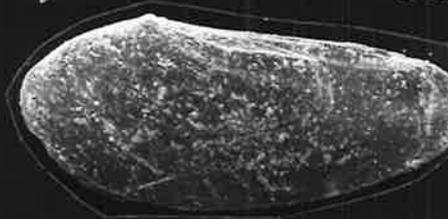
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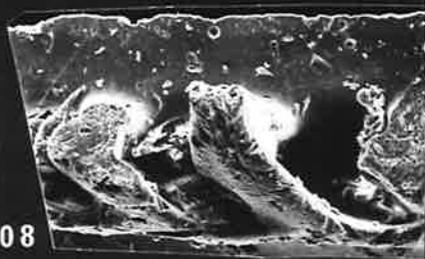
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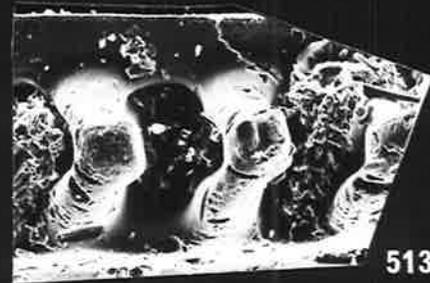
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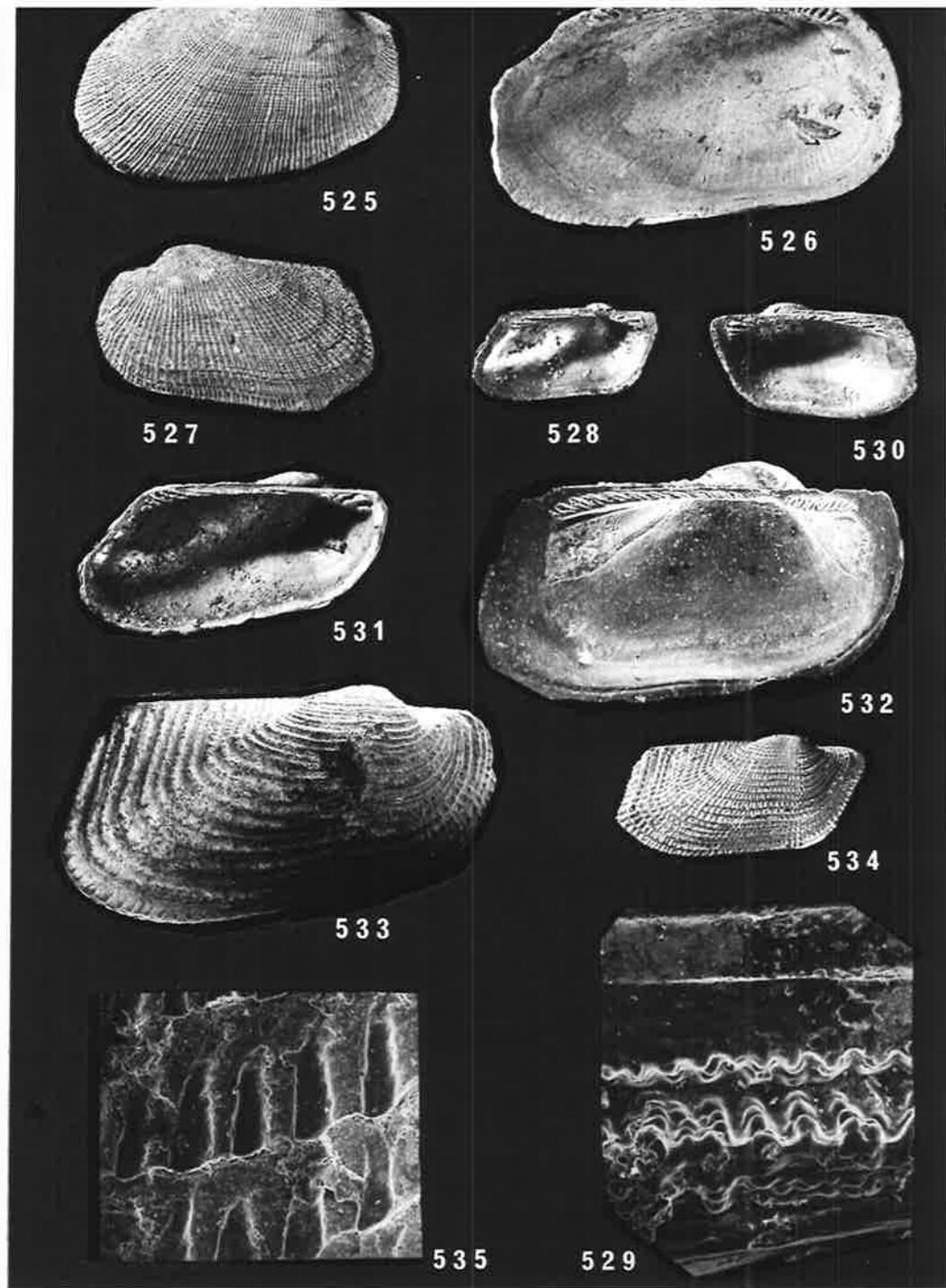
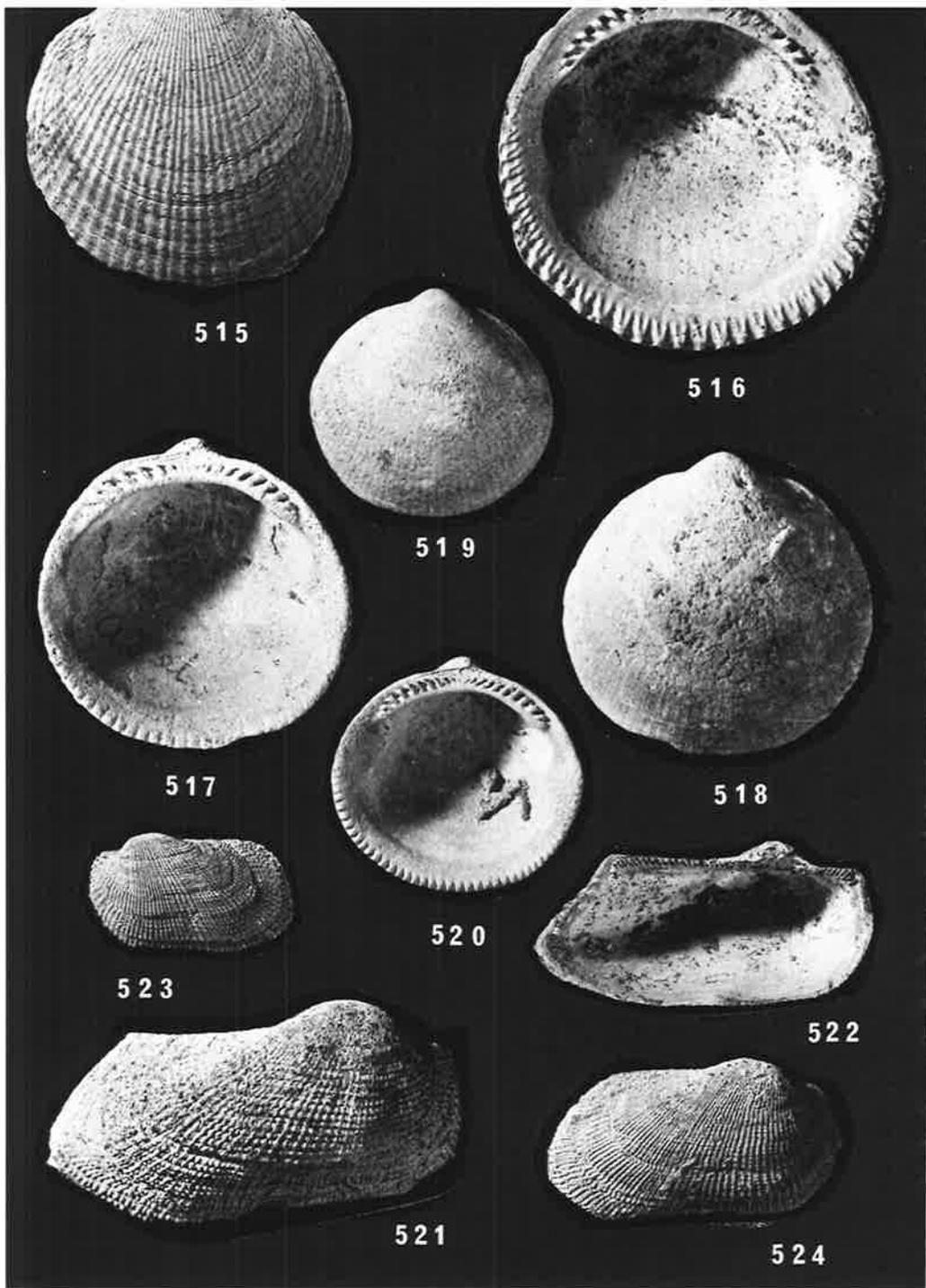


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- FIGS. 515-516 Glycymeris (Glycymeris) lenticularis (Tate):
515) SAM T 1011-A, holotype, LV, outer (x 1.7);
516) SAM T 1011-D, paratype, RV, inner (x 2 .65)
- FIGS. 517-520 Glycymeris (Glycymeris) kurna sp. nov. SAM T
1055-U, holotype, LV: 517) inner (x 2); 518) outer
(x 2). SAM T 1055-W, paratype, LV: 519) outer
(x 2); 520) inner (x 2).
- FIGS. 521-522 Arca (Arca) pseudonavicularis Tate: 521) holotype,
SAM T 1057, RV, outer (x 2.25); 522) paratype,
SAM T 1057-B, LV, inner (x 2.25).
- FIGS. 523-524 Barbatia (Barbatia) limatella Tate: 523) paratype
SAM T 1048-K, LV, outer (x 2.1); 524) paratype,
SAM T 1048-J, RV, outer (x 2.1)
- FIGS. 525-527 Barbatia (Barbatia) limatella Tate. 525) SAM T
1048-E, paratype, RV, outer (x 2); 526) SAM T
1048-D, paratype, RV, inner (x 1.75); 527) SAM T
1048-C, paratype, RV, outer (x 1.95).
- FIGS. 528-535 Porterius (Notogrammatodon) inexpectatus Maxwell
528) SAM T 1056-R, LV, inner (x 2.7); 529) the same,
posterior hinge, particular crenulated teeth (x 100);
530) SAM T 1056-U, RV, inner (x 3.2); 531) SAM T
1056-S, LV, inner, (x 2.7); 532) GSSA M 3437, LV,
inner (x 10); 533) NZGS 9481, topotype, outer, bivalved
specimen RV view (x 9); 534) SAM T 1056-T, RV, outer
(x 3.2); 535) the same, particular ornament (x 40).



FIGS. 536-543 Porterius (Ludbrookella) spinosa subgen. & sp. nov.:

536) GSSA M 3433, paratype, RV fragment (x 2.75).
GSSA M 2820-A, holotype, LV; 537) outer (x 3.85);
538) inner (x 3.85). GSSA M 2820-B, RV, paratype:
539) outer (x 3.85); 540) inner (x 3.85). Paratypes:
541) GSSA M 3427-A, LV, outer (x 3.85); 542) GSSA M
3427-B, LV, outer (x 3.85); 543) GSSA M 3427-D,
particular RV posterior hinge (x 100).

