



Phosphorylation by Aurora B kinase regulates caspase-2 activity and function

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Abstract

Mitotic catastrophe (MC) is an important oncosuppressive mechanism that serves to eliminate cells that become polyploid or aneuploid due to aberrant mitosis. Previous studies have demonstrated that the activation and catalytic function of caspase-2 are key steps in MC to trigger apoptosis and/or cell cycle arrest of mitotically defective cells. However, the molecular mechanisms that regulate caspase-2 activation and its function are unclear. Here, we identify six new phosphorylation sites in caspase-2 and show that a key mitotic kinase, Aurora B kinase (AURKB), phosphorylates caspase-2 at the highly conserved residue S384. We demonstrate that phosphorylation at S384 blocks caspase-2 catalytic activity and apoptosis function in response to mitotic insults, without affecting caspase-2 dimerisation. Moreover, molecular modelling suggests that phosphorylation at S384 may affect substrate binding by caspase-2. We propose that caspase-2 S384 phosphorylation by AURKB is a key mechanism that controls caspase-2 activation during mitosis.

Introduction

Mitosis is a highly dynamic process that is tightly regulated by a series of surveillance mechanisms that ensure the faithful segregation of chromosomes into the nascent daughter cells,

thereby maintaining genomic integrity. Errors in karyokinesis and/or cytokinesis during cell division generate cells carrying abnormal genomic content (i.e. aneuploidy or polyploidy) [1]. The chromosomal passenger complex (CPC) is a key factor in controlling the proper execution of both karyokinesis and cytokinesis, especially through the activity of its kinase subunit Aurora B kinase (AURKB), to prevent the formation of aneuploid and polyploid cells [2]. However, when mitotic errors cannot be corrected, they ultimately trigger mitotic catastrophe (MC), a key mechanism that prevents the proliferation and survival of mitotically aberrant cells through either regulated cell death, such as apoptosis, or cellular senescence [1, 3]. Failure of the MC process leads to persistent genomic instability, a well-characterised hallmark of cancer [4]. Therefore, MC is a critical mechanism to prevent cells from becoming tumourigenic.

Caspase-2, the most evolutionarily conserved member of the mammalian caspase family, was discovered over 2 decades ago [5]. Previous studies using knockout (KO) mice have demonstrated that caspase-2 deficiency promotes tumour development following replicative or oncogenic stress [6–10]. Recent studies have demonstrated that caspase-2 activation and its catalytic activity are critical steps in MC signalling. Our previous studies demonstrated that caspase-2-deficient cells acquire extensive aneuploidy following replicative stress in culture [11], following

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prolonged mitotic arrest caused by inhibition of polo-like kinase 1 (Plk1) [12] and in the bone marrow of aged caspase-2 KO mice [13]. This was partly caused by decreased Bid cleavage, reduced cell death, and clonogenic survival of aberrant mitotic cells [12]. Importantly, cells from mice harboring catalytic-dead caspase-2 (*Casp2*^{C320S}) also showed increased aneuploidy following prolonged mitotic arrest [12], suggesting importance of the activation and enzymatic activity of caspase-2 in apoptotic death of aneuploid cells. Other studies have demonstrated that caspase-2 activation following cytokinesis failure following AURKB inhibition triggers cleavage of MDM2, leading to p53 stabilisation and cell cycle arrest, as an alternative mechanism to prevent aneuploidy and polyploidy [14, 15]. Although this evidence shows that caspase-2 activation and its activity are required to prevent survival and proliferation of cells with mitotic defects, the molecular mechanisms have not been well defined.

Previous studies indicate that activation of caspase-2 can be modulated by phosphorylation in different experimental conditions [16–18]. Caspase-2 activation is inhibited by phosphorylation at S157 by casein kinase-2 in TNF- α -related apoptosis-inducing ligand (TRAIL)-mediated apoptosis in cultured cells [17], S308 (S340 in human) by Cdk1-cyclin B1 during mitosis [16] and S135 (S164 in human) by calmodulin-dependent protein kinase II (CaMKII) in *Xenopus oocytes* under nutrient rich conditions [18]. S340 is localised in the linker region between the large (P19) and small (P14) subunits, suggesting that phosphorylation at these sites acts to prevent caspase-2 processing and activation [16]. Recently, phosphorylation at T180 of caspase-2 by mitogen-activated protein kinase (MAPK) is involved in lipid metabolism [19]. However, it is not known whether phosphorylation at these sites regulates caspase-2 activation and its activity in response to aberrant and/or failed mitosis.

In this study, we identified a number of previously unknown phosphorylation sites in caspase-2. Specifically, we demonstrate AURKB phosphorylates caspase-2 at the highly conserved S384 residue within the small subunit. Furthermore, our data indicate an alternative MC regulatory mechanism through AURKB-mediated inhibitory phosphorylation of caspase-2 and suggest that inhibition of AURKB activity is required to trigger apoptosis or cell cycle arrest following failed mitosis.

Results

Identification of phosphorylation sites in caspase-2

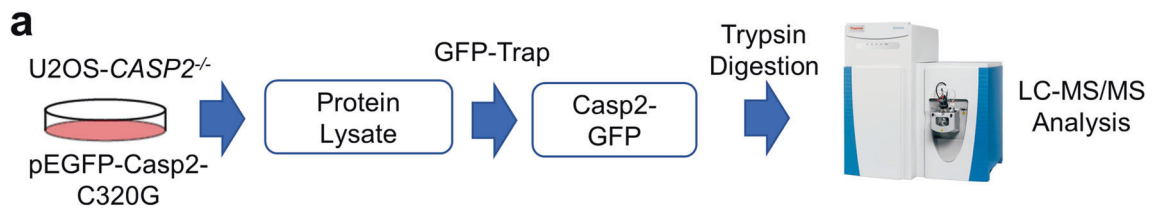
In this study, we mainly utilised U2OS cells as these cells have been used in similar studies [12, 14], arrest effectively in mitosis [20, 21] and undergo cell death following mitotic

exit upon treatment with mitotic drugs [22, 23]. Previous studies have shown that phosphorylation can inhibit caspase-2 activation or regulate its interaction with other molecules under different physiological conditions [16–19]. Therefore, we set out to define all potential caspase-2 phosphorylation sites in viable cells in culture. To identify phosphorylation sites in caspase-2, liquid chromatography-tandem mass spectrometry (LC-MS/MS) experiments were conducted using trypsin-digested GFP immunoprecipitates from U2OS-CASP2^{-/-} cells transiently expressing GFP-tagged catalytically inactive mouse caspase-2-C320G. (Fig. 1a and Supplementary Fig. S1). The annotated peptide sequence demonstrates all of the peptide fragments used for phosphorylation site identification. In total, we identified 11 phosphorylation sites in caspase-2 (Fig. 1b, c), including previously characterised residues S157, S164, T180 and S340 [16, 18, 19], and six novel sites S24, S80, T161, S220, S346 and S384. Phosphorylation at T158 in human caspase-2 has also been reported in the PHOSIDA database, but its role is functionally undefined [24].

Next, multiple amino acid sequence alignment showed that S80, S157, T158, T180 and S220 are conserved in mammals, whereas S340 and S346 are also conserved in *Xenopus* caspase-2. Interestingly, S164 and S384 are most highly conserved, especially S384, which is remarkably well conserved among the different species including *Drosophila* apical caspase Dronc and *C. elegans* CED3 (Fig. 1d). It is important to note that Dronc and CED3 respectively are the only CARD containing caspases in flies and nematodes and are functionally analogous to both mammalian caspase-2 and caspase-9 [25, 26]. This finding suggests that caspase-2 S384 residue might be functionally important.

Phosphorylation regulates caspase-2 activation and function

The first step in caspase-2 activation is homodimerisation via its CARD, followed by autoprocessing, and ectopic expression of caspase-2 triggers cell death [27–29]. To functionally characterise the newly identified phosphorylation sites in caspase-2, we generated phospho-mimetic (Ser/Glu) and phospho-deficient (Ser/Ala or Thr/Val) mutants for each residue and then examined whether transient expression of these mutants affected the processing and activation of caspase-2, by assessing the cleavage of its substrates Bid and MDM2 [14, 15, 30]. As expected, WT caspase-2-GFP but not the C320G mutant cleaved MDM2, generating a p60 cleavage fragment (60 kDa) (Fig. 2a). While most of the phosphorylation mutants could cleave MDM2 to some extent, MDM2 cleavage was completely abolished by expression of the S384E caspase-2 mutant (Fig. 2a). Similarly, we found that all the phosphorylation

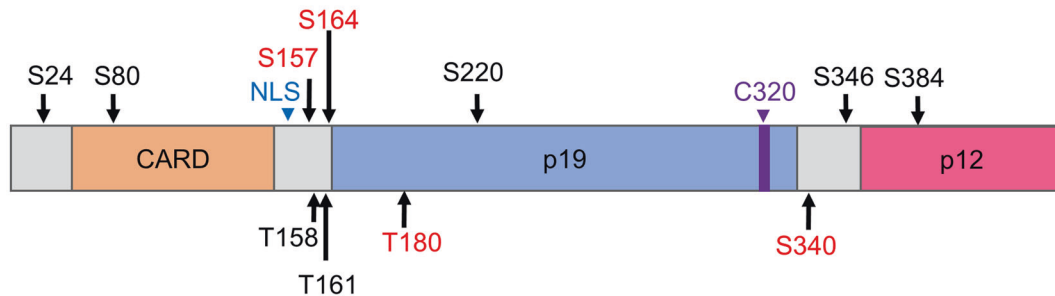


b

Caspase-2 phosphorylation sites identified by LC-MS/MS

Peptide	Position in peptide	Charge	Mass error [ppm]	Peptide location	pS/T site	Localization probability
S(ph)RILAVCGMHPDHQETLK	1	4	-0.4984	24-41	24	1.00
GGs(ph)FSQNVELLNLLPK	3	2	-2.498	78-93	80	0.99
QLRLS(ph)TDATEHS(ph)LDNGDGPPCLLVK	5	3	-0.59	153-177	157	0.98
LST(ph)DAT(ph)EHSLDNGDGPPCLLVK	3	2	-2.3533	156-177	158	0.71
LS(ph)T(ph)DAT(ph)EHS(ph)LDNGDGPPCLLVK	6	3	-1.1089	153-177	161	0.96
LS(ph)T(ph)DAT(ph)EHS(ph)LDNGDGPPCLLVK	9	2	-1.7132	153-177	164	1.00
PCT(ph)PEFYQAHY(ph)QLAYR	3	3	-0.2975	178-193	180	1.00
S(ph)GGDVDHTTLVTLFK	1	2	-0.1515	220-234	220	1.00
NHT(ph)QS(ph)PGCEESDAGKEELMK	5	3	-1.2279	336-355	340	0.98
NHTQS(ph)PGCEES(ph)DAGKEELMK	11	3	-0.585	336-355	346	0.98
RGS(ph)WYIEALTQVFSER	3	2	0.279	382-397	384	1.00

c



d

	24	80	157	158 161	164	180
Human	AADRGRIL	QAKVGS	KLRLS	TDVTEH	SLDNKDG	VCLQVKPCTPE
Mouse	AADRRSRIL	QAKGGS	QLRLS	TDATEH	SLDNGDG	PCLLVKPCPE
Rat	AADRRSRIL	QAKGGS	QSRLS	TDTEH	SLDNGDG	PCLQVKPCPE
Xenopus	-----	MAYRSD	GRQIC	REYREE	SIDDGDG	VTVQL--CSVN
CED3	-----	NSCGTV	NSSFTGCSS	LGYSRRNR	SFSKASG	TQYIFHEEDMN
Dronc	--MQPPEL	QDLNGKPFNMD	KLRNE--P	LGALTP	YVGVVDG	PEVKKSKKIHGDSA

