



Haemagglutinins of *Vibrio cholerae*: Molecular  
Characterization of the Mannose-Fucose Resistant  
Haemagglutinin (MFRHA).

Vicki L. Franzon.

Department of Microbiology and Immunology  
University of Adelaide,  
Adelaide, 5000  
Australia

A thesis Submitted for the degree of Doctor of Philosophy.

July, 1988.

## Abstract

The disease cholera is caused by *V. cholerae* of the O1 serotype. In contrast to organisms such as *Shigella* and *Salmonella*, *V. cholerae* is a non-invasive pathogen. It has been recognized that one of the most essential steps in the onset of cholera is the colonization of the small intestine. Hence considerable interest has been shown in identifying which factors may act as adhesins in the attachment of these organisms. Since 1961, when Bales and Lankford suggested that interaction between *V. cholerae* and red blood cells mimics that of the organism with the intestinal epithelium, a number of workers have become interested in the various haemagglutinins of *V. cholerae* and their properties. Hanne and Finkelstein (1982) have described four distinct haemagglutinins. One of these haemagglutinins is termed the mannose-fucose resistant haemagglutinin (MFRHA) and is found in all *V. cholerae* strains regardless of biotype. The general aim of this thesis is to report the first cloning, sequencing and detailed analysis of a gene encoding one of the *V. cholerae* haemagglutinins and to give some indication of whether the MFRHA protein may play a role in pathogenesis.

Chapter 3 describes the cloning and isolation of the MFRHA gene, characterization of its properties, localization of the coding region to within a 0.72 kb region and identification of the protein products using minicell analysis. The MFRHA gene was isolated from both biotypes and was shown to be identical. Chapter 4 analyzes the genetic organization of the MFRHA gene. This included sequencing of a 1,398 bp segment of *V. cholerae* DNA. Chapter 5 describes the construction of a deletion mutation in the MFRHA gene followed by insertion of an antibiotic marker and introduction of such a mutation into the *V. cholerae* chromosome.

Research of other Gram-negative pathogens suggests haemagglutinins are likely candidates for adhesins. Due to the number of *V. cholerae* haemagglutinins and the lack of characterization, one can only analyze their contribution by cloning the genes and introduction of specific mutations into the chromosome.

# Contents

<b>1</b>	<b>Review of the literature</b>	<b>1</b>
1.1	Introduction . . . . .	1
1.2	History of Cholera . . . . .	3
1.3	The Aetiological Agent . . . . .	4
1.4	Biotype Differentiation . . . . .	5
1.5	Serotype Differentiation . . . . .	5
1.6	Pathogenesis . . . . .	6
1.7	Cholera Toxin (CT) . . . . .	7
1.7.1	Mode of Action of CT . . . . .	8
1.7.2	Genetics of Cholera Toxin . . . . .	9
1.7.3	Antitoxic Immunity . . . . .	14
1.8	Adhesion . . . . .	15
1.8.1	Studies with brush border membranes and RBC's . . . . .	16
1.8.2	Studies with Intact Rabbit Intestinal Mucosa . . . . .	17
1.9	Slime envelope or Slime Agglutinin (SA) . . . . .	18
1.10	Lipopolysaccharide (LPS) . . . . .	19
1.10.1	Structure . . . . .	19
1.10.2	LPS genetics . . . . .	22
1.10.3	Anti-LPS immunity . . . . .	23
1.11	Flagellum, Flagellar Sheath and Proteins . . . . .	24
1.12	Fimbriae (pili) . . . . .	26
1.13	Outer Membrane Proteins . . . . .	28