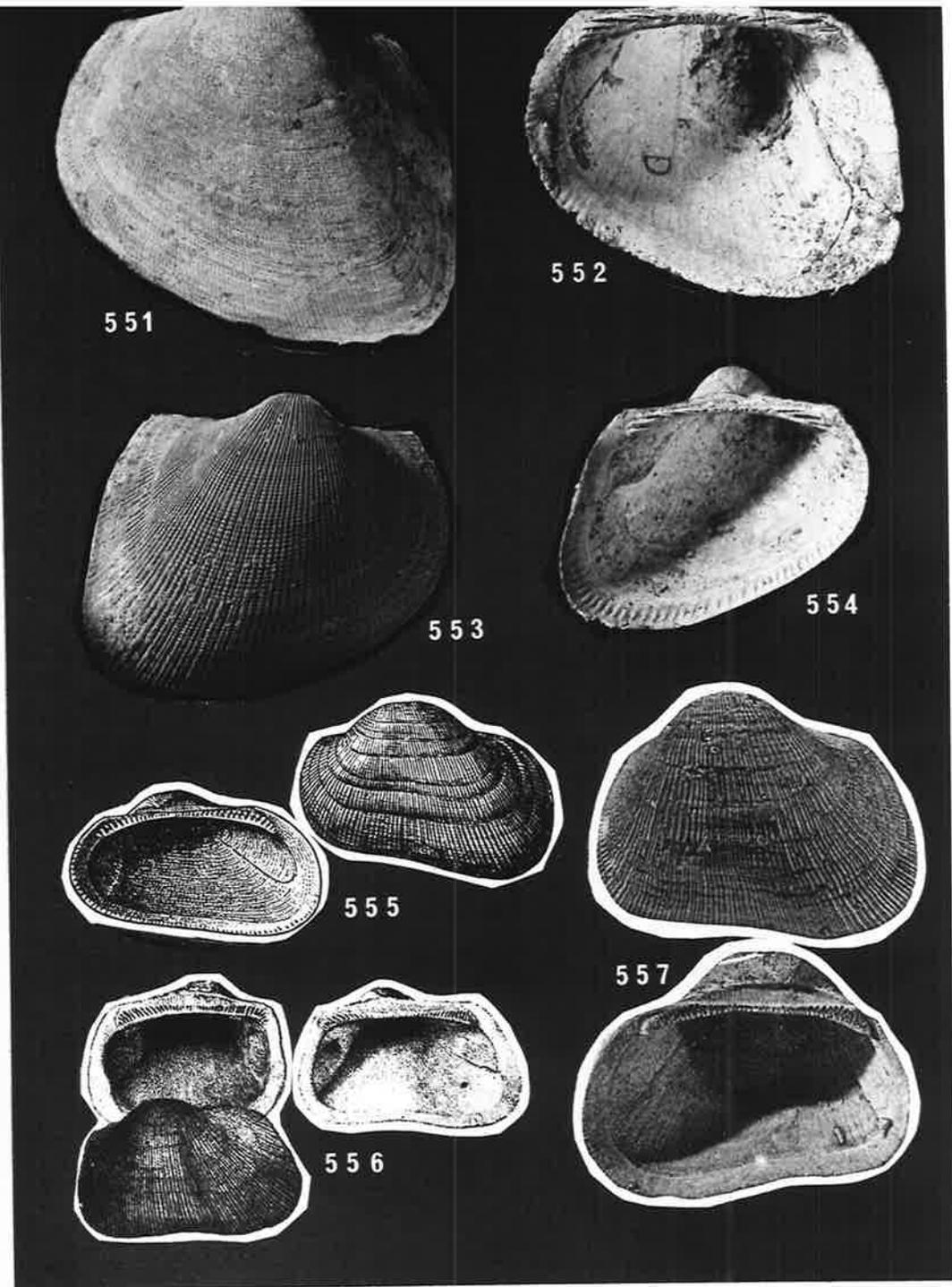
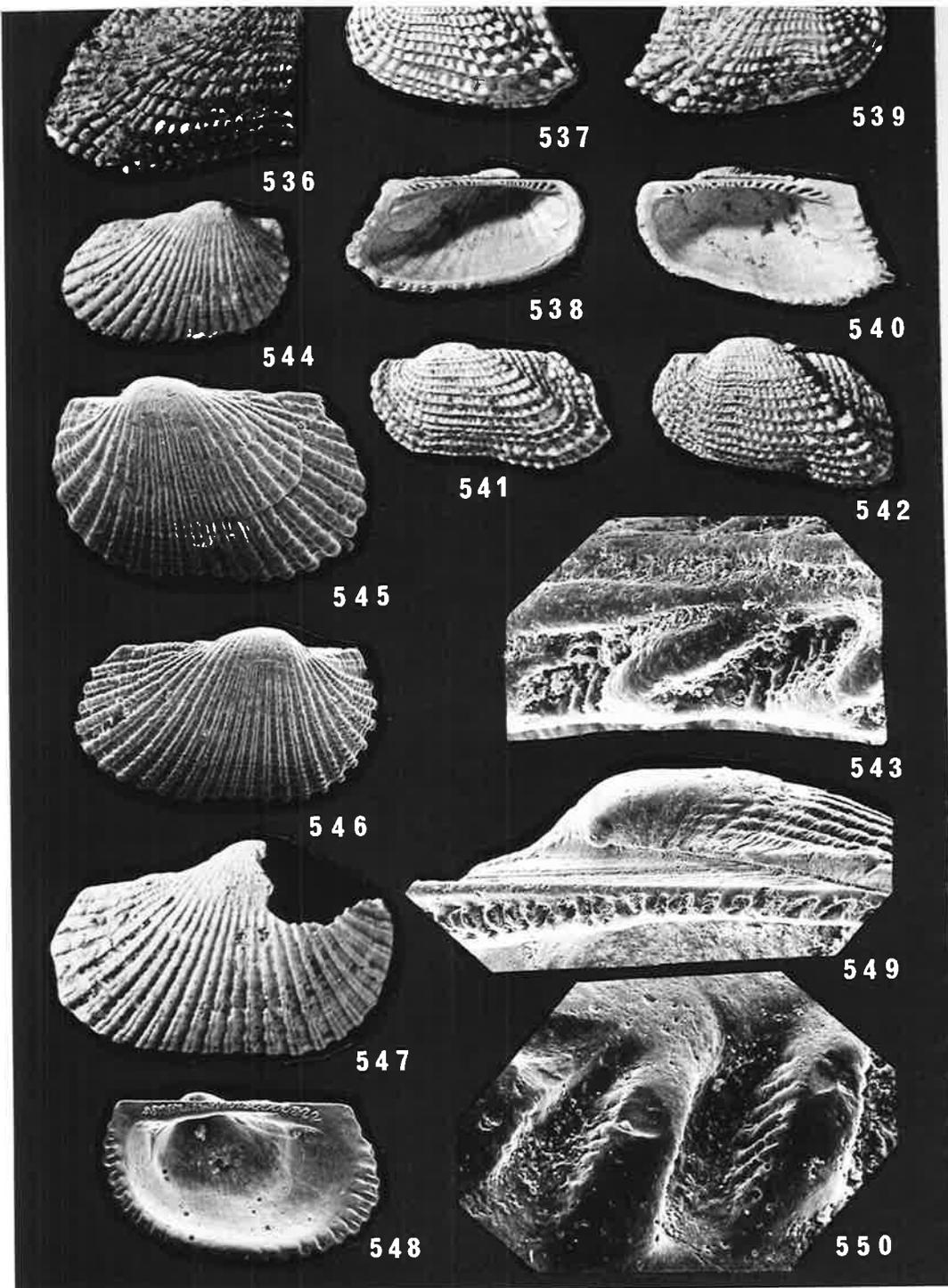
FIGS. 544-550 Grammatodon s.l. margaritatum sp. nov.: 544) Holotype,
GSSA M 3432-A, RV, outer (x 3.8). Paratypes: 545)
GSSA M 3416-C, LV, outer (x 10); 546) GSSA M 3416-D,
RV, outer (x 10); 547) GSSA M 3432-B, RV, senile,
outer (x 3.8). Paratype GSSA M 3416-B, RV; 548) inner
(x 10); 549) cardinal area (x 30); 550) posterior
hinge, particular, tooth striations (x 200).

FIGS. 551-554 Cucullaea (Cucullaea) adelaidensis Tate. 551) SAM T
1047-A, holotype, LV, outer (x 1.8); 552) SAM T 1047-D,
paratype, RV, inner (x 1.65); 553) SAM T 1047-F,
paratype, RV, outer (x 2.45); 554) SAM T 1047-E, paratype,
LV, inner (x 1.5).

FIG. 555 Arca (Barbatia) centenaria Glenn (non Say), figured
by Newell (1969) as Arca centenaria Say.

FIG. 556 Arca centenaria Say, Conrad's 1832 illustration.

FIG. 557 Striarca centenaria (Say), Bird's 1965 illustration.



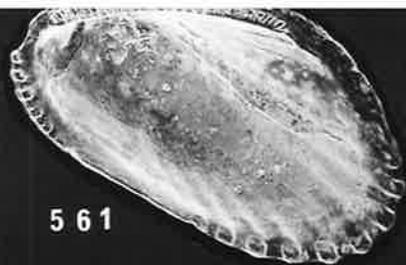
FIGS. 558-564 Scapularca scapulina (Lamarck). SAM P 21252-D:
LV, inner (x 9.2); 559) cardinal area (x 32.3);
560) particular resilifer (x 89.3). SAM P 21252-B:
561) RV, inner (x 8.2); 562) particular anterior
hinge, striations on the last tooth (x 80). SAM P
21252-C: 563) RV, outer (x 10). SAM P 21252-A:
564) LV, outer (x 10).

FIGS. 565-567 Arca lactea Linnaeus. Pheziens, France, Pliocene.
SAM P 21253: 565) LV, inner (x 9.5); 566) cardinal
area (x 14.2); 567) posterior hinge and tooth
striations (x 37).

FIGS. 568-571 Arca quadrilatera Lamarck. Paris Basin, Middle
Eocene, SAM P 21254-A: 568) LV, inner (x 10)
SAM P 21254-B: 569) RV, inner (x 10); 570)
particular, resilifer and hinge gap (x 50);
571) posterior hinge and tooth striations (x 44).



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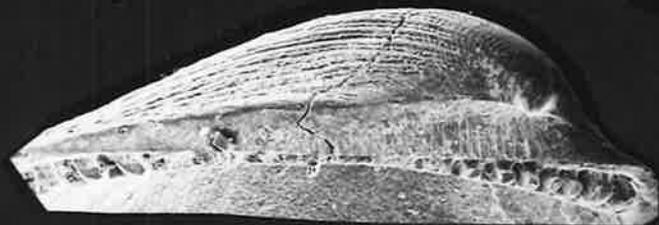
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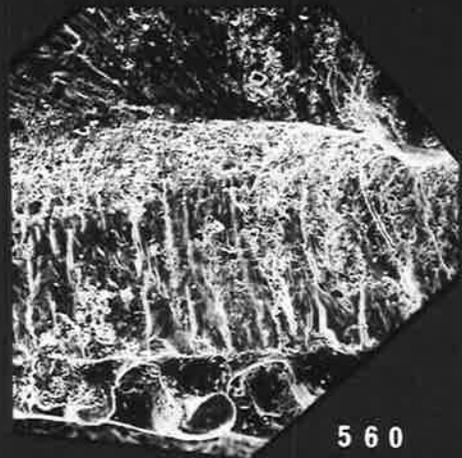
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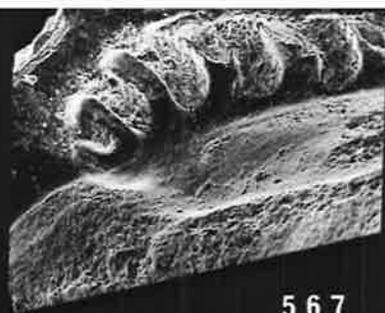
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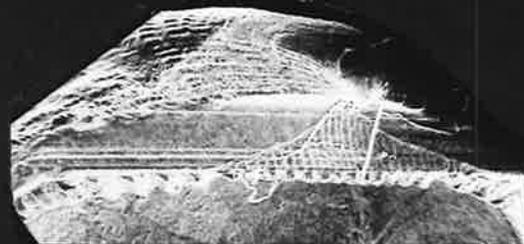
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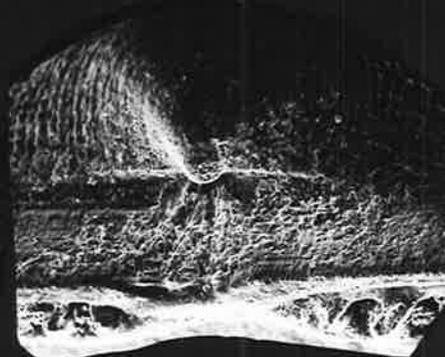
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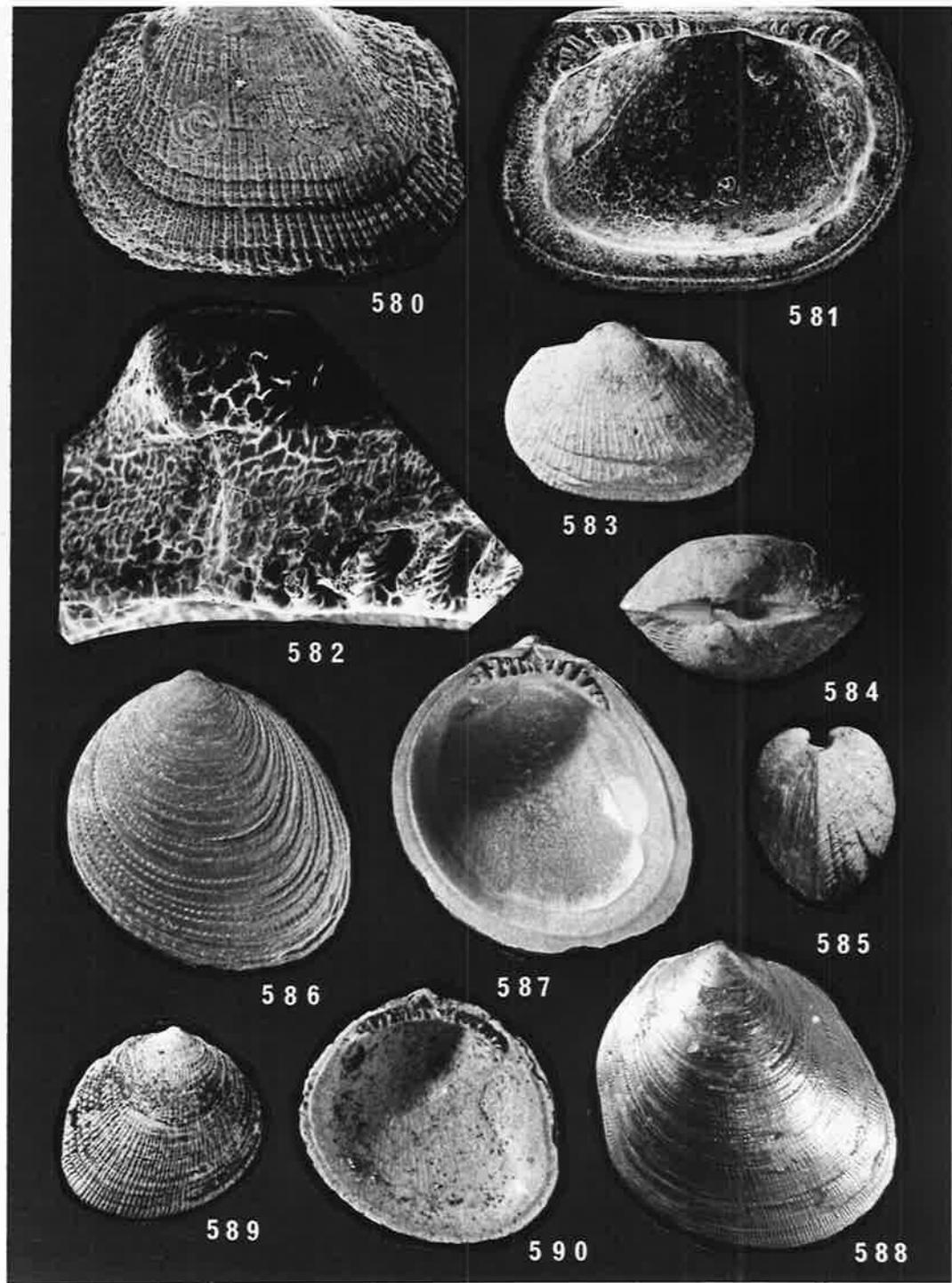
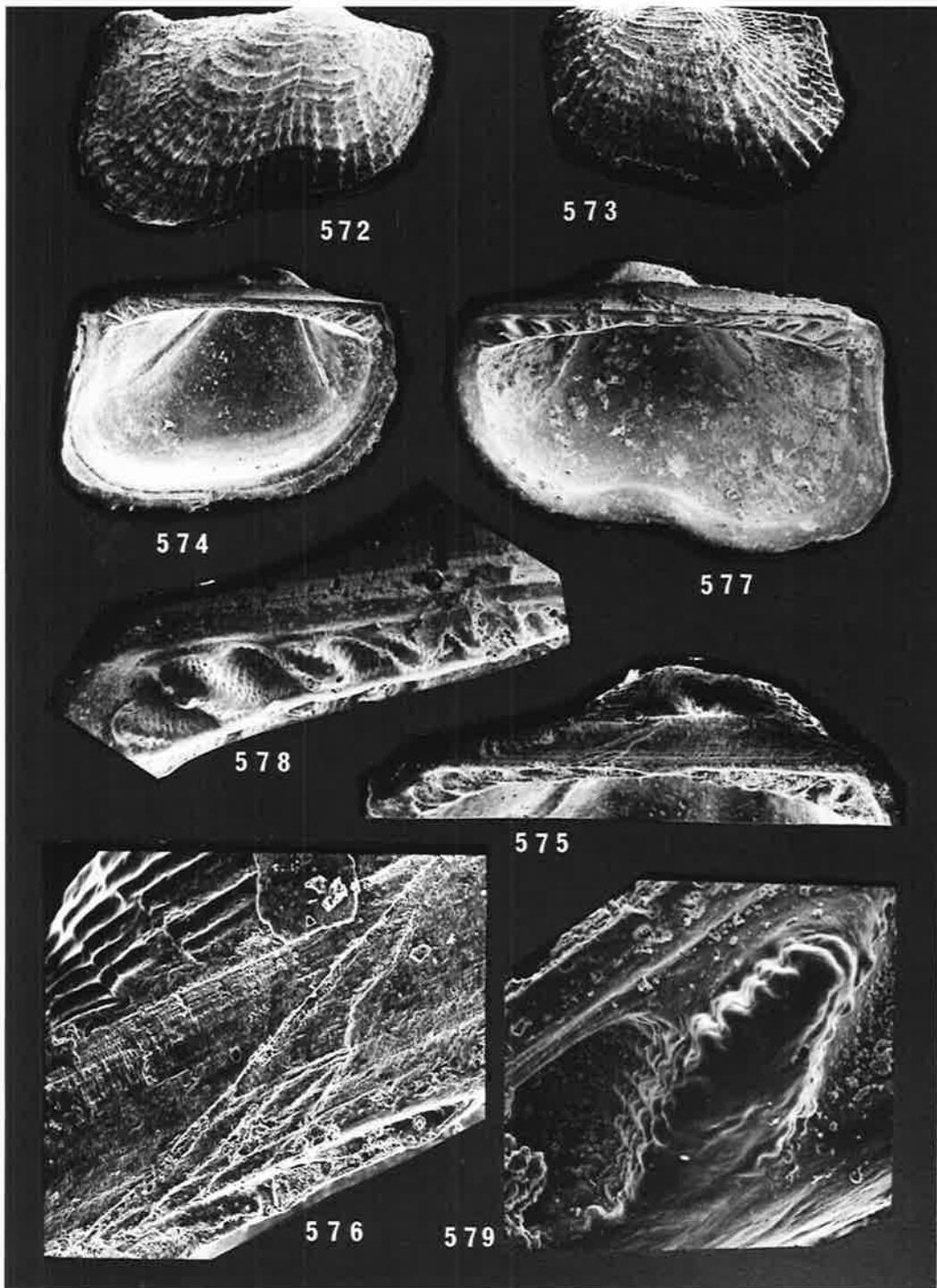