	220	340	346	384
Human	EKELEFRSGGDVD	HAGSPGCEESDAG	TAAMRNTRKGSWYIEAL	
Mouse	EKDLEFRSGGDVD	HTQSPGCEESDAG	NAAMRNTRKGSWYIEAL	
Rat	EKDLEFRSGGDVD	HAQSPGCEESDAG	NAAMRNTRKGSWYIEAL	
Xenopus	TPDLDYRCGGEVD	QSGSPGCEQSDAG	TVSLRNTRKGSWVQDL	
CED3	--QMPTRNGTKAD	AFLRRGWDNRDGP	YVSWRNSARGSWFIQAV	
Dronc	-DQNRIRIGAEKD	NLMPEVYTAQEEL	YVTHRDLDTGSWYIQKF	

site mutants, except S384E, showed reduced levels or loss of full-length Bid, indicating complete cleavage. The C320G mutant did not induce cleavage of Bid as expected

(Fig. 2b). We also observed reduced autoprocessing of S384E, to a similar extent as seen with C320G. This was shown by higher levels of full-length caspase-2-GFP protein

Fig. 1 Proteomic analysis identified six new phosphorylation sites in caspase-2. **a** Schematic diagram of phosphorylation site analysis of caspase-2. **b** Caspase-2 phosphorylation sites identified by LC-MS/MS. The phosphorylated amino acid position, charge, mass error, peptide location in caspase-2, phosphorylated S/T site position in caspase-2 and localisation probability are indicated. Phosphorylated residues are indicated in brackets. Spectra for the peptides in (b) can be found in Supplementary Fig. S2. **c** Phosphorylation sites in caspase-2 identified by mass spectrometry. Functionally reported sites are indicated in red. S (Ser) or T (Thr) (in black) indicates positions of novel potential phosphorylation sites. NLS nuclear localisation signal. C320 catalytic Cys. **d** Multiple amino acid sequence alignment showing conservation of phosphorylation sites in caspase-2 from different species. Newly identified and known phosphorylation sites with high homology are shaded in purple and grey, respectively.

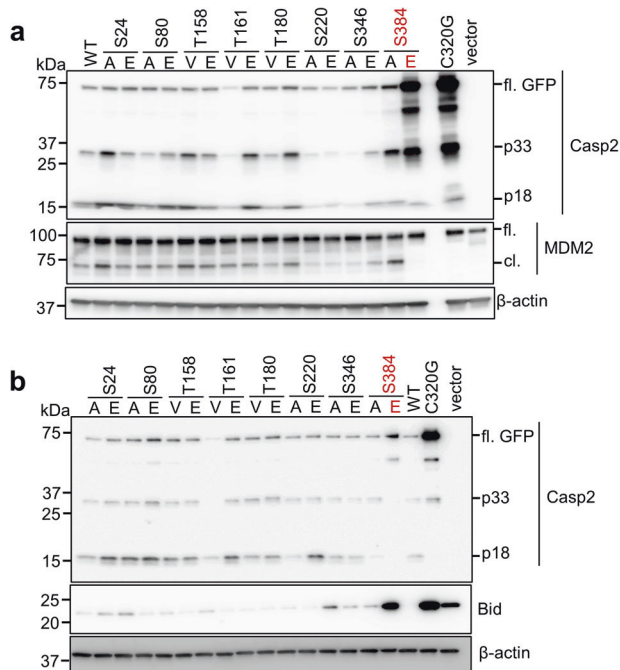


Fig. 2 Caspase-2-S384E mutant does not cleave MDM2 or Bid. **a** U2OS-*CASP2*^{-/-} cells were transfected with 1 μg of GFP mock vector, or GFP-tagged caspase-2 WT, C320G, or phosphorylation site mutants as indicated. Cell lysates were subjected to immunoblotting with the indicated antibodies. β-actin was used as loading control. **b** U2OS-*CASP2*^{-/-} cells were co-transfected with 1 μg of GFP mock vector, or GFP-tagged caspase-2 WT, C320G, or phosphorylation site mutants along with 500 ng of Bid-HA. Cell lysates were subjected to immunoblotting with the indicated antibodies. β-actin was used as loading control.

(74 kDa) and reduced levels of the 18 kDa subunit band compared to WT caspase-2 or other phospho site mutants (Fig. 2a, b). These results demonstrate that the caspase-2 phospho-mimetic S384E mutation affects its autoprocessing and catalytic activity. Consistent with these findings, and similar to the C320G mutant, the expression of S384E also exhibited reduced ability to promote cell death compared to

WT or S384A in *Casp2*^{-/-} immortalised mouse embryonic fibroblasts (MEFs) (Supplementary Fig. S3a, b).

We next assessed the enzymatic activity of the various caspase-2 phosphorylation mutant cell lysates using an in vitro caspase activity assay with the selective fluorogenic caspase-2 substrate, VDVAD-AFC [31, 32]. Several mutants showed significantly reduced VDVADase cleavage activity compared to WT caspase-2, including T180V, S220A, S220E, S346A, S384A and S384E (Supplementary Fig. S4a). Interestingly, only the S384E mutant showed a complete absence of enzymatic activity, similar to C320G or the vector control (Supplementary Fig. S4a), further demonstrating that mutation of this site impairs caspase-2 processing and substrate cleavage (Fig. 2). As the S384A mutation also showed reduced enzymatic activity in vitro, we generated additional unphosphorylatable mutants, including Ser-to-Gly (small amino acid) and Ser-to-Thr (uncharged, polar amino acid like serine) mutations at this site. We observed that, similar to S384A, both the S384G and S384T mutants were still able to cleave MDM2 and could also be processed in transfected cells (Supplementary Fig. S4b). Together, these results indicate that phosphorylation at S384 inhibits caspase-2 activation and activity in cells and that several phosphorylation site mutations can affect the enzymatic activity of caspase-2, but they do not impair its substrate cleavage and cell death inducing activity.

Caspase-2-S384E does not disrupt dimerisation

As homodimerisation is sufficient for initial caspase-2 activation [27], we examined whether the S384E mutation affected caspase-2 dimerisation using the bimolecular fluorescence complementation (BiFC) assay, a visualizing tool of protein–protein interaction [12, 33], with full-length caspase-2-S384E and CASP2-C320A. We blocked mitosis using the Plk1 inhibitor BI2536 (BI), which we have previously demonstrated to result in caspase-2 dimerisation [12]. There were no differences in dimerisation of caspase-2-S384E or CASP2-C320A between control (DMSO) and BI treatment (Fig. 3a, b). Expression of GST recombinant caspase-2-WT, C320G or S384E in *E. coli* showed that WT GST-caspase-2 was cleaved to form an intermediate (~50 kDa) and fully processed large subunit (18 kDa), whereas GST-caspase-2-S384E was not processed and behaved like the recombinant C320G protein (Fig. 3c). This demonstrates that caspase-2-S384E substitution prevents autoprocessing of caspase-2. Caspase assays using VDVAD-AMC showed robust catalytic activity by WT caspase-2 but not by both the C320G and S384E (Fig. 3d), which further confirmed that caspase-2-S384E mutant protein lacks catalytic activity. Our data suggest that phosphorylation at S384 does not prevent the initial caspase-2 dimerisation event, but

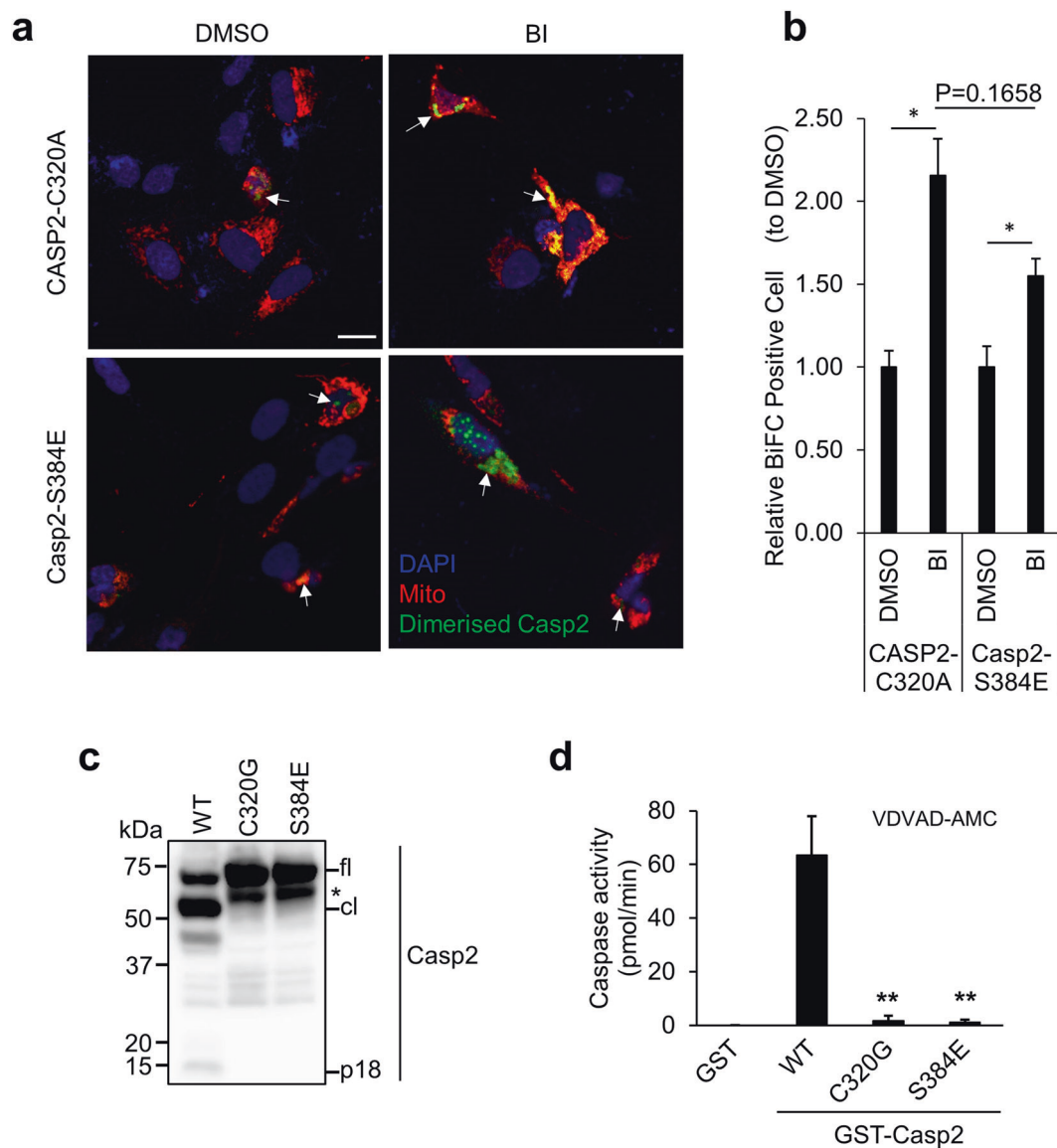


Fig. 3 Caspase-2-S384E mutant can still homodimerise but lacks enzymatic activity. **a** Representative confocal images showing dimerisation of Casp2-S384E or CASP2-C320A by BiFC. U2OS-*CASP2*^{-/-} cells were transfected with Casp2-S384E or CASP2-C320A BiFC constructs, respectively, together with pDsRED-Mito as transfection reporter and treated with DMSO or 100 nM BI2536 (BI) for 24 h. All cells were also treated with z-VAD (20 μ M), pan-caspase inhibitor. DAPI (blue); pDsRED-Mito (red). Arrows indicate dimerised caspase-2 (Green). Scale bar = 20 μ m. **b** Quantitation of relative