1.14	Soluble Proteins . . . . .	31
1.14.1	Haemolysins (Hly) . . . . .	31
1.14.2	DNase . . . . .	34
1.14.3	Neuraminidase . . . . .	36
1.14.4	Soluble Haemagglutinin (SHA) . . . . .	36
1.15	Cell-associated Haemagglutinins . . . . .	40
1.15.1	D-mannose, D-fructose sensitive haemagglutinin . . . . .	40
1.15.2	Fucose-sensitive haemagglutinin . . . . .	42
1.15.3	Mannose-Fucose resistant haemagglutinin . . . . .	42
1.16	Aims of this Study . . . . .	43
<b>2</b>	<b>Materials and Methods</b>	<b>44</b>
2.1	Growth media . . . . .	44
2.2	Chemicals and reagents . . . . .	45
2.3	Enzymes . . . . .	46
2.4	Maintenance of bacterial strains . . . . .	46
2.5	Bacterial strains . . . . .	47
2.6	Plasmids . . . . .	47
2.7	Sources and preparation of red blood cells . . . . .	47
2.8	Haemagglutination assay . . . . .	48
2.9	Haemagglutination inhibition assay . . . . .	48
2.10	Assay for chemotaxis . . . . .	48
2.11	Antisera . . . . .	49
2.11.1	Antisera production . . . . .	49
2.11.2	Selective absorption of antiserum by intact cells . . . . .	49
2.12	Transformation procedure . . . . .	50
2.13	DNA extraction procedures . . . . .	50
2.13.1	Plasmid DNA isolation . . . . .	50
2.13.2	Preparation of <i>V.cholerae</i> genomic DNA . . . . .	52
2.14	Analysis and manipulation of DNA . . . . .	53

2.14.1	DNA quantitation . . . . .	53
2.14.2	Restriction endonuclease digestion of DNA . . . . .	53
2.14.3	Analytical and preparative separation of restriction fragments . . . . .	53
2.14.4	Isolation of DNA fragments less than 1,000bp . . . . .	54
2.14.5	Calculation of restriction fragment size . . . . .	54
2.14.6	<i>In vitro</i> cloning . . . . .	55
2.14.7	Generation of deletions using nuclease <i>Bal31</i> . . . . .	55
2.14.8	Dephosphorylation of DNA using alkaline phosphatase . . . . .	56
2.14.9	End-filling with Klenow fragment . . . . .	56
2.14.10	End-filling with T4 DNA polymerase . . . . .	57
2.14.11	Ligation of Linkers to blunt DNA ends . . . . .	57
2.14.12	Construction of gene banks . . . . .	57
2.14.13	Nick translation method . . . . .	58
2.14.14	Southern transfer and hybridization . . . . .	58
2.14.15	Colony hybridization . . . . .	59
2.15	Transposition with <i>Tn1725</i> . . . . .	59
2.16	Protein analysis . . . . .	60
2.16.1	Minicell procedures . . . . .	60
2.16.2	SDS Polyacrylamide Gel Electrophoresis . . . . .	60
2.16.3	Autoradiography . . . . .	61
2.16.4	Small scale cell envelope isolation . . . . .	61
2.16.5	Whole cell preparation . . . . .	61
2.16.6	Western transfer and protein blotting . . . . .	62
2.16.7	Colony transfer and blotting with antiserum . . . . .	62
2.17	M13 cloning and sequencing procedures . . . . .	63
2.17.1	Preparation of M13 replicative form (RF) DNA . . . . .	63
2.17.2	Cloning with M13mp18 and M13mp19 . . . . .	63
2.17.3	Transfection of JM101 . . . . .	64
2.17.4	Screening M13 vectors for inserts . . . . .	64
2.17.5	Purification of single-stranded template DNA . . . . .	64