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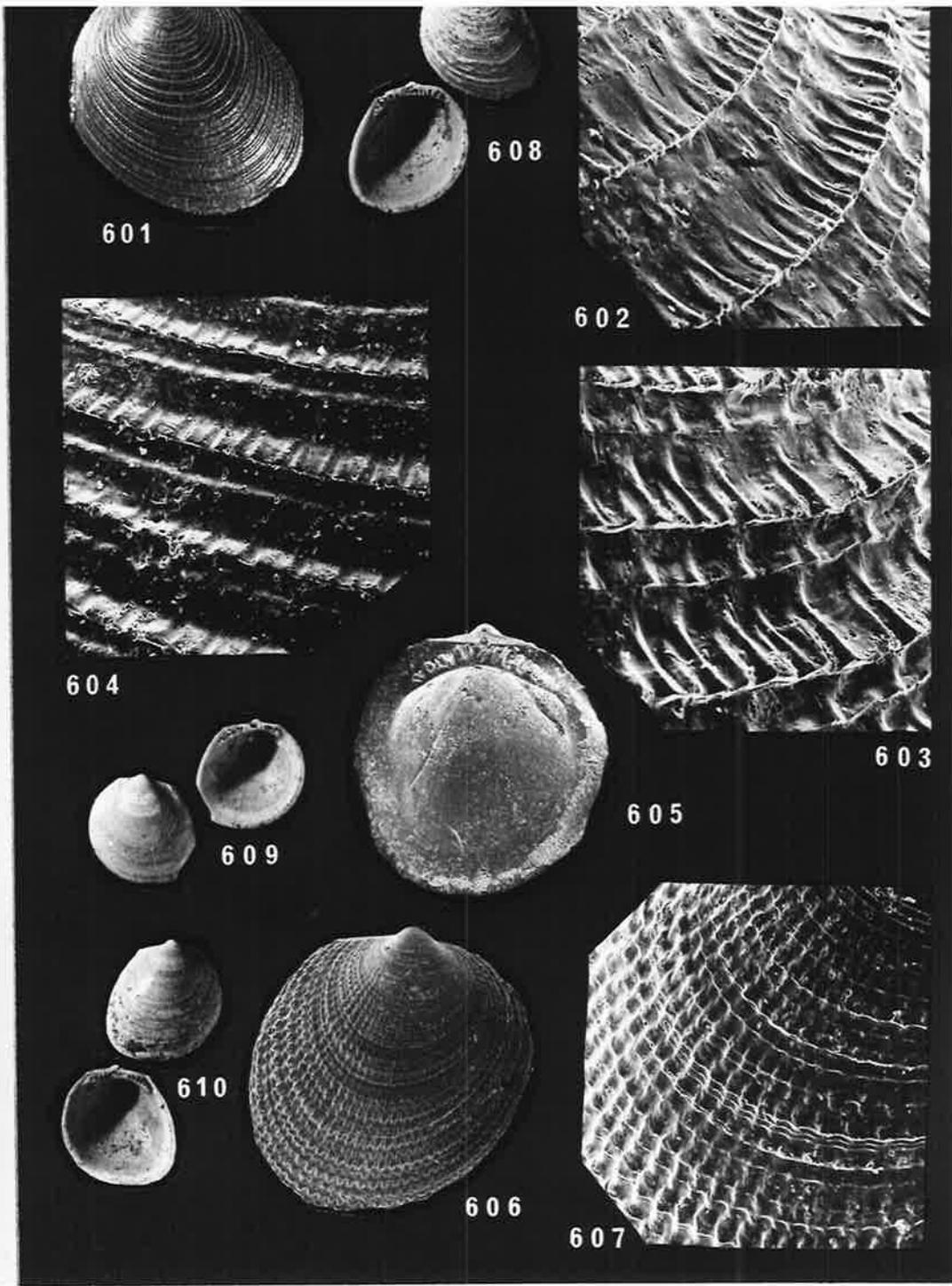
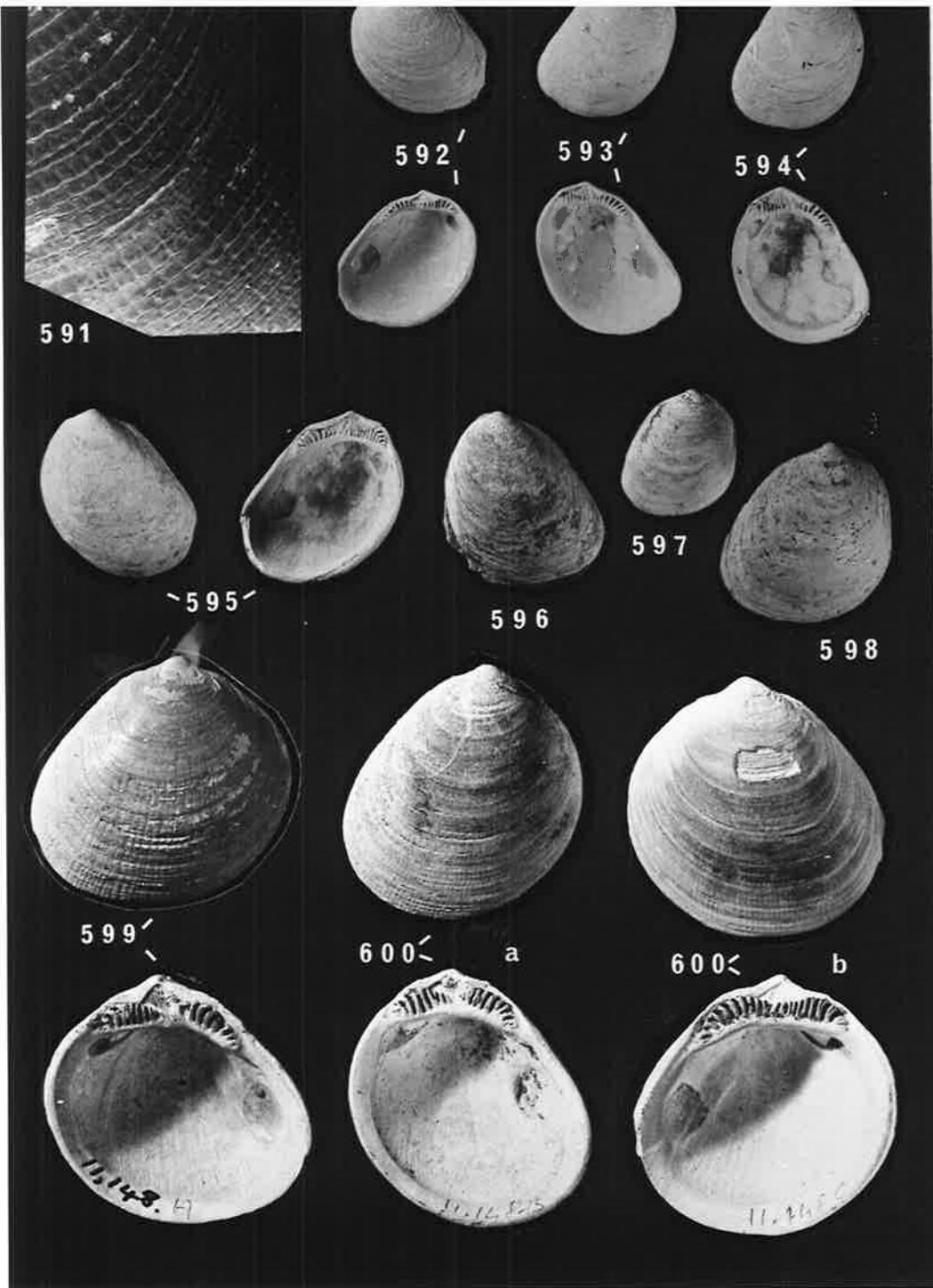


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- FIGS. 572-579 Allasinazella gen. nov. equidens (Tate). 572) SAM T 1058-M, paratype, RV, outer (x 10); 573) GSSA M 3438-B, LV, outer (x 10). GSSA M 3438-A: 574) LV, inner (x 10); 575) cardinal area (x 15); 576) particular resilifer (x 60). SAM T 1058-T, paratype: 577) RV, inner (x 10); 578) anterior hinge (x 32); 579) posterior hinge, particular tooth crenulations (x 150).
- FIGS. 580-582 Arcopsis dissimilis (Tate). 580) GSSA M 3439, LV, outer (x 9.5); 581) GSSA M 3440, LV, inner (x 10); 582) the same, particular resilifer and tooth striations (x 40).
- FIGS. 583-585 Arcopsis januarica (Marwick), NZGS 9481: 583) bivalved specimen, LV, view (x 2.6); 584) umbonal view (x 2.6); 585) anterior view (x 2.6).
- FIGS. 586-587 Limopsis (Limopsis) aurita (Brocchi), Siena, Italia, Pliocene. 586) SAM P 21251-A, LV, outer (x 3.5); 587) SAM P 21251-B, RV, inner (x 4).
- FIGS. 588 L. (Limopsis) insolita (Sowerby) BMNH 12530-1, bivalved specimen, LV View (x 2.35).
- FIGS. 589-590 L. (Limopsis) multiradiata Tate. GSSA M 2804: 289) RV, outer (x 3.3); 590) inner (x 4.2).



- FIGS. 591-598 L. (Limopsis) insolita (Sowerby). 591) BMNH 12530-1, particular ornament (x 98); 592) BMNH 12530-2, LV, outer and inner views (x 1); 593-4) BMNH 12530-7/8, RVs, outer and inner views (x 1); 595) BMNH 12530-6, LV, inner and outer views (x 1); 596-8) BMNH 12530-3/-4/-5, bivalved specimens, LV, RV, and RV views, respectively (x 1). To note the variability in outline and inner morphology.
- FIGS. 599-600 L. (Limopsis) campa Allan. Topotypes; inner and outer views (all x 1.9): 599) NZGS 11 148-A; 600a) NZGS 11 148-B; 600b) NZGS 11 148-C).
- FIGS. 601-604 L. (Limopsis) zitteli Ihering. GSSA M 2082: 601) outer, LV (x 3.6); 602) particular, microriblets in juvenile stages (x 60); 603) particular, microriblets in adult stages (x 60); 604) particular, ventral margin, riblets of 'morningtonensis' type (x 40).
- FIGS. 605-607 L. (Limopsis) multiradiata Tate. juveniles: 605) GSSA M 3429, LV, inner (x 10); 606) GSSA M 3430), RV, outer (x 10); 607) the same, particular ornament (x 30).
- FIGS. 608-610 L. (Limopsis) zitteli Ihering. Juveniles. Topotypes of 'Limopsis waihaoensis Allan', NZGS 9508-A/-C, outer and inner views; LV, LV, and RV, respectively (all x 2.45).