BiFC-positive cells are indicated. * $p < 0.05$. **c** Immunoblot analysis of recombinant GST-caspase-2-WT, C320G and S384E protein expression and autoprocessing. *, truncated GST-caspase-2. **d** Caspase activity of recombinant GST-caspase-2-WT, C320G and S384E protein, was assessed by cleavage of VDVAD-AMC. GST protein was used as a control. Data represented as mean \pm SEM from three independent experiments. ** $p < 0.01$ compared to GST-caspase-2-WT.

inhibits autoproteolysis and downstream activation, and therefore likely exerts a dominant negative function.

Caspase-2 S384 is a target of Aurora B kinase

We used computational tools and databases including PHOSIDA, Scansite and KinasePhos 2.0 and identified AURKB as a potential kinase for phosphorylation of S384 in caspase-2. The consensus phosphorylation motifs for

AURKB are (K/R)1-3-X-(S/T) or (K/R)-(R/K)-X0-2-(S/T), where X indicates any amino acid [34, 35]. Such a sequence is present around S384 in caspase-2 (Fig. 1d). We first performed a GST-Casp2 pull-down assay to examine whether AURKB directly binds to caspase-2 and found that GST-Casp2 full-length (GST-Casp2-fl) coprecipitated with AURKB (Fig. 4a). Furthermore, a small peptide fragment of caspase-2 comprising the S384 residue (aa 363–423; GST-Casp2₃₆₃₋₄₂₃) also coprecipitated with AURKB (Fig. 4b).

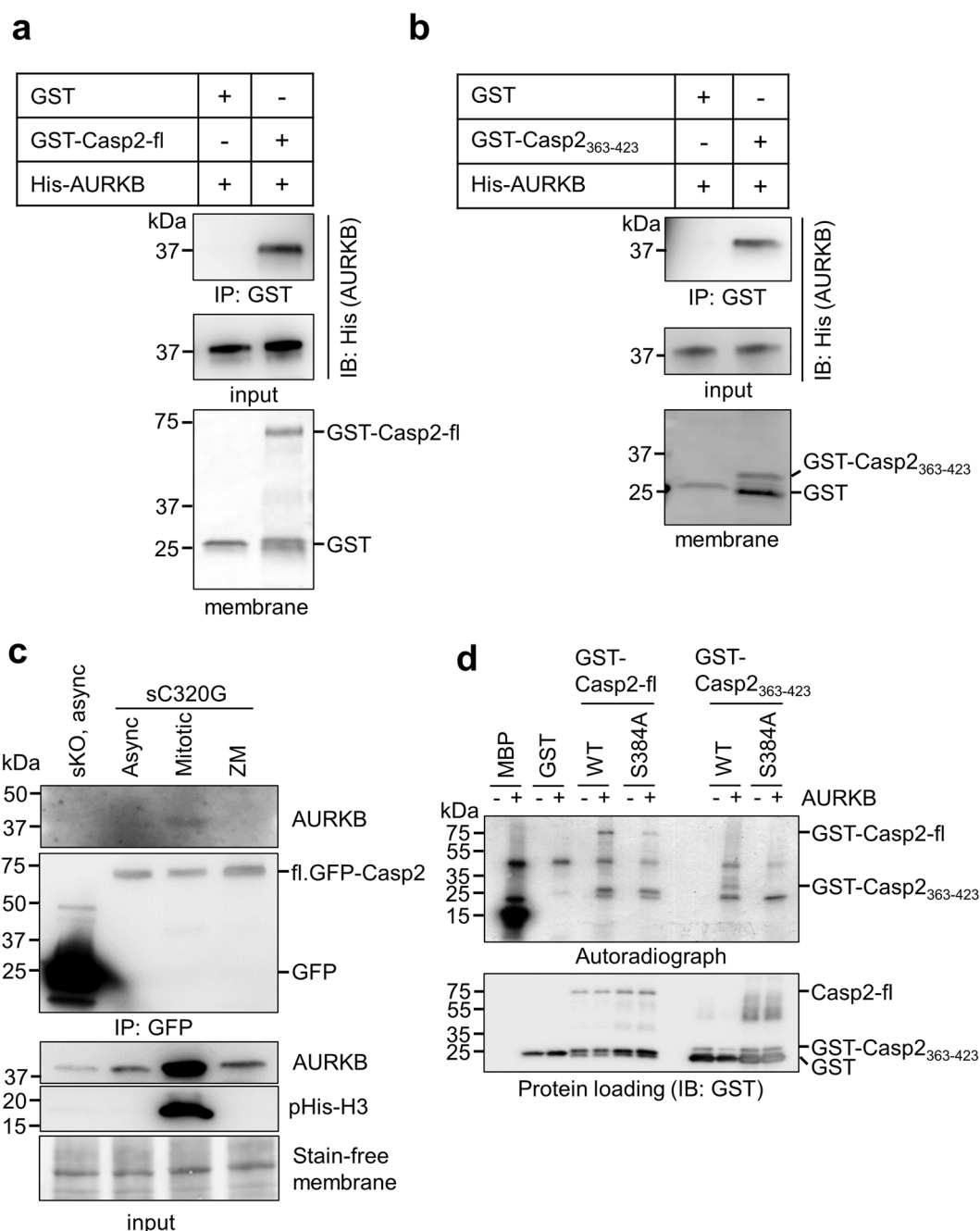


Fig. 4 Aurora B kinase phosphorylates caspase-2 at S384 in vitro. **a, b** In vitro interaction between purified recombinant GST-Casp2-C320G (GST-Casp2-fl) or GST-Casp2₃₆₃₋₄₂₃-WT and His-AURKB was determined by GST pull-down assay. Stain-free membrane shows protein loading amount for GST or GST-Casp2. Antibodies used are indicated. **c** U2OS-CASP2^{-/-} cells stably expressing GFP (sKO) or GFP-caspase-2-C320G (sC320G) were used. GFP or GFP-caspase-2-C320G were immunoprecipitated (IP) with GFP-Trap from the asynchronous (Async), mitotic or ZM447439 (ZM)-treated cells

and subjected to immunoblotting with the indicated antibodies. Phospho-histone H3 (pHis-H3) was used as mitosis marker. Stain-free membrane was used as loading control. **d** GST-Casp2-fl WT, GST-Casp2-fl S384A, GST-Casp2₃₆₃₋₄₂₃ WT, GST-Casp2₃₆₃₋₄₂₃ S384A or GST was subjected to in vitro phosphorylation by incubation with AURKB and [γ -³²P]-ATP and analysed by autoradiography. IB with GST antibody shows equal loading. MBP, myeloid basic protein was used as positive control for AURKB phosphorylation.

To examine the interaction between caspase-2 and AURKB in cells, we performed a coimmunoprecipitation (Co-IP) experiment using U2OS-CASP2^{-/-} cells stably expressing GFP (sKO) or GFP-caspase-2-C320G (sC320G). AURKB

coimmunoprecipitated with caspase-2 predominantly in the mitotic cell fraction but not in asynchronous or AURKB inhibitor (ZM) treated cells (Fig. 4c). Next, we performed an in vitro phosphorylation assay using AURKB with either

GST-Casp2-fl WT or the S384A mutant. The result demonstrated that full-length caspase-2 is phosphorylated by AURKB (Fig. 4d). Furthermore, *in vitro* phosphorylation using GST-Casp2₃₆₃₋₄₂₃ WT or S384A confirmed that S384 of caspase-2 is phosphorylated by AURKB (Fig. 4d). Notably, AURKB phosphorylation was almost completely abolished in the GST-Casp2₃₆₃₋₄₂₃ S384A fragment but was only diminished in the GST-Casp2-fl S384A (Fig. 4d), suggesting that AURKB might phosphorylate additional residues besides S384. Interestingly, Aurora A kinase (AURKA) did not phosphorylate S384 (Supplementary Fig. S5), indicating that this residue is a specific site for AURKB. Together, these data suggest that S384 of caspase-2 is target of AURKB and is predominantly phosphorylated in mitosis.

Caspase-2-S384E expression results in increased polyploidy and resistance to cell death following mitotic stress

To further characterize the functional consequence of phosphorylation at S384, we established a series of U2OS cells stably expressing GFP (sWT) or U2OS-CASP2^{-/-} cells stably expressing GFP (sKO), GFP-tagged caspase-2-C320G (sC320G) or S384E (sS384E clones #1 and #2) (Supplementary Fig. S6). These cell lines were treated with the AURKB inhibitor ZM447439 (ZM) [36] to induce polyploidy and then cleavage of caspase-2 and its substrate MDM2, was analysed by immunoblotting (IB) and DNA content was measured by flow cytometry. Consistent with previous reports [14, 15], sWT cells showed cleavage of caspase-2 (p18 kDa), MDM2 and increased p53 and p21 accumulation in response to AURKB inhibition (Fig. 5a). The sKO cells still showed increased p53 levels following ZM treatment, albeit less than sWT cells, but p21 levels were not increased and cleavage of MDM2 was absent, which is consistent with previous findings (Fig. 5a) [14]. Interestingly, similar to sKO and sC320G, the two independent sS384E cell lines also showed absence of MDM2 cleavage and p21 increase in response to ZM, while still exhibiting induction of p53 (Fig. 5a). DNA content analysis of sKO cells showed significantly increased polyploid (>4*N*) cell accumulation compared to sWT cells (Fig. 5b, c), which is consistent with previous reports [14, 15]. Similarly, sC320G and sS384E cell lines also showed increased polyploidy following ZM treatment (Fig. 5b, c).