2.17.6	Dideoxy sequencing protocol . . . . .	65
2.17.7	DNA sequencing gels . . . . .	67
2.17.8	Analysis of DNA sequences . . . . .	68
2.18	Animal experiments . . . . .	68
2.18.1	Infant mouse cholera model . . . . .	68
2.18.2	Virulence tests . . . . .	68
2.18.3	Adherence to HEp-2 cells . . . . .	69
<b>3</b>	<b>Molecular Cloning of the Mannose-Fucose-Resistant Haemagglu-</b>	
	<b>tinin of <i>Vibrio cholerae</i></b> . . . . .	<b>73</b>
3.1	Introduction . . . . .	73
3.2	Results . . . . .	74
3.2.1	Testing antiserum specificity . . . . .	74
3.2.2	Detection and isolation of the mannose-fucose resistant haemagglutinin clone . . . . .	76
3.2.3	Sugar inhibition and RBC activity . . . . .	78
3.2.4	Western blot analysis . . . . .	78
3.2.5	Effect of <i>tol</i> mutants on expression and cellular location of the cloned haemagglutinin. . . . .	81
3.2.6	Proteolytic activity . . . . .	83
3.2.7	Zincov inhibition . . . . .	85
3.2.8	Restriction analysis of pPM471 . . . . .	85
3.2.9	Localization of the DNA in pPM471 which encodes the haemagglutination activity . . . . .	88
3.2.10	Identification of the gene products of pPM471 . . . . .	96
3.2.11	Re-introduction of the cloned HA gene into <i>V. cholerae</i> . . . . .	98
3.2.12	Cloning of the MFRHA gene from the El Tor biotype . . . . .	100
3.3	Discussion . . . . .	102
<b>4</b>	<b>Genetic Organization of the Gene Encoding the MFRHA</b> . . . . .	<b>107</b>
4.1	Introduction . . . . .	107

4.2	Results . . . . .	108
4.2.1	Location of promoter . . . . .	108
4.2.2	Direction of transcription . . . . .	109
4.2.3	Generation of fragments for nucleotide sequencing . . . . .	112
4.2.4	Nucleotide sequence determination . . . . .	114
4.2.5	Regulatory sequences affecting expression of the MFRHA . . . . .	119
4.2.6	ORF1 signal sequence . . . . .	121
4.2.7	Codon usage . . . . .	123
4.2.8	Restriction endonuclease cleavage sites . . . . .	123
4.2.9	ORF 2 . . . . .	128
4.3	Discussion . . . . .	129
5	Construction of Defined Mutations in the <i>Vibrio cholerae</i> chromo- some . . . . .	133
5.1	Introduction . . . . .	133
5.2	Results . . . . .	134
5.2.1	Construction of a MFRHA deletion: type1 . . . . .	134
5.2.2	Construction of a MFRHA deletion: type 2 . . . . .	135
5.2.3	Insertion of a kanamycin resistance cartridge . . . . .	136
5.2.4	Subcloning into plasmid pRK290 . . . . .	136
5.2.5	Mobilization of pPM1147 from <i>E. coli</i> into <i>V. cholerae</i> . . . . .	138
5.2.6	Construction of a <i>V. cholerae</i> MFRHA <sup>-</sup> strain . . . . .	138
5.2.7	Colony hybridization . . . . .	138
5.2.8	Southern hybridization . . . . .	141
5.2.9	Distribution of MFRHA gene in <i>V. cholerae</i> . . . . .	144
5.2.10	Adherence to HEp-2 cells . . . . .	148
5.2.11	Virulence in the infant mouse cholera model . . . . .	148
5.2.12	Affect of motility . . . . .	148
5.2.13	Chemotaxis . . . . .	148
5.2.14	Virulence of motile strains in the infant mouse cholera model . . . . .	150

5.3	Discussion . . . . .	150
<b>6</b>	<b>Discussion</b>	<b>153</b>
6.1	Introduction . . . . .	153
6.2	Cloning and characterization of the gene encoding the MFRHA . . .	155
6.3	Localization of the coding region . . . . .	156
6.4	The MFRHA is distinct from the Tcp pilus . . . . .	156
6.5	Identification of protein products . . . . .	157
6.6	Nucleotide sequence determination . . . . .	159
6.7	Primer extensions . . . . .	160
6.8	Northern hybridization . . . . .	160
6.9	Construction of specific mutations . . . . .	161
6.10	Comparison with the Pap pilus . . . . .	161
6.11	Virulence . . . . .	163
6.12	Role of the MFRHA . . . . .	166
6.13	Future prospects . . . . .	167
<b>7</b>	<b>Bibliography</b>	<b>169</b>