- FIG. 611 L. (Limopsis) zitteli Ihering. GSSA M 2816, outer shells.
Lot showing the variability range in outline (all x 1).
- FIG. 612-614 L. (Limopsis) zealandica Hutton, Topotypes, NZGS
9520-A/-C, outer shells; RV, RV, and LV respectively
(all x 1.9)

- FIG. 615 L. (Limopsis) zitteli Ihering. The same as in Fig. 611,
inner views showing the variability of the cardinal
area and of the hinge (all x 1).
- FIGS. 616-618 L. (Limopsis) zealandica Hutton. The same as in Figs.
612-4, inner views (all x 1.9).

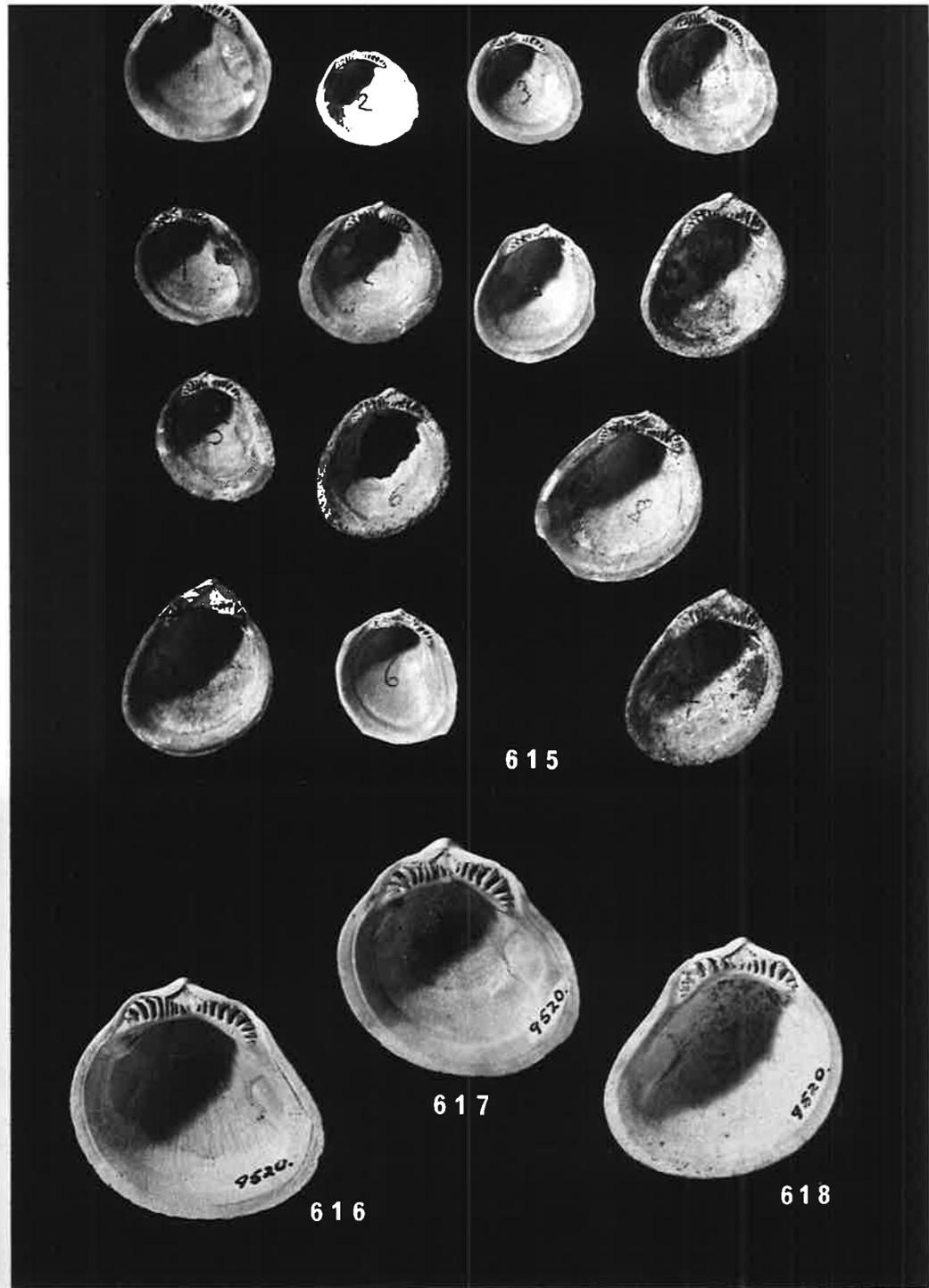
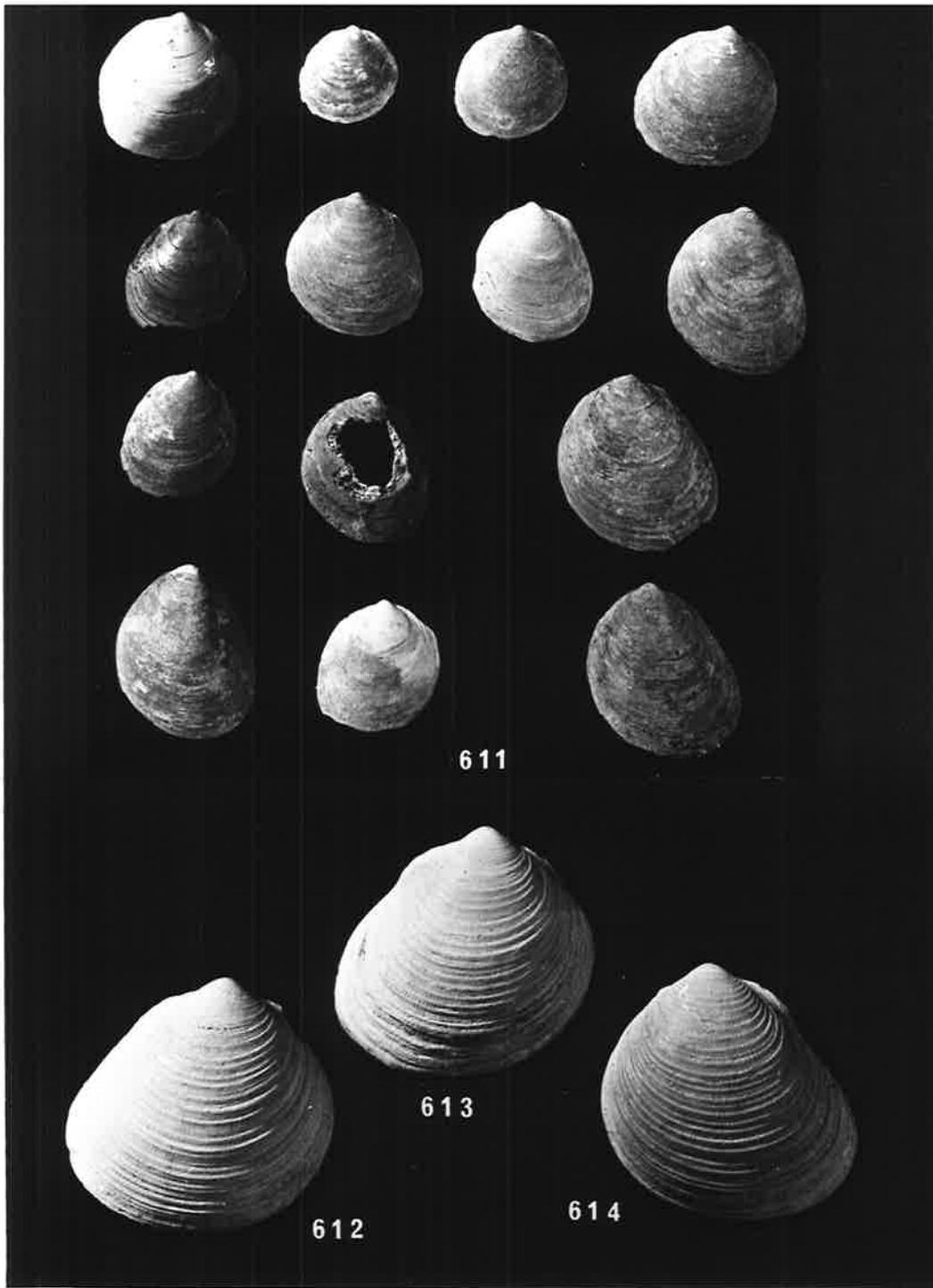
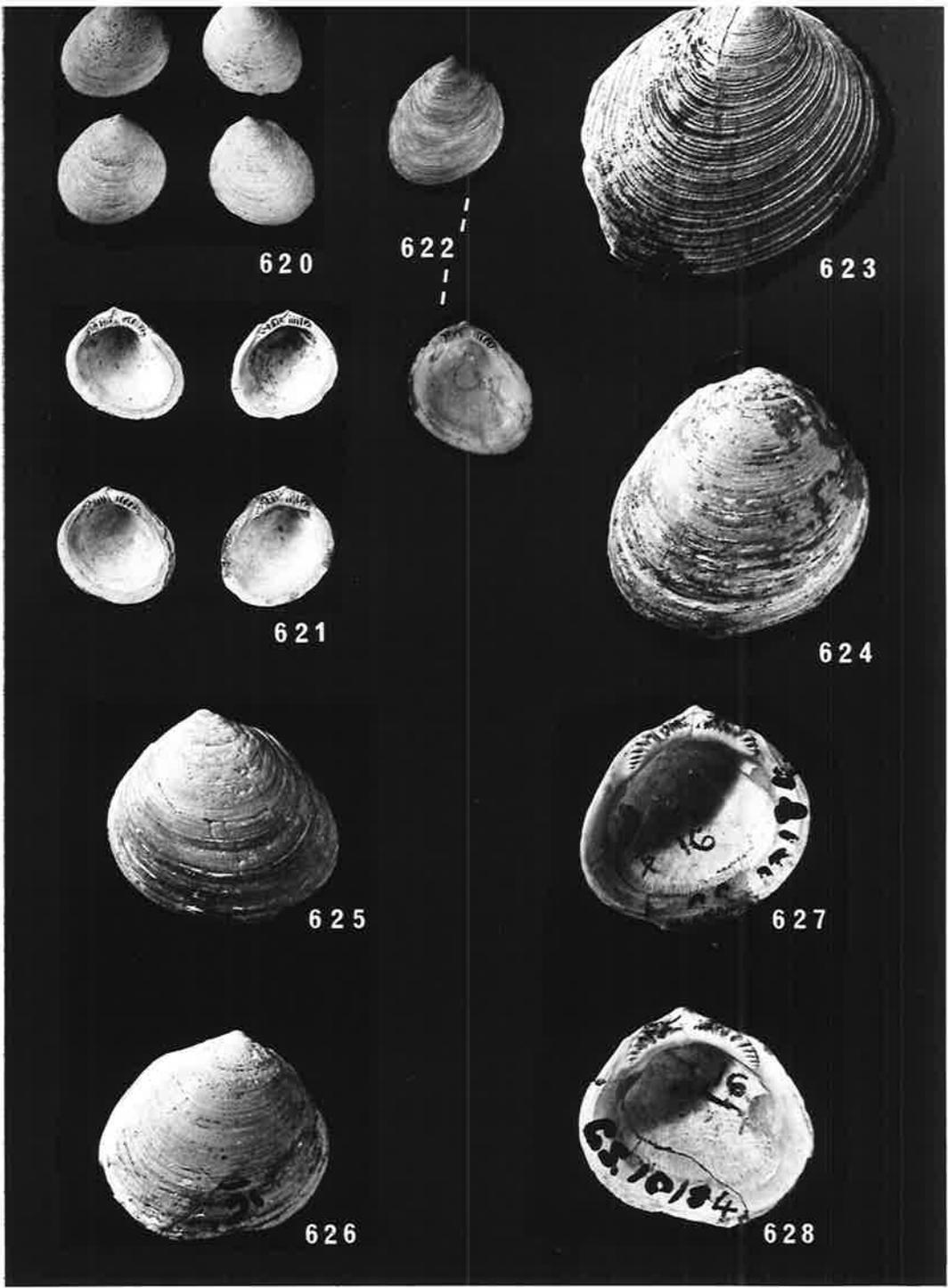
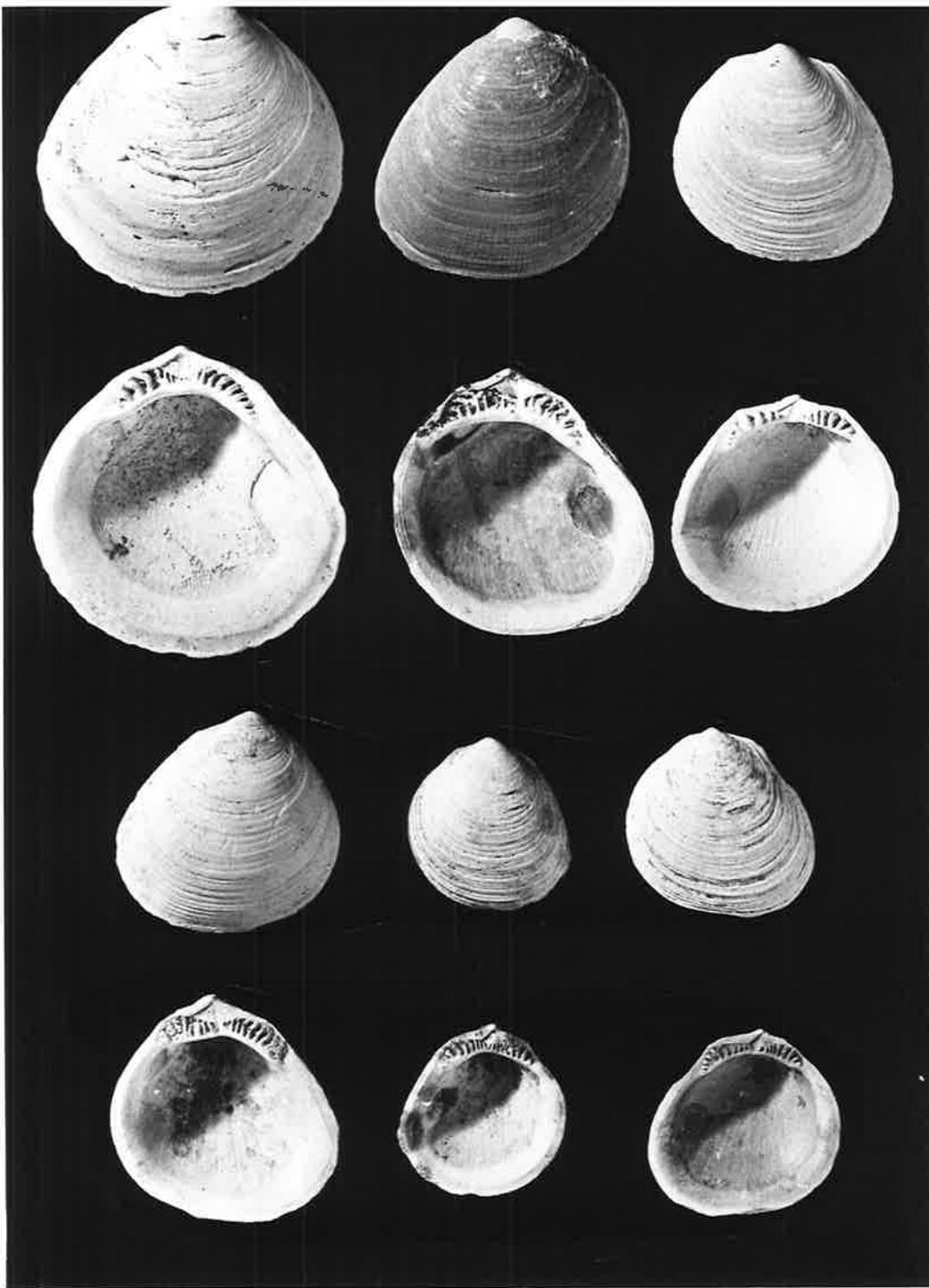