We also used an alternative approach to induce polyploidy by treating cells with blebbistatin to inhibit non-muscle myosin II, which plays an essential role in the constriction of the actomyosin contractile ring during cytokinesis [37, 38]. Importantly, similar results were observed when the U2OS stable cell lines were treated with blebbistatin, although the accumulation of p53 reduced

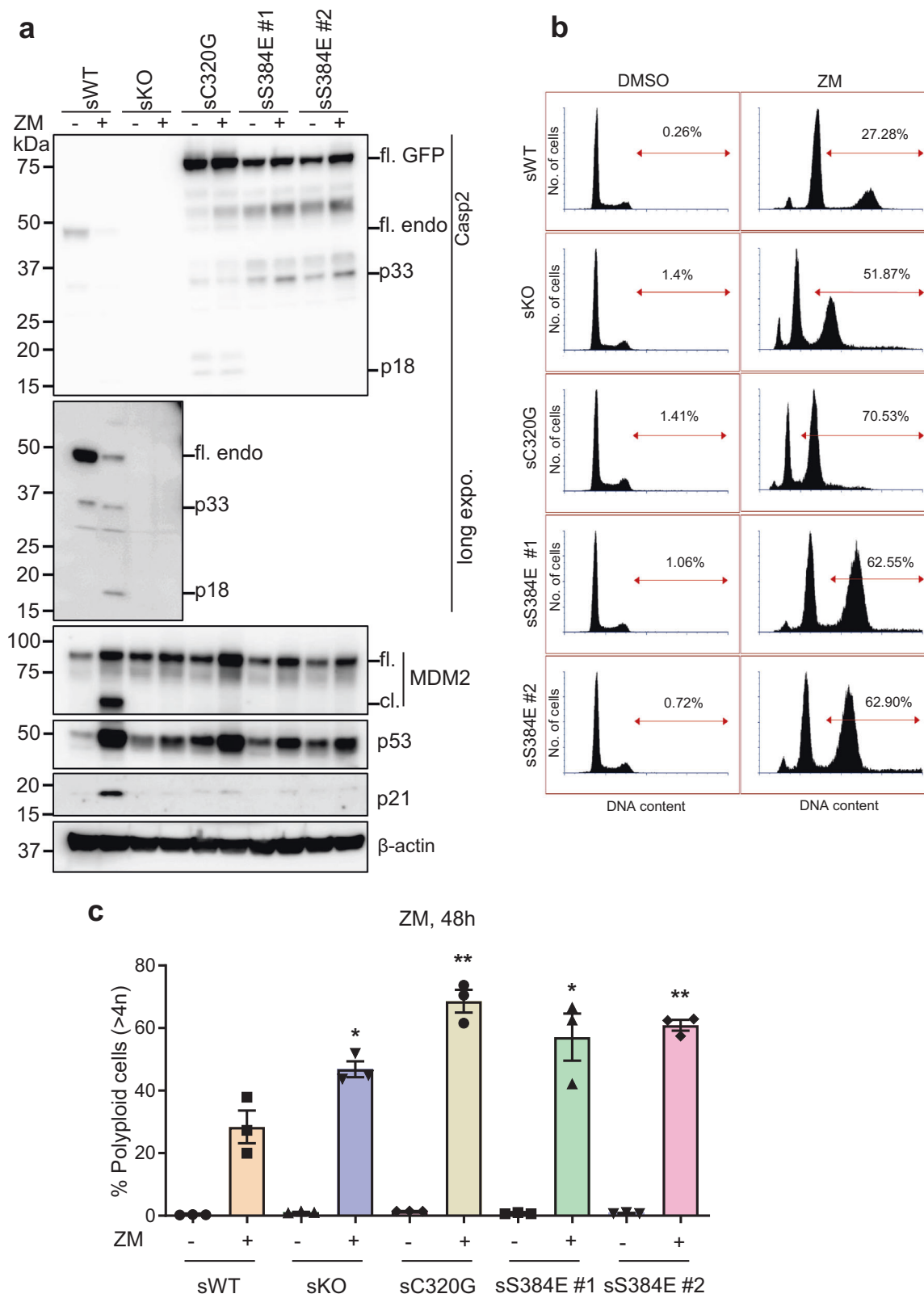
(Fig. 6a), particularly when compared to treatment with the AURKB inhibitor, AZD1152 [36] (Supplementary Fig. S7a) in U2OS parental and CASP2^{-/-} cells. Polyploidy was also significantly increased in sKO, sC320G and sS384E cell lines compared to the sWT although the increase is not as robust as with AURKB inhibitor treatment (Fig. 6b and Supplementary Fig. S7b).

We also treated U2OS stable cell lines with the Plk1 inhibitor, BI2536 (BI), which induces mitotic arrest, chromosome missegregation, aneuploidy and caspase-2 dependent apoptotic cell death [12]. Following treatment with BI, the sKO, sC320G and sS384E cell lines were more resistant to apoptosis than sWT cells (Fig. 7a). The sWT cells also showed caspase-2 activation, cleavage of PARP, caspase-3 and Bid, demonstrating BI-induced cell death (Fig. 7b). As previously reported, sKO and sC320G cells showed reduced cleavage of PARP, Bid and caspase-3 [12] and this was also observed in the sS384E cell lines (Fig. 7b). Taken together, these experiments confirmed that phosphorylation of S384 inhibits caspase-2 activity and its cell death function.

We then examined whether re-expression of caspase-2 WT, S384A or S384E can rescue caspase-2-mediated MC signalling, in two different cell lines, U2OS-CASP2^{-/-} and A549-CASP2^{-/-} cells. Only the caspase-2 WT and S384A expressing cells showed caspase-2 activation and complete processing (p18) following ZM treatment, whereas C320G and S384E did not (Supplementary Fig. S8a, b). Also, MDM2 cleavage was observed in caspase-2 WT or S384A expressing U2OS-CASP2^{-/-} and A549-CASP2^{-/-} cells, but not in cells expressing C320G or S384E (Supplementary Fig. S8a, b). Interestingly, more MDM2 cleavage was observed in S384A expressing cells compared to the cells expressing WT caspase-2. These data further indicate that phosphorylation at S384 in caspase-2 is critical to regulate caspase-2 activation and activity in MC.

Discussion

In the present study, we identified six new phosphorylation sites in mouse caspase-2 by LC-MS/MS. We used the catalytically inactive caspase-2-C320G mutant, because unlike WT caspase-2, it is not associated with cytotoxicity when expressed ectopically [27]. This allowed us to identify new phosphorylation sites associated with full-length caspase-2 zymogen. Of the six newly identified phosphorylation sites, we found that S384, which is highly conserved, is critical for the regulation of caspase-2 activity and function. Our studies utilising re-expression of caspase-2 mutants in caspase-2 deficient cells demonstrated that the caspase-2-S384E phospho-mimetic mutant prevents caspase-2 activation and its catalytic activity. Firstly, S384E caspase-2



shows reduced autoprocessing of recombinant protein in cells. While we observed some caspase-2 cleaved products (p33 and p18) in S384E and C320G overexpressing cells,

recombinant protein did not show caspase-2 autoprocessing or catalytic activity. These cleaved products were reduced or absent when the C320G or S384E mutants were

◀ **Fig. 5 Caspase-2-S384E expressing cells fail to cleave MDM2 and show increased polyploidy following AURKB inhibition.** GFP-expressing U2OS (sWT) and U2OS-*CASP2*^{-/-} expressing GFP (sKO), GFP-caspase-2-C320G (sC320G) or GFP-caspase-2-S384E (sS384 #1 and #2) were treated with DMSO or 2 μM ZM447439 (ZM) for 48 h and subjected to immunoblot and DNA content analysis. **a** Representative immunoblots (from three independent experiments) of cell lysates from treated stable cell lines. Antibodies used for immunoblotting are as indicated. β-actin was used as loading control. **b** Representative flow cytometric profiles of the DNA content in cells following ZM treatment. The percentage represents polyploid cells (>4*N*), which was calculated using FACS Express 6 cell cycle analysis module. **c** Graph comparing percentage of cells with polyploid (>4*N*) DNA content following ZM treatment. Mean ± SEM; *n* = 3. **p* < 0.05; ***p* < 0.01 compared to sWT + ZM.

transiently expressed in U2OS-*CASP2*^{-/-}/*CASP3*^{-/-} cells (Supplementary Fig. S9) indicating the cleavage is likely caused by caspase-3 [39] or other cellular proteases. Secondly, S384E was unable to cleave the known caspase-2 substrates, MDM2 and Bid. Thirdly, caspase-2-S384E can still homodimerise but is catalytically inactive. Lastly, expression of the caspase-2-S384E shows predominantly nuclear localisation, similar to the C320G mutant, which may also impede activation by other caspases in the cytosol. Overall our results suggest that while S384E does not affect dimerisation, the first step of caspase-2 activation, it prevents autoproteolytic processing, cytosolic localisation and complete caspase-2 activation. This may be in part caused by steric hindrance on the substrate binding site. These data suggest that S384E caspase-2 likely acts as a dominant negative protein.

Caspase-2 is the only member of the caspase family that preferentially recognises a pentapeptide, such as VDVAD or ADVAD, as substrates over a tetrapeptide sequence [31, 32, 40]. T380 and Y420 are key residues that have been shown to be critical for binding the P5 residue [41]. These residues form hydrogen bonds with P5 Val in VDVAD or Ala in ADVAD pentapeptide substrates [41]. Also, W385 forms a hydrogen bond with P4 Asp. Interestingly, S384 localises next to W385 and close to T380 although it may not directly interact with the substrate [41]. This suggests that phosphorylation at S384 may cause an interference with substrate binding (Fig. 8a, b) preventing the catalytic activity of caspase-2.

We examined four different substitution mutants of S384 and found that only the S384E phospho-mimetic mutant was enzymatically inactive. Although various other amino acid substitutions on S384 may affect cleavage ability of caspase-2, we have shown here that the region directly surrounding S384 of caspase-2 can bind to AURKB in vitro and in vivo (Fig. 4a–c), and S384 is phosphorylated by AURKB (Fig. 4d). In addition, expression of the phospho-deficient S384A mutant in caspase-2 deficient cells, can rescue the cleavage of MDM2 following aberrant mitosis.

However, phosphorylation of S384 is unable to rescue MC signalling (as demonstrated by expression of S384E) (Supplementary Fig. S8). These findings, demonstrate that phosphorylation of S384 by AURKB is critical to prevent caspase-2 activation and activity.