FIG. 619 L. (Limopsis) zitteli Ihering. GSSA M 3436, lot from Sutherlands, South Canterbury, New Zealand (Awamoan, Early Miocene): outer and inner views (all x 3).

FIGS. 620-622 L. (Limopsis) valida Singleton. NMV P 31297: 620) outer views (x 1); 621) inner views (x 1); 622) SAM P 21252, cast of the holotype, RV, inner and outer views (x .9).

FIG. 623 L. (Limopsis) morningtonensis Pritchard. SAM T 1020-A, RV, outer (x 2.8).

FIGS. 624-628 L. (Limopsis) lawsi King. Topotypes: 624) NZGS 10 184-A, bivalved specimen, RV view (x 1.8); 625) NZGS 10 184-B, LV, outer (x 1.75); 626) the same, inner (x 1.75); 627) NZGS 10 184, RV, outer (x 1.75); 628) the same, inner (x 1.75).

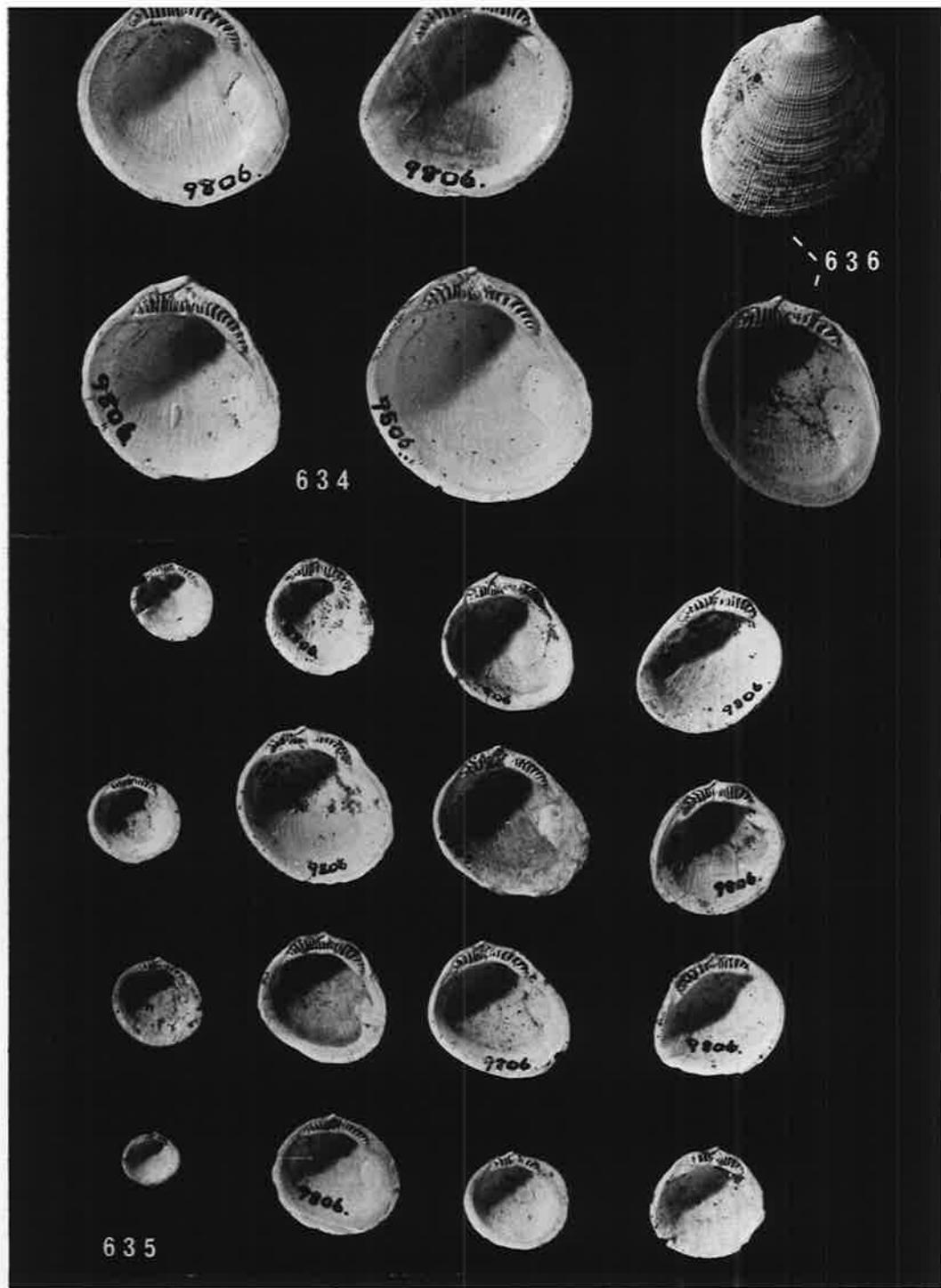
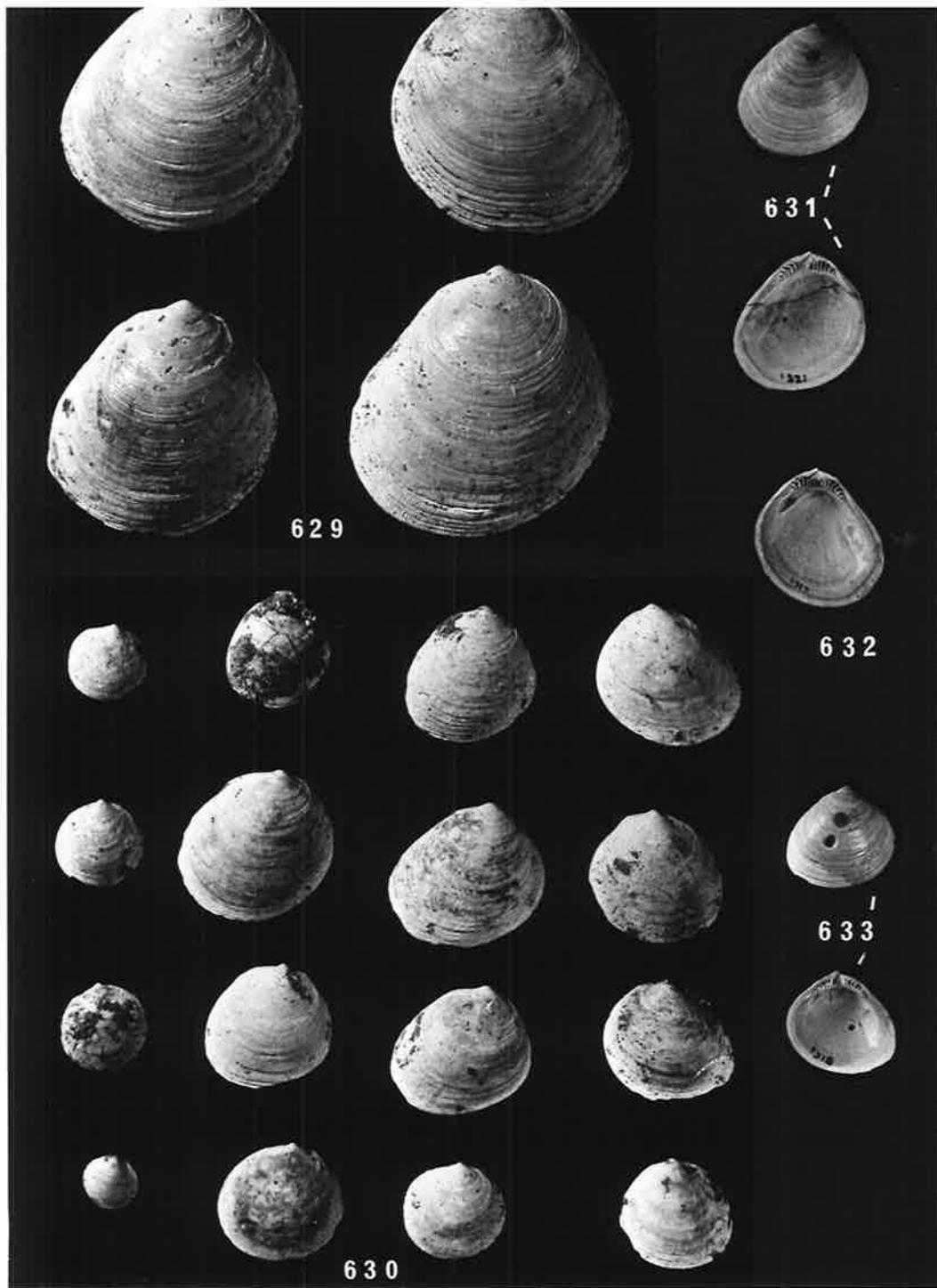


FIGS. 629-630 L. (Limopsis) zitteli Ihering. Topotypes of 'L. parma Marwick', Shell Gully, Chatton, New Zealand (Duntroonian, Middle Oligocene), ZNGS 9806: 629) seniles, outer (x 1.72); 630) lot showing the variability range in outline, outer views (all x 1).

FIGS. 631-633 L. (Limopsis) zitteli Ihering. Types of 'L. chapmani Singleton': 631) holotype, MUGD 1317, RV, outer and inner views (x .95); 632) paratype, MUGD 1321, LV, inner (x .95); 633) paratype, MUGD 1318, RV, inner and outer views (x .95).

FIGS. 634-635 L. (Limopsis) zitteli Ihering. The same as in Fig. 628, inner view (x 1.46); 635) the same as in Fig. 629, inner views showing the variability in cardinal area and in hinge (all x 1).

FIG. 636 L. (Limopsis) maccoyi Chapman. GSSA 3431, topotype, RV: 636) outer view and inner view (x 1.9).