Multiple kinases are responsible for phosphorylation of caspases [42]. Our study identified AURKB as an important additional kinase that phosphorylates caspase-2 at S384. We also examined other kinases (including CaMKII, PKA, PAK1, Cdk1, Plk1 and MAPK) but none of them phosphorylated the S384 site in caspase-2 (data not shown). Analysis with the GPS 5.0 software (<http://gps.biocuckoo.cn>) predicted five possible additional AURKB phosphorylation sites in caspase-2, of which only two, S24 and S157, were identified by MS in this study. We tested S24 phospho-mutants but found this residue was not essential for caspase-2 function. While S157 may be an AURKB target site, our data indicate that phosphorylation on S384 is sufficient to prevent activation and catalytic activity of caspase-2. AURKB has various critical functions in mitosis, including chromatin condensation, kinetochore-microtubule error correction and cytokinesis [43]. Therefore, our findings suggest that AURKB phosphorylation maintains caspase-2 in an inactive state throughout cell division. Notably, caspase-2 is also regulated by Cdk1-mediated inhibitory phosphorylation at S340 during mitosis [16]. However, while Cdk1 activity is inhibited following degradation of its cyclin B partner at anaphase onset, AURKB remains active until late cytokinesis [44]. Importantly, AURKB, along with its CPC partners, can delay the final separation (i.e. abscission) of the two daughter cells in the presence of lagging chromatin at the cleavage site to prevent aneuploidy and polyploidy [45]. Therefore, it appears that caspase-2 is under tight regulation by multiple mitotic kinases to prevent any accidental cell death during cell division. Our studies using the S384E mutant expression in U2OS cells and recombinant S384E caspase-2 protein in vitro demonstrate that phosphorylation at S384 prevents caspase-2 catalytic activity and function. Therefore, the inhibition of S384 phosphorylation by specific targeting of this site, or by targeting AURKB may be an effective mechanism of activating caspase-2 in cycling cells. However, it would be important to decipher whether S384 dephosphorylation could effectively activate caspase-2 during other cell cycle stages or in senescent cells. Interestingly, blebbistatin, which does not inhibit AURKB, also causes cytokinesis failure and caspase-2 activation indicating potentially differential regulation of caspase-2 at different mitotic stages. Importantly, the caspase-2-S384E was also not activated following blebbistatin treatment, suggesting an important role for AURKB-mediated phosphorylation in regulating caspase-2 activation during general cytokinesis failure. It has been shown that PP1 can

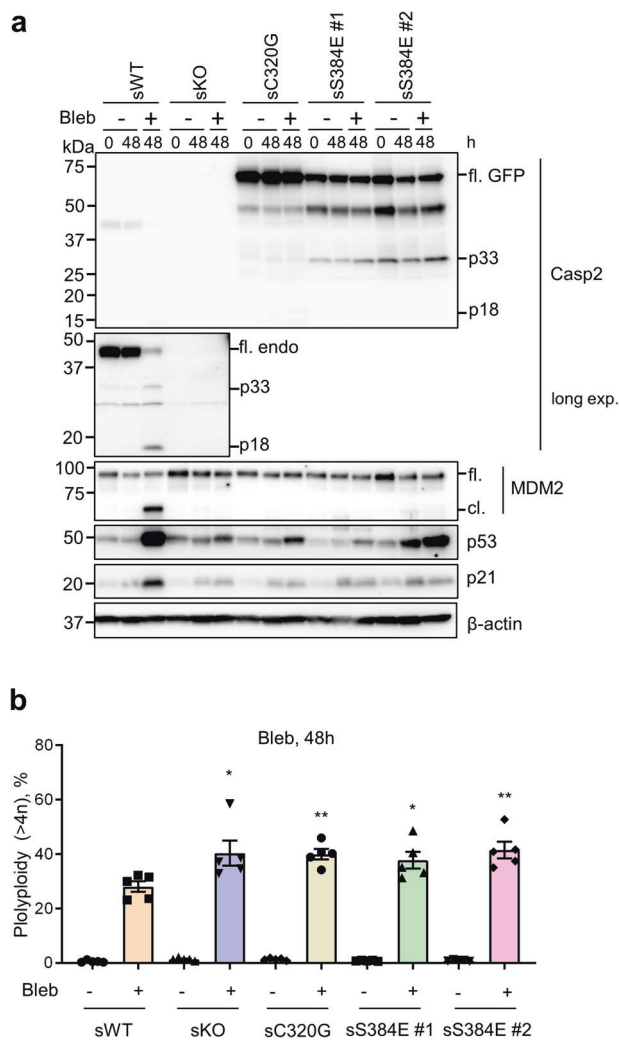


Fig. 6 Caspase-2-S384E expressing cells fail to cleave MDM2 and show increased polyploidy following cytokinesis block with blebbistatin. GFP-expressing U2OS (sWT) and U2OS-CASP2^{-/-} expressing GFP (sKO), GFP-caspase-2-C320G (sC320G) or GFP-caspase-2-S384E (sS384 #1 and #2), were treated with DMSO or 50 μM blebbistatin (Bleb) for 48 h and subjected to immunoblot and DNA content analysis. **a** Representative immunoblots (from five independent experiments) of cell lysates from treated stable cell lines. Antibodies used for immunoblotting are as indicated. β-actin was used as loading control. **b** Graph comparing percentage of cells with polyploid (>4N) DNA content following Bleb treatment. The percentage of polyploid cells (>4N) was calculated using FACS Express 6 cell cycle analysis module. Mean ± SEM; *n* = 5. **p* < 0.05; ***p* < 0.01 compared to sWT + Bleb.

dephosphorylate caspase-2 S340 residue, leading to caspase-2 activation and cell death [16, 46]. Therefore, identification of the phosphatase(s) of S384 that can trigger caspase-2 activation may provide an important mechanism to eliminate polyploid cells.

We propose that during normal mitosis caspase-2 is maintained in an inactive state by both Cdk1 and AURKB-mediated phosphorylation until anaphase onset and then

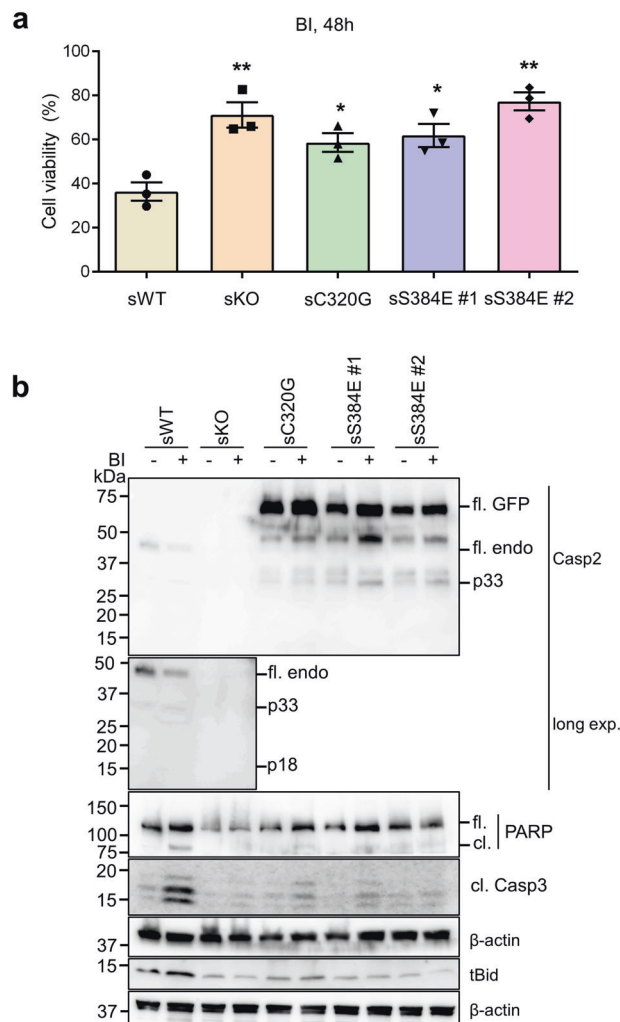


Fig. 7 Caspase-2-S384E expressing cells are resistant to aberrant mitosis-mediated cell death. GFP-expressing U2OS (sWT) and U2OS-CASP2^{-/-} expressing GFP (sKO), GFP-caspase-2-C320G (sC320G) or GFP-caspase-2-S384E (sS384 #1 and #2), were treated with DMSO or 100 nM BI2536 (BI) for 48 h and subjected to immunoblot and MTS assay. **a** Plot showing percentage of viable cells following BI treatment. Mean ± SEM; *n* = 3. **p* < 0.05; ***p* < 0.01 compared to sWT. **b** Representative immunoblots (from three independent experiments) of cell lysates from treated stable cell lines. Antibodies used for immunoblotting are as indicated. β-actin was used as loading control.

only by AURKB-mediated phosphorylation at S384 until abscission. If MC is triggered before anaphase, by failure of chromosome alignment for example, inhibition/reduction of Cdk1 and AURKB, and dephosphorylation at both S340 and S384 would activate caspase-2 to promote cell death. If instead there are irrecoverable problems specifically during cytokinesis, such as persistent lagging chromatin, then the inactivation/reduction of AURKB would enable dephosphorylation at S384 to trigger caspase-2 activation and apoptosis (Fig. 8c). Reduced AURKB levels have previously been shown to cause MC as well as premature

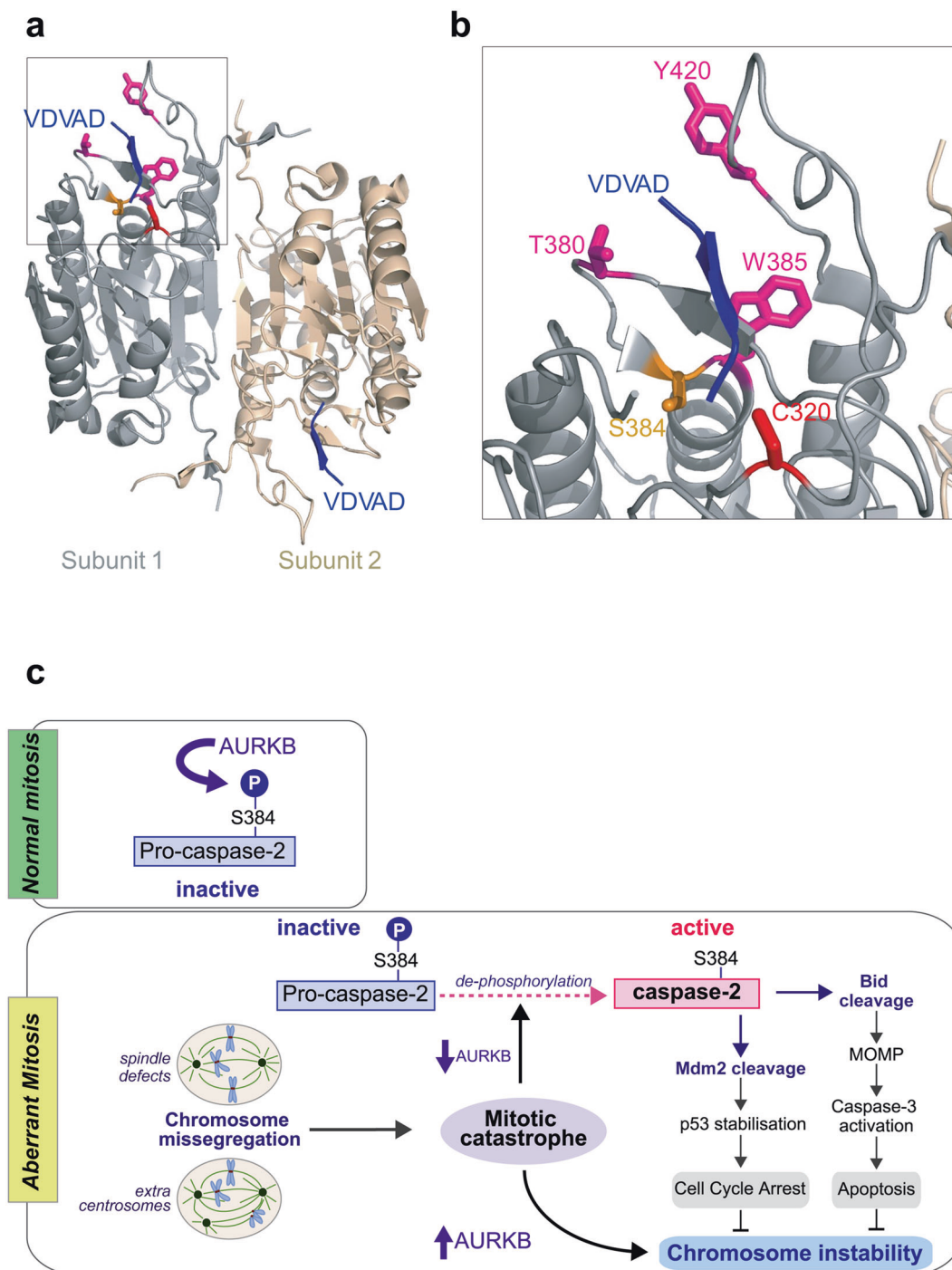


Fig. 8 Phosphorylation at S384 affects the conformation of the substrate binding pocket. **a** The crystal structure of human active caspase-2/VDVAD (PDB ID: 3R6G) [41] complex. Crystallised caspase-2 is composed of two p19/p12 heterodimers. **b** Magnified view showing position of S384 and surrounding amino acid residues important for substrate binding. T380 and Y420 and W385 are shown in magenta and interact with P5 and P4 residues [41]. S384 and C320 are shown in orange and red, respectively. **c** Schematic showing regulation of caspase-2 by AURKB-mediated phosphorylation in mitosis. During normal mitosis, S384 of caspase-2 is phosphorylated by AURKB, maintaining caspase-2 in inactive state. In response to

aberrant mitosis, such as cytokinesis failure and chromosome missegregation, reduced AURKB activity enables S384 dephosphorylation by protein phosphatase(s) (PP). This leads to caspase-2 activation that can result in two outcomes: either cleavage of MDM2, which leads to p53 stabilisation and cell cycle arrest, or cleavage of Bid, which leads to outer mitochondrial membrane permeabilization (MOMP), caspase-3 activation and apoptotic cell death. Increased AURKB levels (and reduced caspase-2 activation) are associated with chromosome instability. Therefore, these caspase-2 activation pathways limit cells becoming aneuploid or polyploid, preventing chromosome instability. Dashed lines indicate unknown mechanisms.

mitotic exit and we now demonstrate that this directly causes caspase-2 activation as a mechanism to prevent survival of cells that slip through mitosis and become polyploid.

Overexpression and enhanced kinase activity of AURKB in various tumour types are well documented and associated with therapy resistance and low survival rate in various cancers. Therefore, AURKB is considered as a potential therapeutic target in cancer [47]. Together with our data presented here, this suggests that in tumour cells, AURKB may act to maintain caspase-2 in a phosphorylated inactive state, thereby preventing apoptotic cell death that contributes to therapy resistance. We therefore propose that the phosphorylation state of caspase-2 may predict apoptosis sensitivity and treatment response in cancer.

Cytokinesis failure induced by AURKB inhibition has previously been shown to induce caspase-2 activation, MDM2 cleavage and accumulation of p53 and cell cycle arrest in both U2OS and A549 cells [14, 15]. We have also previously observed increased polyploidy in U2OS-*CASP2*^{-/-} cells, as a consequence of the accumulation of uncleaved, full-length MDM2, but still associated with increased p53 levels [14]. This suggested the presence of an MDM2-independent p53 response following aberrant mitosis, in caspase-2 deficient cells [14]. In this study, we now demonstrate that, similar to *CASP2*^{-/-} cells or C320G expression, cells stably expressing the caspase-2-S384E mutant acquire increased polyploidy associated with lack of MDM2 cleavage. Importantly, we have also observed this, using a different cytokinesis inhibitor, blebbistatin [37], that induces polyploidy. These data confirm that our results are a consequence of general cytokinesis failure and phosphorylation of caspase-2 at S384 can prevent the canonical MC response and exacerbate polyploidy.

We have previously demonstrated that caspase-2 activation and its catalytic activity are required for Bid-mediated apoptotic cell death of aneuploid cells [12]. Our findings now demonstrate that the caspase-2-S384E cell line does not show Bid cleavage and is more resistant to apoptosis caused by Plk1 inhibition. This further suggests that as well as causing accumulation of MDM2, and dampening p53-mediated cell cycle arrest, the S384E mutant prevents caspase-2-mediated apoptosis.

Taken together, our data suggest that AURKB-mediated phosphorylation at S384 of caspase-2 is a critical mechanism that regulates its activation and cellular functions. As activation of caspase-2 is required following MC to prevent survival of polyploid cells following failed mitosis and failure to repair damaged DNA [1, 3], it will be important to examine in future, whether phosphorylation of caspase-2 at S384 also prevents its tumour suppressor function.

Materials and methods

Antibodies

The following antibodies were used: anti-caspase-2 (clone 11B4; produced in-house [48] and available from Merck #MAB3507); anti-GFP (600-101-215, Rockland, Limerick, PA, USA); anti-MDM2 (OP46, EMD Millipore, Billerica, MA, USA); anti-p53 (sc-126, Santa Cruz Biotechnology, Dallas, TX, USA); anti-p21 (5567430, BD Bioscience, Franklin Lakes, NJ, USA); anti-PARP (9542, Cell Signalling Technology, Beverly, MA, USA); anti-Bid (2002, Cell Signalling Technology); anti-cleaved caspase-3 (9664, Cell Signalling Technology); anti-HA (H6908, Sigma-Aldrich, St. Louis, MO, USA); anti-GST (G7781, Sigma-Aldrich); anti-Phospho-Histone H3 (9701, Cell Signalling Technology); anti-His (ab18184, Abcam, Cambridge, MA, USA) and anti- β -actin (A1978, Sigma-Aldrich).

Cell culture and transfection

U2OS (ATCC, Manassas, VA, USA), U2OS-*CASP2*^{-/-}, U2OS-*CASP2*^{-/-}/*CASP3*^{-/-}, A549, A549-*CASP2*^{-/-} and stable cell lines were maintained in Dulbecco's Modified Eagles Medium (DMEM, Sigma-Aldrich) supplemented with 10% foetal bovine serum (JRH Biosciences, Lenexa, KS, USA), 0.2 mM L-glutamine (Sigma-Aldrich), 15 mM HEPES (Sigma-Aldrich) and 100 μ M penicillin/streptomycin (Sigma-Aldrich) in a humidified incubator at 37 °C with 10% CO₂. A549 and A549-*CASP2*^{-/-} cell lines were provided by Prof. Villunger (Medical University of Innsbruck, Austria). Where indicated, cells were treated with 100 nM BI2536 (Axon Medchem, Netherlands), 2 μ M ZM447439 (Sigma-Aldrich), 400 nM AZD1152-HQPA (Sigma-Aldrich), or 50 μ M blebbistatin (Selleck Chemicals, Houston, TX, USA) for 24 or 48 h. MEFs were maintained in DMEM (Sigma-Aldrich) supplemented with 0.2 mM L-glutamine (Sigma-Aldrich), 15 mM HEPES (Sigma-Aldrich), 10% foetal bovine serum (JRH Biosciences) and 50 μ M β -mercaptoethanol (Sigma-Aldrich), non-essential amino acid mix (Sigma-Aldrich), 100 μ M penicillin/streptomycin (Sigma-Aldrich). Cells were cultured in a humidified incubator at 37 °C with 10% CO₂. Transfection of plasmid DNA was performed using Lipofectamine 3000 (Life Technologies, Carlsbad, CA, USA) according to the manufacturer's instructions. All cell lines were tested for mycoplasma contamination.

Cloning of caspase-2 phospho site mutants and other constructs

All phosphorylation site mutants were generated by Quik-Change site-directed mutagenesis of pEGFP-mouse caspase-2

WT [49] or pcDNA-DEST47-mouse caspase-2-WT (for T161A) as PCR template. All primer sequences are outlined in Supplementary Table S1. To produce BiFC-caspase-2-S384E constructs, the caspase-2-S384E was amplified from the pEGFP-caspase-2-S384E using the following PCR primers: VN-caspase-2-S384E (forward: 5'-AAAGATCTCGC GGCGCCGAGCGGGAG-3' and reverse: 5'-AATCTAGA CGTGGGTGGGTAGCCTGG-3' with added BglIII and XbaI site); VC-caspase-2-S384E (forward: 5'-AAAGATCTCCGC GGCGCCGAGCGGGAG-3' and reverse: 5'-AACTCGAG TCGTGGGTGGGTAGCCTGG-3' with added BglIII and XhoI site). The amplified fragments were cut with BglIII/XbaI or BglIII/XhoI and subsequently subcloned in-frame into pBiFC-VN173 and pBiFC-VC155 (gifts from Lisa Bouchier-Hayes, Baylor College of Medicine, Houston, TX, USA), respectively. For bacterial expression, pGEX4T3-caspase-2-WT [27] was used as a template for PCR-based mutagenesis to introduce the S384E, S384A or C320G mutations, followed by insertion into the BamHI and XhoI sites of pGEX4T3. To obtain the small fragment of caspase-2 containing S384 (Casp₂₃₆₃₋₄₂₃-WT or -S384A), the corresponding fragments (aa 363–423) were amplified from the pGEX4T3-caspase-2-WT or -S384A using the following PCR primers: (forward: 5'-AAAGGATCCGACATGAT ATGTGGCTATGCTTG-3' and reverse: 5'-AAACTCGAG TTAGCCAGGGGCATAG CCTTC-3') and subcloned into pGEX-4T-3 using BamHI and XhoI sites. To obtain human AURKB expression plasmid construct, full-length AURKB was amplified from cDNA made from U2OS cells using the following PCR primers: (forward: 5'-AAACATATGG CCCAGAAGGAGAACTCC-3' and reverse: 5'-AAACTCG AG GGCGACAGATTGAAGGGC-3') and subcloned into pET-32a vector plasmid via NdeI and XhoI sites. Each construct was confirmed by DNA restriction enzyme digestion and DNA sequencing. All DNA restriction enzymes were purchased from NEB (Ipswich, MA, USA).