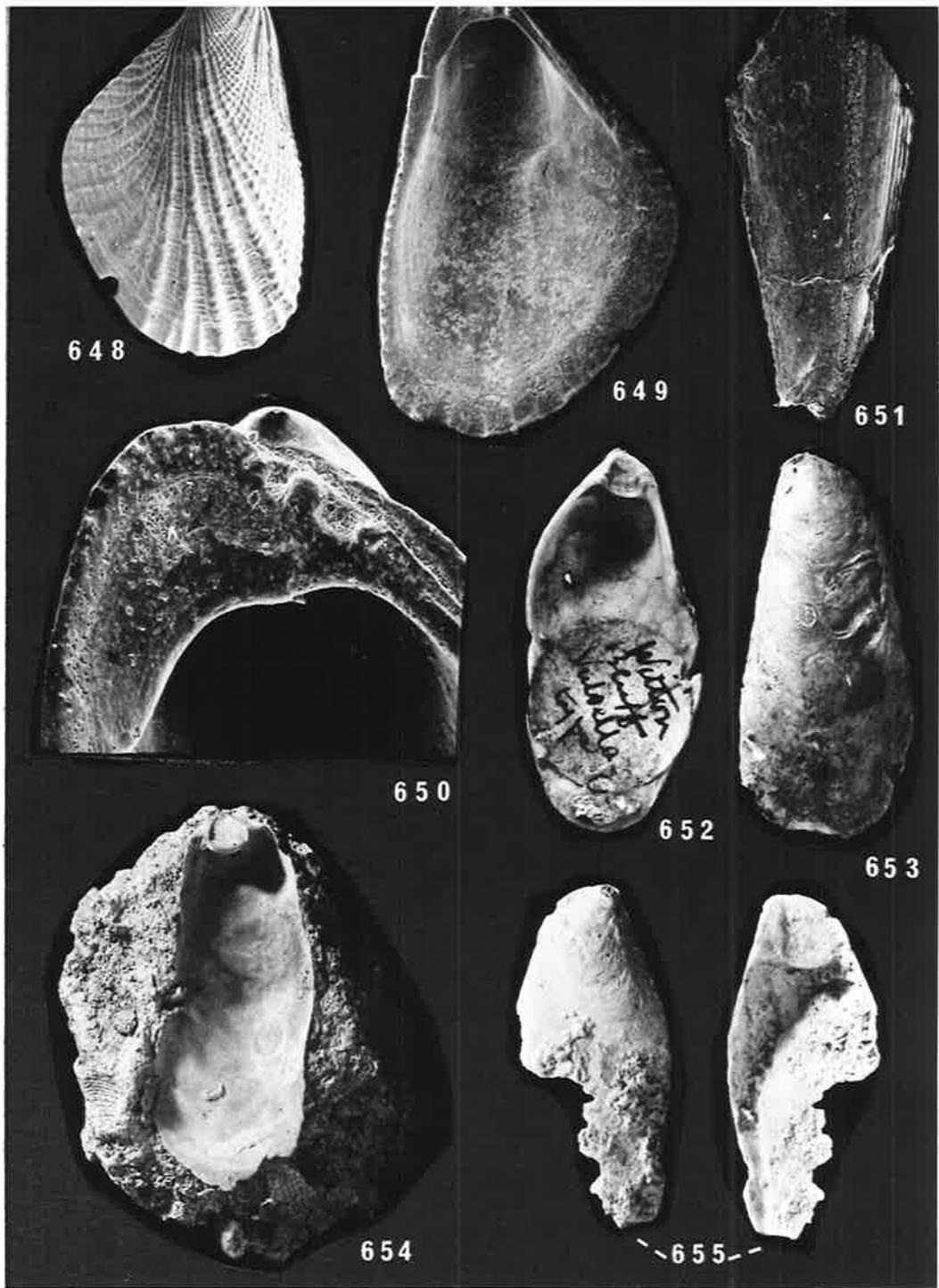
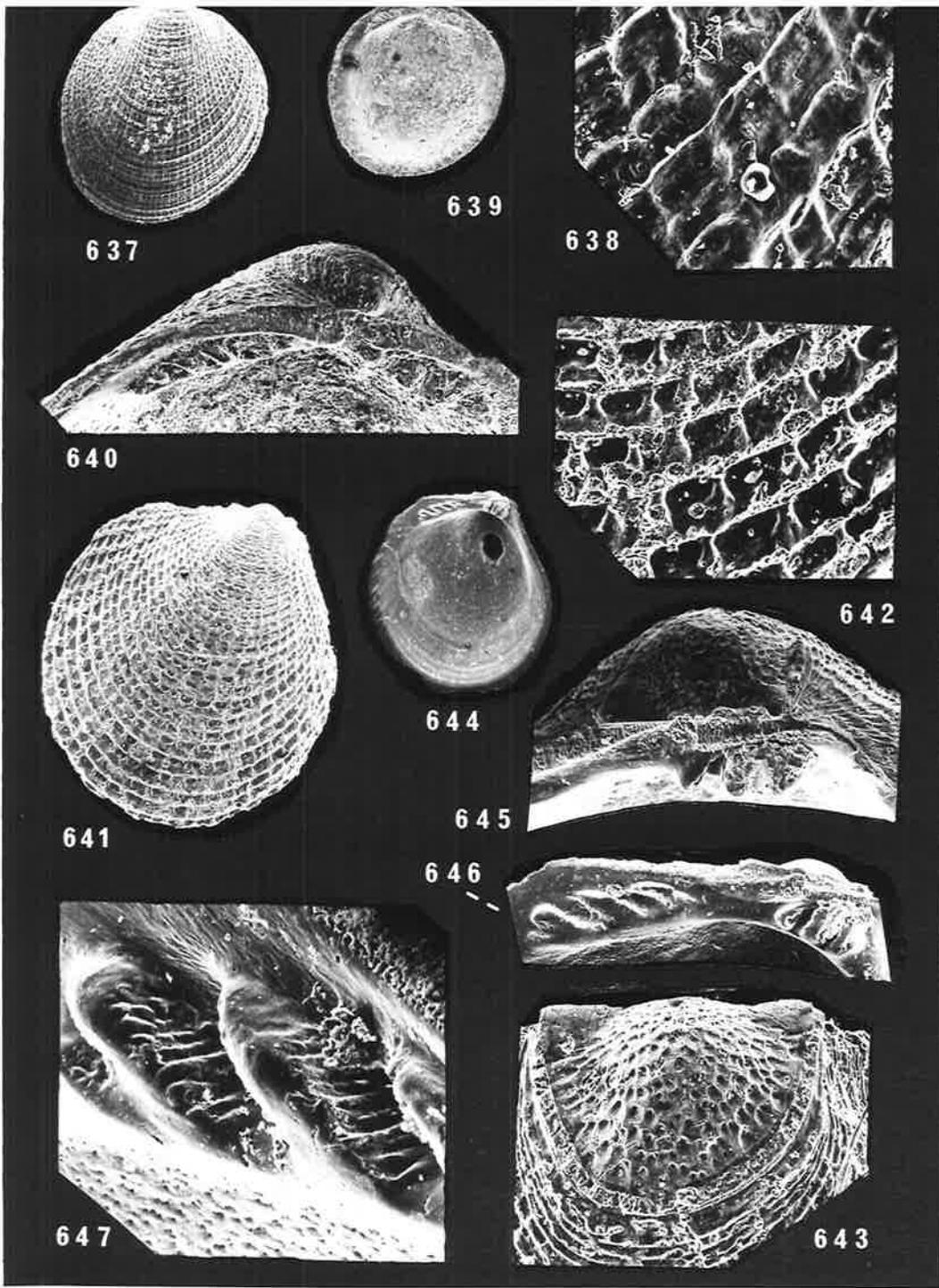
FIGS. 637-640 Limopsis (Pectunculina) cancellata sp. nov. SAM P 21205-A
holotype, RV: 637) outer view (x 10); 638) particular,
ornament (x 80). Paratype, SAM P 21205-B, LV: 639) inner view
(x 10); 640) cardinal area and hinge (x 30).

FIGS. 641-647 Limarca angustifrons Tate. GSSA M 2871-A, RV: 641)
outer (x 20); 642) particular, ornament (x 70); 643)
prodissoconch (x 70). GSSA M 2871-B, LV: 644) inner
view (x 10); 645) cardinal area, resilifer, and prodis-
soconch (x 50); 646) hinge (x 30); 647) particular,
striations, posterior teeth (x 130).

FIGS. 648-650 Septifer (Septifer) sp. nov. 648) GSSA M 3441-A,
LV, outer (x 10); 649) GSSA M 3441-B, RV, inner
(x 9.5); 650) the same, particular hinge (x 40).

FIG. 651 Pinna sp., SAM P 21255 (x 9.5).

FIGS. 652-655 Vulsella laevigata Tate: 652) SAM T 975-A, holotype,
LV, inner (x 1.9); 653) SAM T 975-B, paratype, LV,
outer (x 1.9); 654) SAM P 21237-A, RV, inner (x 1.7);
655) SAM P 21237-B, RV, outer (x 2.7) and inner
(x 2.8), respectively.



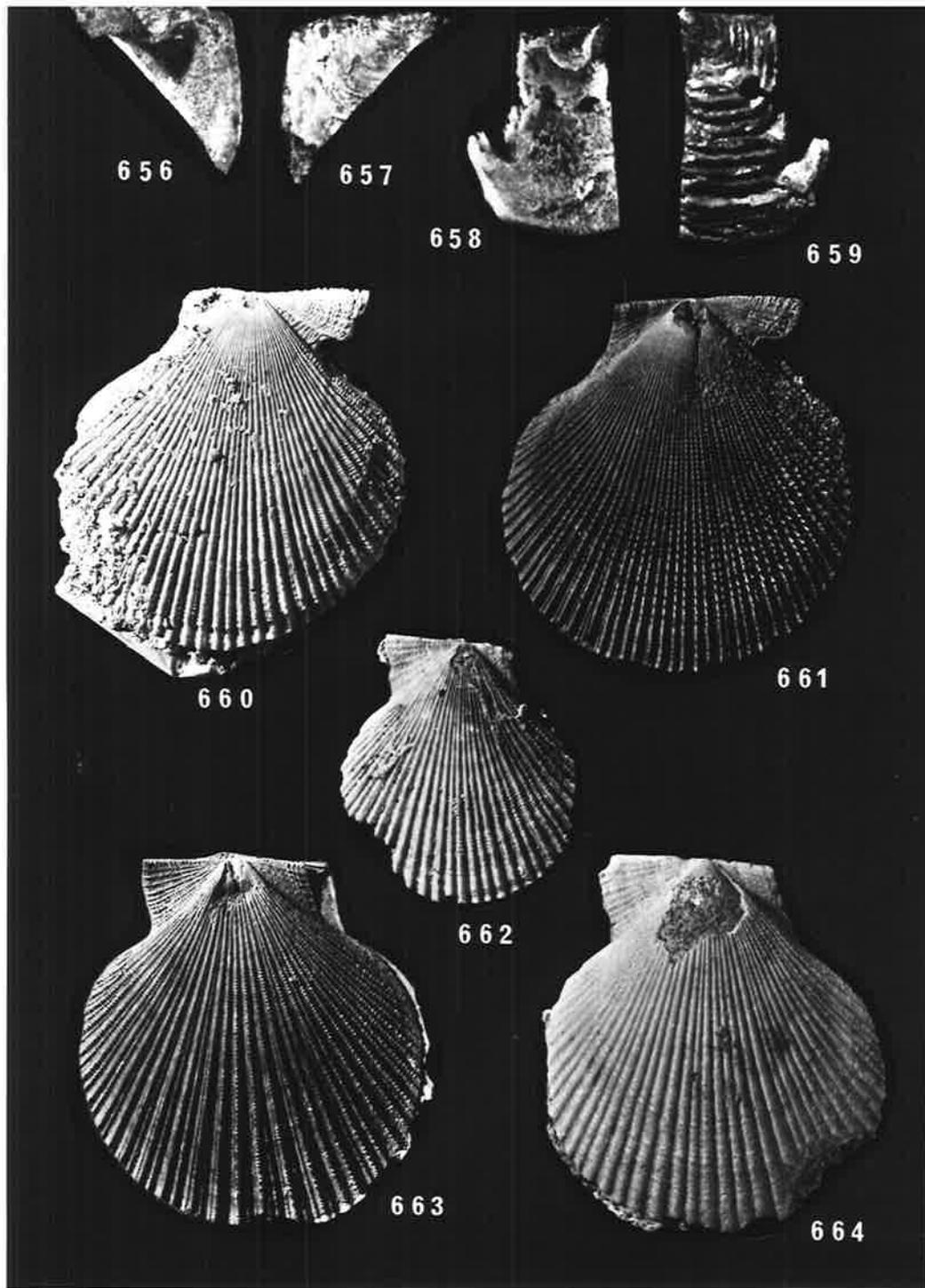
FIGS. 656-659 Pinctada (Pinctada) sp. nov. 656-57) SAM P 21256-A,
fragment of RV: inner (x 4), and outer (x 4);
658-59) SAM P 21256-B, fragment of LV: inner (x 4)
and outer (x 4).

FIGS. 660-663 Chlamys (Chlamys) peroni (Tate). 660) SAM P 21241,
RV, outer (x 1.9); 661) SAM T 930-C, holotype, RV,
outer (x 1.5); 662) SAM P 21240, LV, outer (x 1.9);
663) SAM T 930-E, paratype, bivalved specimen, LV
outer view (x 1.3).

FIG. 664 C. (Chlamys) flindersi (Tate), SAM P 21239-A, LV,
outer (x 1.3).

FIGS. 665-667 C. (Chlamys) flindersi (Tate). 665) SAM T 931-B,
paratype, RV, outer, (x 1.3); 666) SAM T 931-D,
paratype, LV, outer (x 1.55); 667) SAM P 21239-B,
RV, outer (x 1.3).

FIGS. 668-670 C. (Chlamys) aldingensis (Tate). 668) SAM P 21242-A,
RV, outer (x 1.65); 669) SAM P 21242-B, LV, outer
(x 1.65); 670) SAM P 21242-C, LV, outer (x 1.7).



FIGS. 671-675 Spondylus (Spondylus) tortachillensis sp. nov. 671) SAM T 987-A, holotype, LV view (x 1.4); 672) the same, RV view (x 1.4); 673) SAM P 21238, 1 paratype, LV, outer (x 1.6).

FIGS. 674-681 Dimya sigillata Tate. 674) SAM P 21233-A, RV, outer (x 4.4); 675) SAM P 21233-B, LV, inner (x 4.4); 676) SAM P 21234-C, RV, inner (x 10.9); 677) SAM P 21233-E, RV, inner, attached to a Bryozoa (x8.16); 678) the same, particular hinge (x 20); 679) SAM P 21234-B, LV, inner, (x 8.5); 680) SAM P 21233-D, LV, outer, laterally tilted (x 6.15); 681) SAM P 21234-A, RV, outer, attachment area broken (x 7.47).