Expression and purification of recombinant proteins

The plasmids pGEX4T3-caspase-2-WT, -S384E or -S384A, pGEX4T3-Casp₂₃₆₃₋₄₂₃ (WT or S384A) or pET-32a-AURKB were transformed into *E. coli* BL21(DE3)pLysS (Promega, Madison, WI, USA). Single colonies were inoculated into 50 mL LB with 100 µg/mL ampicillin and cultured overnight at 37 °C with shaking at 180–200 rpm. The next day, 10 mL overnight culture was diluted into 400 mL LB with ampicillin (100 µg/mL) and cultured at 30 °C. To induce recombinant proteins isopropyl-β-D-thiogalactoside (IPTG, Sigma-Aldrich) was added (0.1 mM) when OD₆₀₀ was between 0.4 and 0.6 and the culture was incubated for 24 h at 16 °C. Bacterial cells were harvested by centrifugation at 5000 rpm for 10 min at 4 °C. For GST-tagged recombinant protein purification, the pellet was

resuspended in 20 mL of cold lysis buffer (25 mM HEPES, 10% sucrose, 0.1% CHAPS, 1 mM EDTA, 10 mM DTT, pH 7.4) with protease inhibitor (Sigma-Aldrich), followed by sonication (10 × 20-s pulses). Triton X-100 (Sigma-Aldrich) was then added to a final concentration 0.1% and incubated for 1 h at 4 °C with rocking, followed by centrifugation at 15 000 rpm for 30 min at 4 °C. Supernatant was transferred to a new 50 mL falcon tube and 250 µL bed volume of PBS-washed glutathione Sepharose beads (GE Healthcare Australia) was added to the lysate, followed by overnight incubation at 4 °C with rocking. Supernatant containing beads were transferred to a column in a 4 °C cold room and flow through was collected. Beads were then washed four times with 10 mL PBST (PBS, 0.1% Triton X-100). GST-tagged recombinant protein was eluted in 10 bead volumes of elution buffer (50 mM Tris, 10 mM reduced glutathione, pH 8.0) in 10 fractions. For AURKB purification, bacterial pellet was resuspended in 20 mL of cold His-lysis buffer (50 mM NaH₂PO₄, 300 mM NaCl, 10 mM imidazole, pH 8.0) with protease inhibitor (Sigma-Aldrich), followed by sonication (10 × 20-second pulses) and centrifugation at 15 000 rpm for 30 min at 4 °C. The supernatant was transferred to a new 50 mL falcon tube and 250 µL bed volume of Ni-Sepharose (GE Healthcare, Waukesha, WI, USA) was added to the lysate, followed by overnight incubation at 4 °C with rocking. Lysate containing beads were transferred to a column in the cold room and flow-through was collected. Beads were washed four times with 10 mL of wash buffer (50 mM NaH₂PO₄, 300 mM NaCl, 20 mM imidazole, pH 8.0). His-AURKB was eluted in 10 bead volumes of elution buffer (50 mM NaH₂PO₄, 300 mM NaCl, 250 mM imidazole, pH 8.0) and collected in 10 fractions. Purified proteins were checked by SDS-PAGE analysis. Pooled eluates were dialysed against 50 mM Tris-HCl, pH 7.5. Protein concentration was quantitated with BCA Protein Assay (Thermo Fisher Scientific, Rockford, IL, USA) and stored in aliquots at –80 °C until used.

Purification of caspase-2-C320G-GFP

U2OS-CASP^{2-/-} cells were seeded into 16 × 100 mm dishes at a total density of 1.5 × 10⁶ cells/dish. Cells were treated with 20 µM Z-VAD for 2 h and then transfected with 6 µg pEGFP-caspase-2-C320G for 48 h. The cells were harvested and lysed in RIPA buffer [25 mM Tris-HCl pH 7.4, 150 mM NaCl, 1% nonyl-phenoxypolyethoxyethanol (NP-40), 1% sodium deoxycholate, 0.1% sodium dodecyl sulfate (SDS)] supplemented with 1X Halt Protease and Phosphatase Inhibitor Cocktail, EDTA (Thermo Fisher Scientific), and 10 µM N-ethylmaleimide (Sigma-Aldrich). Lysed cells were sonicated, and lysates cleared by centrifugation at 13 200 rpm (4 °C). GFP-Trap (ChromoTek, Germany) was used to purify caspase-2-C320G-GFP,

according to the manufacturer's instruction. Briefly, 150 μ L GFP-Trap_MA bead suspension was equilibrated by washing with RIPA buffer three times. Cell lysates were added to the GFP-Trap_MA beads and incubated at 4 °C overnight with gentle rocking. The beads were then separated magnetically until the supernatant was clear. The supernatant was collected as 'flow-through' sample for SDS-PAGE and western blot analysis. The beads were resuspended in 1 mL ice-cold wash buffer (10 mM Tris-HCl, 150 mM NaCl, 0.5 mM EDTA, pH 7.5), briefly spun and magnetically separated. The supernatant was removed, and washing was repeated four more times with high salt wash buffer (10 mM Tris-HCl, 500 mM NaCl, 0.5 mM EDTA, pH 7.5). Bound GFP-caspase-2-C320G was eluted by adding 50 μ L elution buffer (0.2 M glycine pH 2.5), followed by incubation for 2 min and magnetic separation. The supernatant was transferred to a new tube and 5 μ L of 1 M Tris (pH 10.4) was added for neutralisation. Elution was repeated three times. A sample of each fraction (10 μ L) was subjected to SDS-PAGE analysis and Coomassie blue staining (Supplementary Fig. S1). 'Elution 1' was utilised for proteomic analysis.

Mass spectrometry and data analysis

Protein samples were resuspended in 6 M urea, 100 mM DTT and 100 mM Tris-HCl (pH 7.0) and subjected to protein digestion using FASP (filter aided sample preparation) [50]. Peptides were collected and acidified with formic acid (FA) to a 1% final concentration. Solvent was removed in a CentriVap concentrator (Labconco, Kansas, MO, USA) and peptides were resuspended in Milli-Q water containing 1% acetonitrile (ACN) and 1% FA. Samples were analyzed by nanoflow LC-MS/MS on a nanoAcquity system (Waters, Milford, MA, USA) coupled to a Q-Exactive Mass Spectrometer (Thermo Fisher Scientific) through a nanoelectrospray ion source (Thermo Fisher Scientific). Peptide mixtures were directly loaded onto a 250 mm column with 75 μ m inner diameter (nanoAcquity UPLC 1.7 μ m BEH130 C18) on a 120 min linear gradient from 1 to 35% buffer B (A: 99.9% Milli-Q water, 0.1% FA; B: 99.9% ACN, 0.1% FA) at a 400 nL/min constant flow rate. The Q-Exactive was operated in a data-dependent mode, switching automatically between one full-scan and subsequent MS/MS scans of the ten most abundant peaks. The instrument was controlled using Exactive series version 2.6 and Xcalibur 3.0. Full-scans (m/z 350–1850) were acquired with a resolution of 70 000 at 200 m/z . The ten most intense ions were sequentially isolated with a target value of 10 000 ions and an isolation width of 3 m/z and fragmented using HCD with normalized collision energy of 27 and stepped collision energy of 15%. Maximum ion accumulation times were set to 50 ms for full MS scan and 200 ms for MS/MS. Underfill

ratio was set to 2% and dynamic exclusion was enabled and set to 60 s.

The raw files were analysed using the MaxQuant [51, 52] software (version 1.5.8.3). The database search was performed using mouse protein sequences obtained from Uniprot including isoforms with strict trypsin specificity allowing up to two missed cleavages. The minimum required peptide length was set to seven amino acids. Carbamidomethylation of cysteine was set as a fixed modification while N-acetylation of proteins N-termini, oxidation of methionine, phosphorylation of S/T/Y and GlyGly on lysine were set as variable modifications. During the MaxQuant main search, precursor ion mass error tolerance was set to 4.5 ppm and fragment ions were allowed a mass deviation of 20 ppm. PSM and protein identifications were filtered using a target-decoy approach at a false discovery rate of 1%.

Protein pull-down assays

To examine the direct interaction between caspase-2 and AURKB, GST pull-down assay was carried out. GST, GST-Casp2-C320G or GST-Casp2_{363–423}-WT (1 μ g) was incubated with 500 ng His-AURKB in 300 μ L incubation buffer (50 mM Tris-HCl, 10 mM MgCl₂, 0.1 mM EDTA, 0.01% Brij 35, pH 7.5) for 2 h with rotation, at room temperature. This was followed by addition of 10 μ L glutathione-agarose beads (GE Healthcare) and further incubation for 1 h with rotation at room temperature. After four washes with incubation buffer, beads were heated in 2x Laemmli buffer for 5 min at 95 °C. Samples and 10% inputs were then subjected to SDS-PAGE and IB.

To examine the interaction between caspase-2 and AURKB in cells, U2OS-CASP2^{-/-} cells stably expressing GFP (sKO) or GFP-Casp2-C320G (sC320G) were used. For mitotic cells, sC320G cells were synchronized in G1/S by treatment with 2 mM thymidine (Sigma-Aldrich) for 24 h. After washing with PBS, the cells were released into complete media for 3 h, followed by treatment with 100 ng/mL nocodazole (Sigma-Aldrich) for 14 h. Mitotic cells were collected by gentle shaking of the culture flasks. For AURKB inhibition, sC320G cells were treated with AURKB inhibitor, ZM447439 (ZM, 2 μ M) for 24 h. The cells were lysed in NP-40 lysis buffer (150 mM NaCl, 1% NP-40, 50 mM Tris-HCl, pH 8.0) containing 1X Halt Protease and Phosphatase Inhibitor Cocktail, EDTA (Thermo Fisher Scientific). The cell lysates were then incubated with GFP-Trap for 2 h at 4 °C with gentle rocking, followed by four washes in lysis buffer. Bound GFP or GFP-caspase-2-C320G was eluted by adding 25 μ L elution buffer (0.2 M glycine, pH 2.5) as described above. The pooled elutes were neutralised by adding 5 μ L of 1 M Tris (pH 10.4) and subjected to IB. Forty micrograms of the cell lysates were used as input.

Caspase-2 dimerisation assay

BiFC analysis for caspase-2 activation was performed as described previously with minor modifications [53]. Briefly, U2OS-*CASP2*^{-/-} cells were seeded onto 13 mm glass coverslips (Thermo Fisher Scientific) and incubated overnight at 37 °C in a 10% CO₂ incubator. The next day, the medium was replaced with complete culture medium containing 20 μM Z-VAD-FMK (Sigma-Aldrich) 2 h prior to transfection. The cells were then co-transfected with 150 ng of the pBiFC-HA-Casp2(S384E)-VC155 and pBiFC-HA-Casp2(S384E)-VN173 (mouse caspase-2) for BiFC and 10 ng of pDsRed-Mito (Clontech, Takara Bio USA, Mountain View, CA, USA) as a transfection reporter plasmid, using Fugene HD reagent (Promega). pBiFC-HA-CASP2 (C320A)-VC155 and pBiFC-HA-CASP2 (C320A)-VN173 (human caspase-2) were used as system control [12]. After incubation for 2 h, DMSO (control) or 100 nM BI2536 (Sigma-Aldrich) was added to the medium and cells were incubated a further 24 h at 37 °C in a 10% CO₂ incubator. Cells were then fixed with 4% paraformaldehyde (Sigma-Aldrich) in PBS and BiFC imaged by confocal microscopy using a Leica TCS SP8 (Leica, Wetzlar, Germany). At least 100 cells were counted in five different areas in two independent experiments to quantify BiFC-positive cells.

Caspase activity assay

Caspase activity assay using VDVAD-AFC was carried out as previously described [54]. The protein concentration of cell lysates were quantified by BCA Protein Assay (Thermo Fisher Scientific) according to the manufacturer's instructions and 50 μg of cell lysate was mixed with caspase-2 activity buffer (0.1 M MES [2-(N-morpholino) ethanesulfonic acid], 10% sucrose, 0.1% CHAPS, 0.5 mM EDTA, pH 6.5) to a total volume of 50 μL. VDVAD-AFC (100 μM, Sigma-Aldrich) diluted in 50 μL of caspase-2 activity buffer was added to each sample to give a final volume of 100 μL per well. Fluorescence was measured in 10 min intervals for 170 min at 37 °C, with excitation = 400 nm, emission = 505 nm, to quantitate caspase-2 activity. For GST recombinant proteins, 20 μL of crude *E. coli* extract was used.

Generation of KO cell lines

U2OS-Caspase-2 KO (U2OS-*CASP2*^{-/-}) cell line was generated using CRISPR/Cas9 technology as described previously [55]. pSpCas9(BB)-2A-Puro (PX459) was a gift from Feng Zhang (Addgene plasmid #48139; <http://n2t.net/addgene:48139>; RRID:Addgene_48139) [55]. The sgRNA oligos (sense: 5'-caccgacaaaatgttctcatcc-3'; anti-sense: 5'-aacggatgaagaacattttggtc-3') were annealed,

phosphorylated by T4 PNK (NEB) and inserted into BbsI (NEB) site of pSpCas9(BB)-2A-Puro (PX459) vector plasmid. To establish the KO cell line, 2 × 10⁵ U2OS cells were transfected with 2 μg of sgRNA and Cas9 expressing plasmid using Fugene HD reagent (Promega) according to the manufacturer's instruction. pSpCas9(BB)-2A-GFP and pSpCas9(BB)-2A-Puro (PX459) were also transfected as a negative control and positive control, respectively, for puromycin treatment. The cells were treated with 1 μg/mL puromycin for 2 days until all negative control cells had died. The surviving cells were harvested and re-seeded into three 10 cm dishes at 1 × 10³ cells/dish for isolating single colonies. To screen candidate single clones, the sgRNA-targeted area was amplified by PCR (forward: 5'-tggtggaagccaactgttgaacc-3'; reverse: 5'-tctcagaaaggaaggcaagacacg-3') and the PCR products were analysed by heteroduplex analysis in PAGE gel as described previously [56] and DNA sequencing. DNA sequencing results were used for insertion and deletion analysis in the targeted gene area by TIDE software (<http://tide.nki.nl>). Loss of caspase-2 expression in the candidate clones was validated by western blot.

The U2OS-*CASP2*^{-/-}/*CASP3*^{-/-} cell line was generated using one of the U2OS-*CASP2*^{-/-} clones, as above. The sgRNA oligos (sense: 5'-caccgattacataaacccatctc-3'; anti-sense: 5'-aacagagatgggtttatgtataatc-3') used for cloning into pSpCas9(BB)-2A-Puro (PX459) vector plasmid. The sgRNA-targeted area was amplified by PCR (forward: 5'-gcctcactcaggtcatttt-3'; reverse: 5'-tgtaggtcctgcccaatctc-3') for genetic analyses. Western blot was performed to validate loss of caspase-3 expression.

Immunofluorescence microscopy

U2OS cells stably expressing GFP (sWT) and U2OS-*CASP2*^{-/-} cells stably expressing GFP (sKO), GFP-Casp2-C320G (sC320G) or GFP-Casp2-S384E (sS384E) were generated using pEGFP mock, pEGFP-Casp2-C320G or pEGFP-Casp2-S384E plasmids, respectively. For immunofluorescence microscopy, cells were grown on coverslips overnight and fixed in 4% paraformaldehyde in PBS for 10 min at room temperature. The fixed cells were washed in TBS, permeabilized and blocked with blocking solution (3% BSA, 0.1% Triton X-100 in TBS) for 1 h at RT. Samples were incubated overnight with goat anti-GFP antibody at 1:500 dilution in blocking solution. After three washes in TBS/0.1% Triton X-100, coverslips were incubated for 60 min with anti-goat IgG-Alexa568 (Molecular Probes, Thermo Fisher Scientific) diluted in blocking solution. Samples were washed another three times (as above) and coverslips were mounted onto glass slides with mounting medium containing DAPI (Life Technologies). Images were taken with a Zeiss LSM-800 confocal microscope (Zeiss, Germany).

Immunoblotting

Protein samples were denatured for 5 min at 95 °C and run on Mini or Midi TGX Stain-Free Precast Gels (Bio-Rad, Hercules, CA, USA) at 110 V for 60 min (mini gels) or at 140 V for 60 min (midi gels) in SDS-PAGE running buffer (250 mM Tris, 192 mM glycine, 0.06% SDS). The proteins were transferred onto PVDF membrane (Bio-Rad) using a Trans-Blot Turbo (Bio-Rad), according to the manufacturer's instructions, in transfer buffer (25 mM Tris, 192 mM glycine, 20% methanol, 0.05% SDS). Membranes were then incubated in blocking buffer [TBST (20 mM Tris, 150 mM NaCl pH 7.4, 0.05% Tween- 20)], 5% (w/v) skim milk powder (Diploma) for 1 h at RT, followed by incubation with primary antibody overnight at 4 °C. Following membrane washing in TBST for 4 × 15 min, membranes were incubated with horse radish peroxidase (HRP)-conjugated secondary antibodies for 1 h at RT. Membranes were then washed in TBST for 4 × 10 min, followed by development with Enhanced Chemiluminescence reagent (Pierce Chemical Co., Rockford, IL, USA). Fuji LAS4000 System (GE Healthcare) or ChemiDoc MP (Bio-Rad) were used for detecting luminescence signals.

DNA content analysis

DNA content analysis was performed as described previously [14]. Briefly, harvested cells were washed with ice-cold 1X PBS followed by fixation in 70% ice-cold ethanol in PBS. Fixed cells were incubated overnight at -20 °C. Cells were centrifuged at 1200 rpm for 5 min then resuspended in 2 mL 1X PBS for rehydration. Cells were centrifuged a second time and pellets resuspended in 1 mL 0.25% Triton X-100 (Sigma-Aldrich) in PBS. Following a final centrifugation step, cell pellets were resuspended in 400 µL staining solution [25 µg/mL propidium iodide (Sigma-Aldrich), 40 µg/mL RNase A (Sigma-Aldrich)] and incubated for 2 h at room temperature in the dark. The stained cells were stored at 4 °C until flow cytometric analysis on a Fortessa (BD Biosciences). For data in Fig. 5b, Fig. 6b and Supplementary Fig. S7b, approximately 25 000 cells were analysed and the percentage of polyploid cells (>4*N*) was calculated using FACS Express 6 cell cycle analysis module.

Cell death and viability assay

For cell death assays, WT or *Casp2*^{-/-} immortalised MEFs [11] were co-transfected with GFP-caspase-2-WT, -S384A or -S384E and β-gal expressing plasmids. After 24 h, cells were fixed and incubated with an X-gal containing solution. Blue (transfected) cells were counted for apoptotic

morphology with a stereomicroscope (Nikon, Tokyo, Japan). At least 300–400 cells were counted in at least 20 different fields for each construct. Cell viability was determined using the MTS assay. Briefly, cells were seeded in triplicate at 5000 cells per well in 50 µL complete culture media in a 96-well microplate (BD Biosciences) and cultured overnight at 37 °C with 10% CO₂. Culture medium (50 µL) containing DMSO or 200 nM BI2536 was added, followed by 48 h incubation at 37 °C with 10% CO₂. Twenty-five microliters of MTS/PMS (96:4) reagent was added to each well. The plate was then incubated in a humidified, 10% CO₂ incubator for 4 h, followed by reading absorbance at 490 nm using a FLUOstar Omega (BMG Labtech, Ortenberg, Germany). Control wells (no cells), were used to detect the cell-free background absorbance.

In vitro kinase assay

In vitro kinase assays were performed as described previously [57]. Proteins were incubated with 38 ng of Aurora B kinase (PV6130 Life Technologies) or 125 ng of Aurora A kinase (PV3612 Life Technologies), 0.1 mM ATP (Sigma-Aldrich), 5 µCi of [γ -³²P]-ATP (3000 Ci/mmol, 10 mCi/mL) (PerkinElmer, Waltham, MA, USA) in kinase buffer (20 mM HEPES [pH 7.5], 2 mM MgCl₂, 1 mM DTT) in a final reaction volume of 12.5 µL. After 30 min incubation at 30 °C with constant agitation, 12.5 µL of 2x Laemmli sample buffer were added to stop the reaction. Samples were heated for 10 min at 90 °C and loaded on a 4–20% Tris-Glycine Precast Gel (Thermo Fisher Scientific). Protein was transferred onto a nitrocellulose membrane using the iBlot Dry Blotting System. Membranes were then exposed to Kodak BioMax XAR Films (Sigma-Aldrich) at -80 °C. The membranes were then blocked and subjected to IB for GST.

Statistical analysis

The data are presented as mean ± standard error of the mean (SEM) and were considered statistically significant when $p < 0.05$. A two-sided Student's *t* test was used to analyse differences between data groups unless otherwise stated. All statistical analyses were conducted using GraphPad Prism, Version 6.05 (GraphPad, supplied by University of South Australia; GraphPad Inc., CA, USA).

Data availability

The mass spectrometry proteomics data have been deposited to the ProteomeXchange Consortium via the PRIDE [58] partner repository with the dataset identifier PXD017866, and are available via ProteomeXchange.

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Author contributions YL designed and performed experiments, analysed data and wrote the paper; DDB helped with the generation of mutants and performed cell biology experiments; JJS and AIW carried out MS analysis; LC and PPD'A performed in vitro phosphorylation and IB experiments; JMM generated the caspase-2 structure modelling; LD and SK designed and supervised the study, analyzed data, acquired funding and wrote the paper. All authors discussed the results and commented on the manuscript.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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