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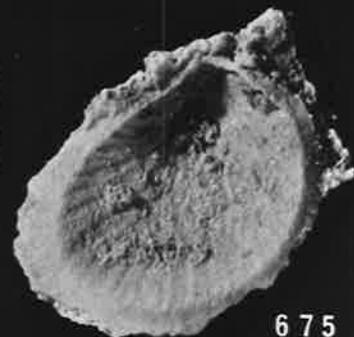


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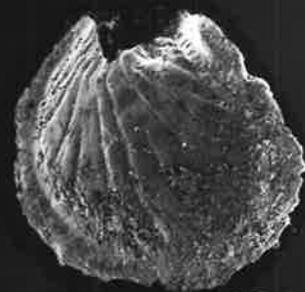
*Aldinga
truncata
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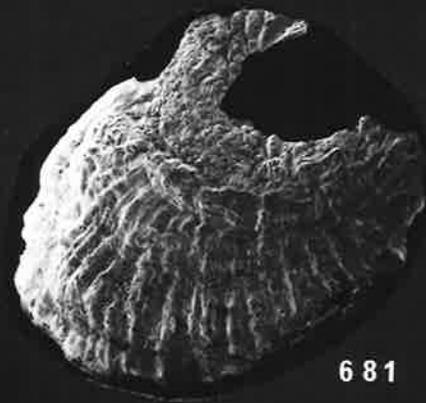
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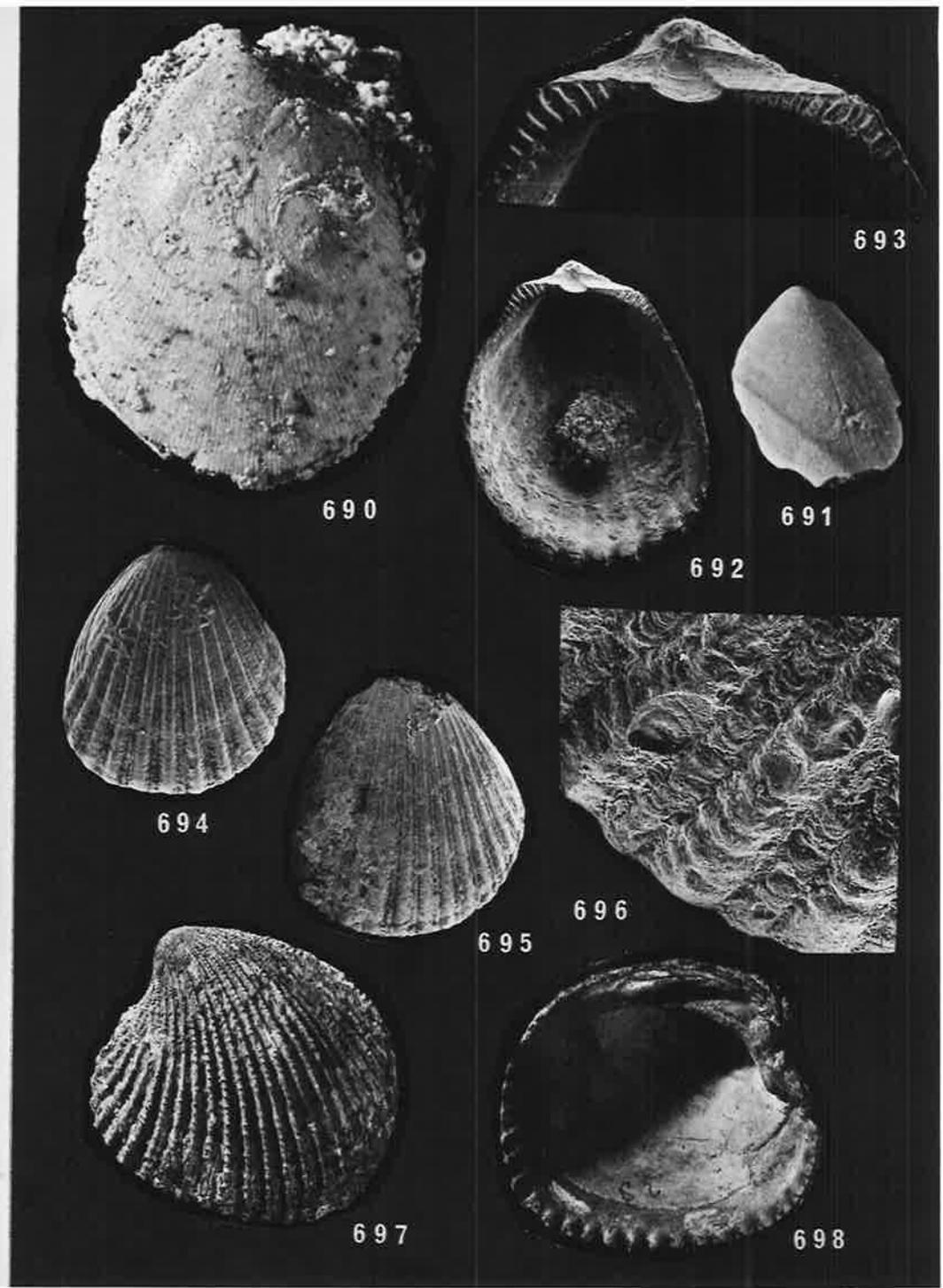
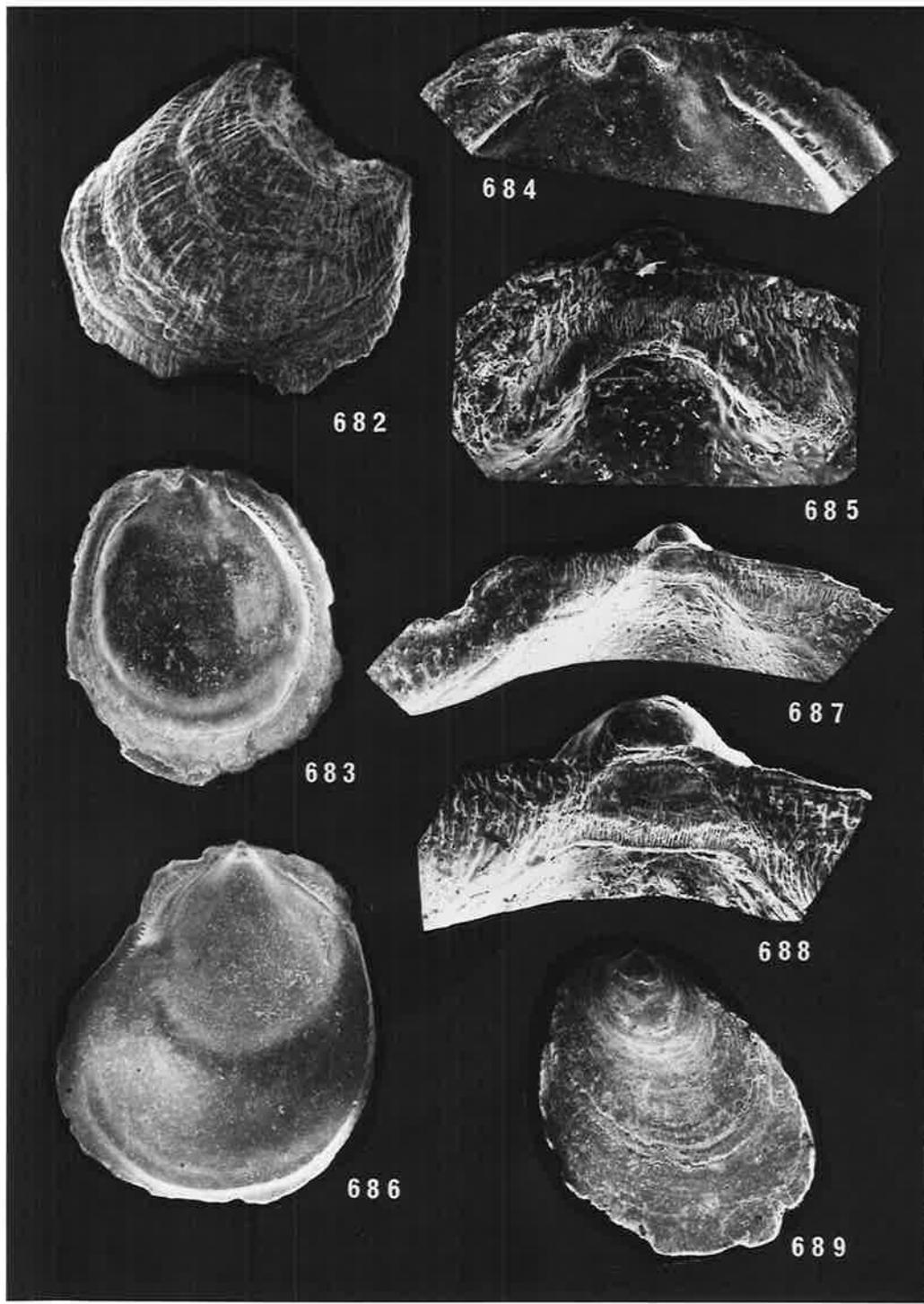
FIGS. 682-689 Dimya asseretoi sp. nov. SAM P 21206-A, holotype, RV: 682) outer (x8.75); SAM P 21206-B, paratype, RV; 683) inner (x 10); 684) hinge (x 20); 685) particular tooth and tooth striations (x 70). SAM P 21206-C, paratype, LV; 686) inner (x8.78); 687) hinge (x 30); 688) particular fossette and ligamental striations (x 70). SAM P 21206-D, LV; 689) outer (x 8.6)

FIG. 690 Ctenoides sp. nov. aff. linguliformis (Tate), SAM P 21244, ?RV, outer (x 2.8).

FIG. 691 Divarilima cf. polyactina (Tate) SAM P 21249, RV, outer (x 3.9).

FIGS. 692-696 Limea (Isolimea) alticosta (Tate). 692) SAM P 21257, RV, inner (x 9); 693) the same, hinge (x 21.85); 694) SAM P 21236-A, RV, outer (x 10); 695) P 21236-B, RV, outer (x 10); 696) SAM P 21236-C, LV, particular abraded ornament (x 40).

FIGS. 697-698 Paraglans latissima (Tate). SAM T 1130-B, paratype, LV: 697) outer (x 1.5); 698) inner (x 1.7)



FIGS. 699-701 Corbula (Caryocorbula) pyxidata Tate. 699) SAM P 21235, RV, inner (x 8.92); 700-701) SAM P 21250, LV, inner and outer (x 3.8).

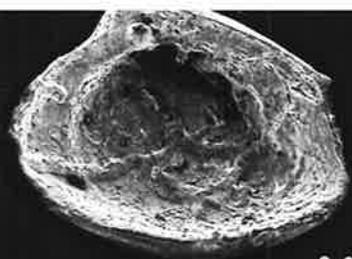
FIGS. 702-704 Hiatella (Hiatella) ?vera (Deshayes) . 702-03) SAM T 853-B, outer and inner LV (x 3.9); 704) SAM P 21248, mould, RV (x 3.9).

FIGS. 705-708 Jouannetia (Pholadopsis) cuneata Tate. SAM T 350-A, holotype: 705) RV view (x 1.8); 706) LV view (x 1.8); 707) anterior view (x 2.7). SAM P 21243: 708) LV view (x 1.75).

FIG. 709 Clavagella (Clavagella) ?lirata (Tate), SAM T 322-D (x 1.9).

FIGS. 710-712 C. (Clavagella) lirata Tate. 710) SAM T 323, holotype (x 2.9); 711-12) SAM T 322-C,-E, moulds (x 1.7 and x 1.85, respectively).

FIG. 713 Pycnodonte (Pycnodonte) sp. cf. tatei (Suter). SAM P 21245, LV, inner (x 1.3).



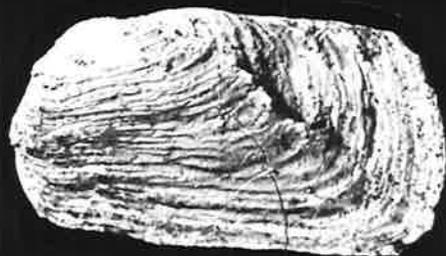
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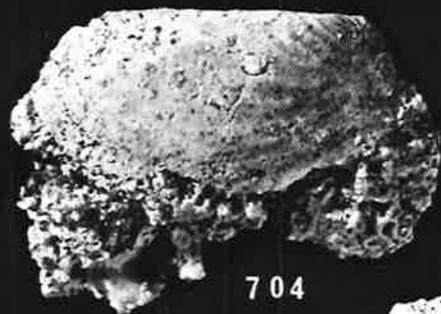
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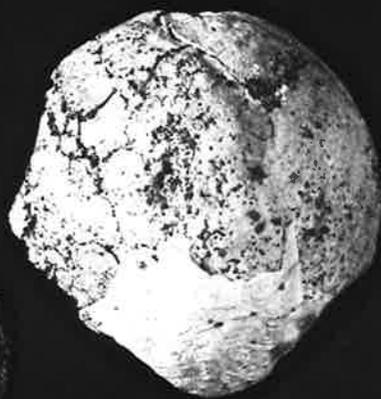
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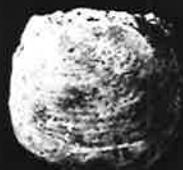
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FIGS. 714-716 Pycnodonte tatei (Suter). 714) SAM P 21247-B,
RV, inner (x 1.7); 715) SAM P 21247-C, RV,
outer (x 1.75); 716) SAM P 21247-A, bivalved,
LV view (x 1.3).

FIGS. 717-718 P. (Pycnodonte) tatei (Suter), SAM P 21247-D,
LV, outer and inner (x 1.7).

FIGS. 719-720 Notostrea subdentata (Hutton). SAM P 21258,
topotype, Trelissick Basin, New Zealand
(Duntroonian, Middle Oligocene). LV, outer
(x 1.76) and inner (x 1.76). Posterior vermicula-
tions obscured by matrix.



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FIGS. 721-723 Pycnodonte (Phygraea) andreaei sp. nov. SAM P 21259, holotype, LV: 721) inner (x 1.9); 722) outer (x 1.85); 723) lateral posterior view (x 2.15).

FIGS. 724-731 Pycnodonte (Phygraea) tarda (Hutton). 724) SAM T 916-G, RV, outer, (x 1.7); 725) SAM T 916-D, LV, outer (x 1.65); 726) SAM T 912-E, LV, outer (x .96); 727) SAM T 912-C, LV, outer (x 1.15); 728) SAM T 912-B, LV, inner (x 1.4); 729) SAM T 916-A, LV, outer (x 1.0); 730) SAM T 912-D, LV, outer (x 1.65); 731) SAM T 916-E, RV, outer (x 1.4).

FIGS. 732-739 P. (Phygraea) tarda (Hutton). Topotypes, Tioriori, Chathams Islands (Waipawan, Middle Paleocene). NZGS 1177-D, RV: 732) inner (x 2.42); 733) hinge (x 3.13); 734) outer (x 2.54). NZGS 1177-A, LV: 735) inner (x 1.35); 736) outer (x 1.31). NZGS 1177-C, LV: 737) inner (x 1.35); 738) outer (x 1.35). NZGS 1177-B, LV: 739) outer (x 1.08).

FIGS. 740-742 P. (Phygraea) tarda (Hutton). 740) SAM T 916-C, LV, outer (x 1.41); 741) SAM T 912-A, LV, outer (x .9); 742) SAM T 916-B, LV, outer (x 1.45).

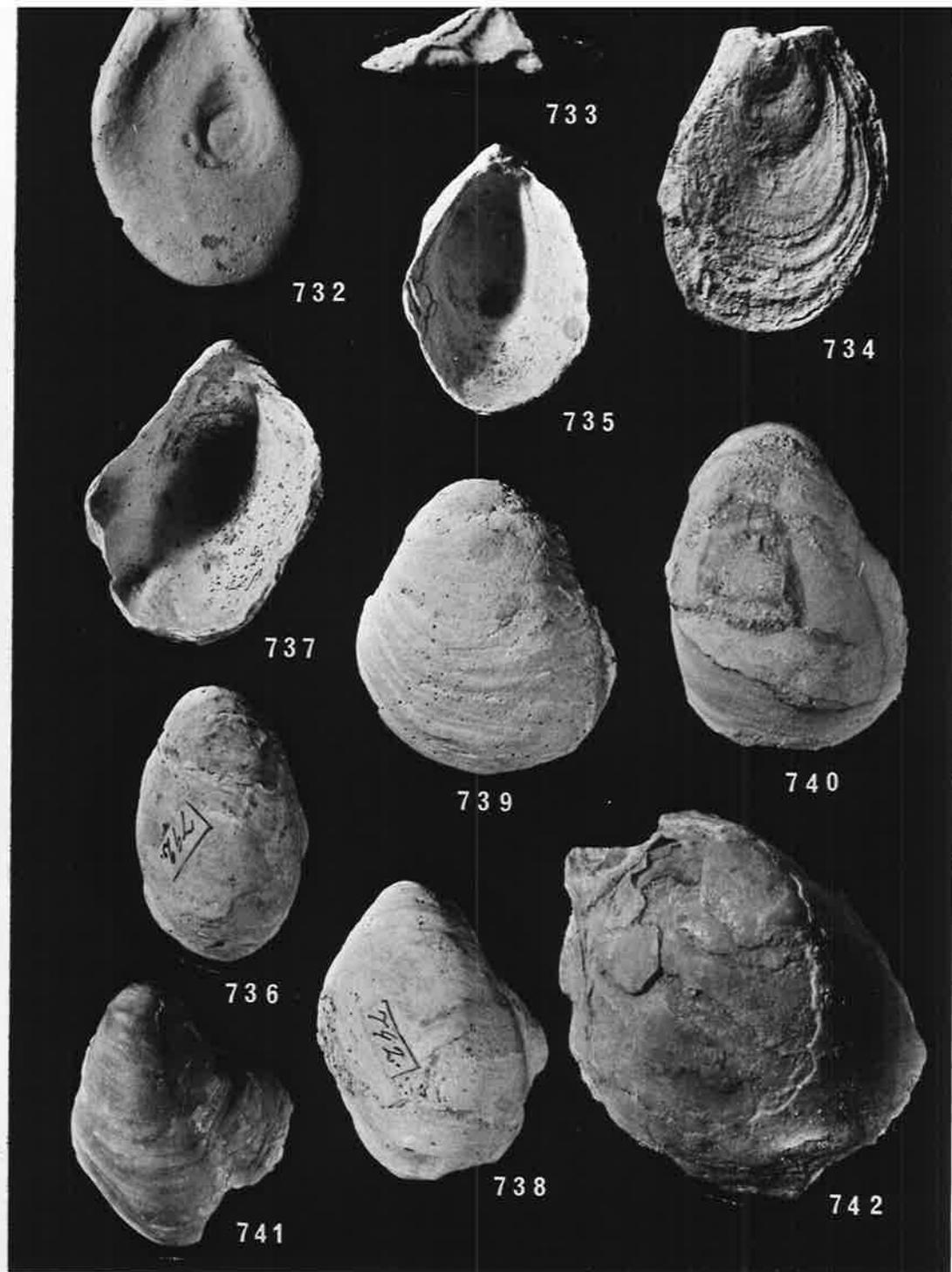
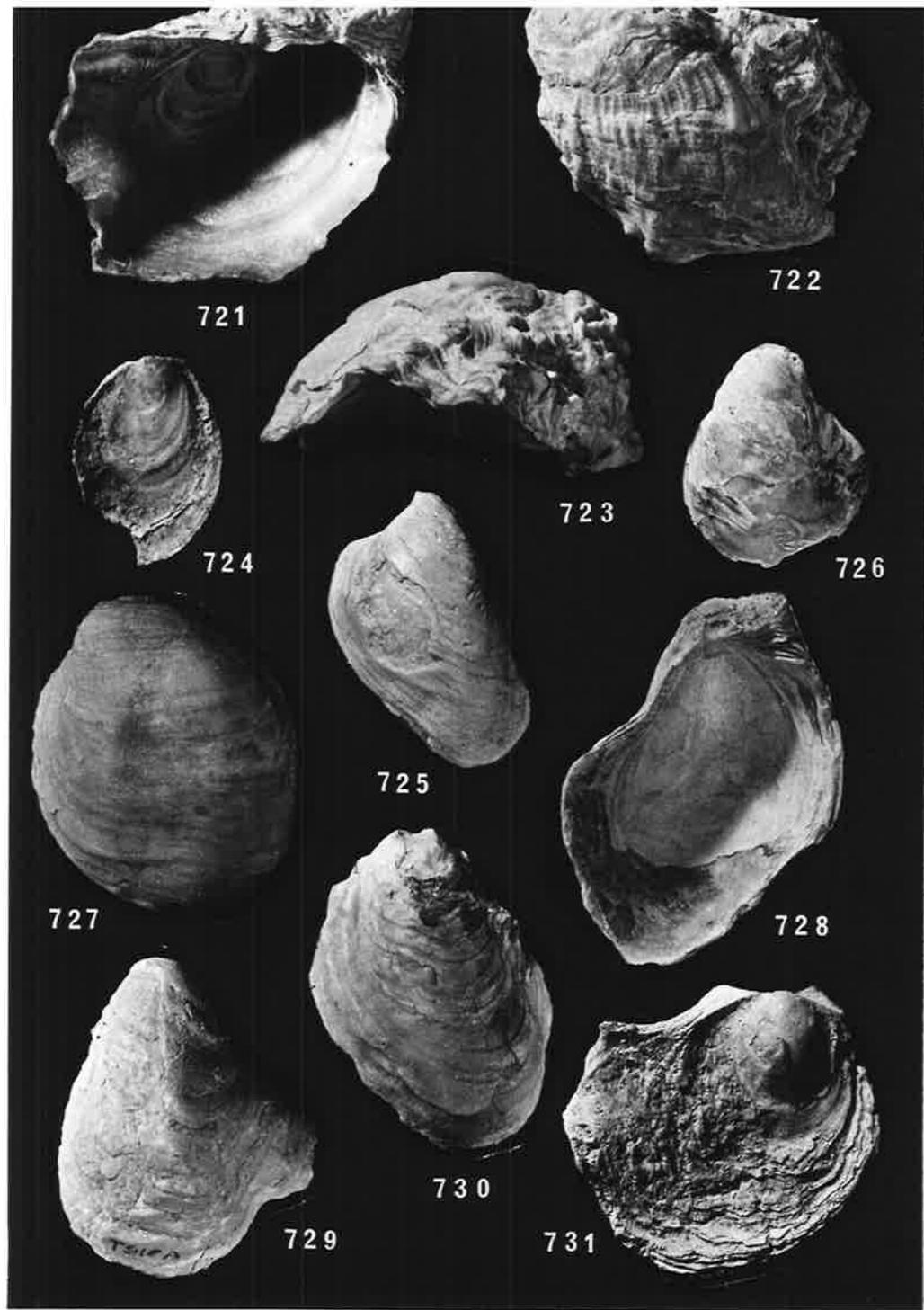
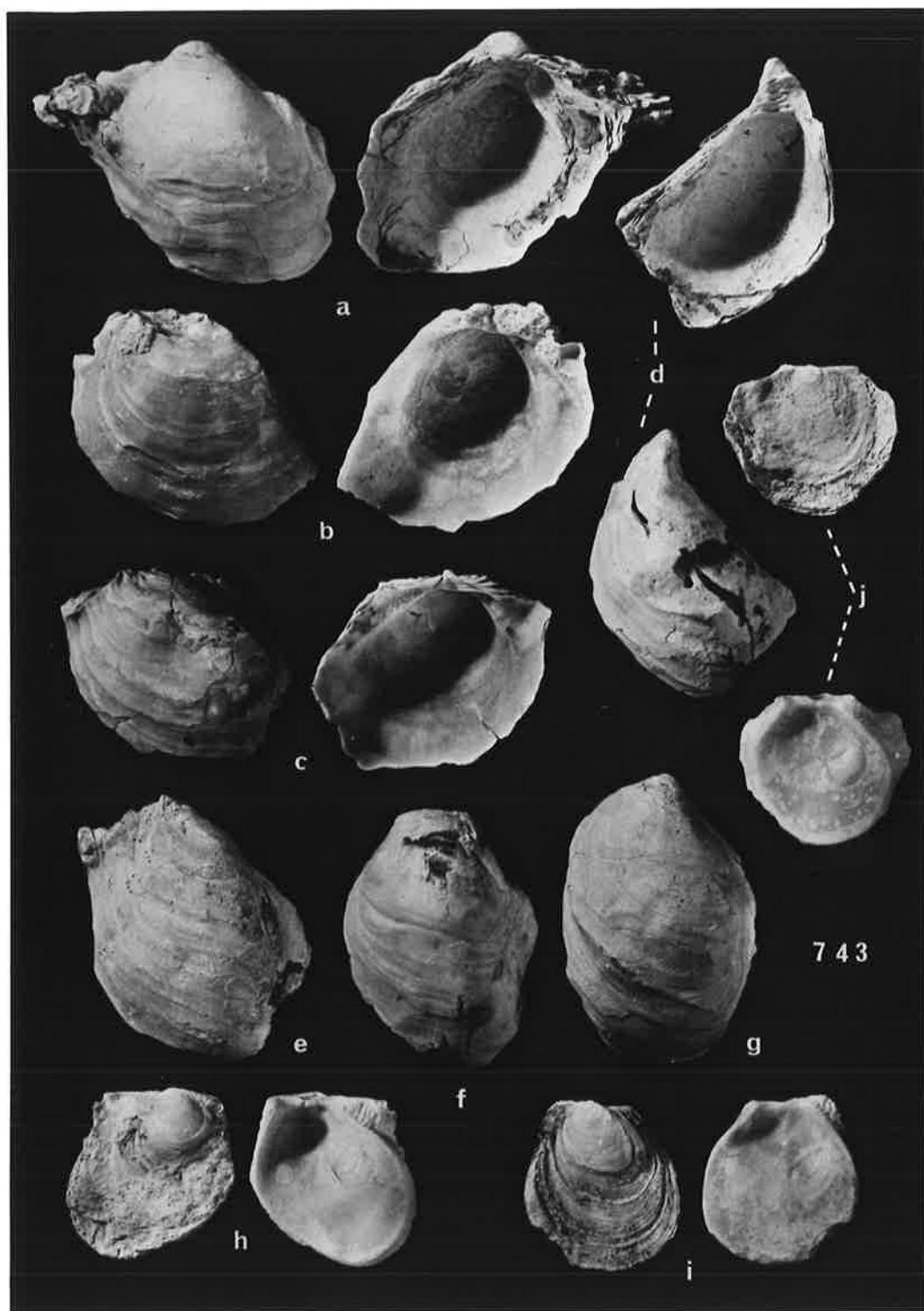


FIG. 743

P. (Phygraea) tarda (Hutton). SAM P 21260, morphae,
lot showing the variability LV and RV outlines and
morphology. LV: a) outer (x 1.23) and inner (x 1.11);
b-c) outer and inner (all x 1.17); d) outer and inner
(x 1.19); e) outer (x 1.17); f) outer (x 1.19); g)
outer (x 1.2). RV: h-j) inner and outer (all x 1.32).



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**NOTES ON THE GENUS *PSEUDOMALAXIS* FISCHER (MOLLUSCA:
GASTROPODA) AND ITS FOSSIL SPECIES IN AUSTRALIA**

By M. F. BUONAIUTO

ADELAIDE

February, 1975

Buonaiuto, M.F. (1975). Notes on the genus *Pseudomalaxis* Fischer (Mollusca Gastropoda) and its fossil species in Australia. *Transactions of The Royal Society of South Australia*, 99(1), 21-30.

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REVISION OF THE AUSTRALIAN TERTIARY SPECIES ASCRIBED
TO *LIMATULA* WOOD (MOLLUSCA, BIVALVIA)

By M. F. BUONAIUTO

ADELAIDE
February, 1977

Buonaiuto, M.F. (1977). Revision of the Australian tertiary species ascribed to Limatula Wood (Mollusca, Bivalvia). *Transactions of The Royal Society of South Australia*, 101(1), 21-33.

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REVISION OF THE COMPOSITE SPECIES *LIMA BASSI* TENISON
WOODS (MOLLUSCA, BIVALVIA)

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NOTE: This publication is included in the print copy of the thesis held in the University of Adelaide Library.