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# Trophic niche segregation allows range-extending coral reef fishes to co-exist with temperate species under climate change 3

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#### 20 Abstract

Changing climate is forcing many terrestrial and marine species to extend their ranges poleward to stay within the bounds of their thermal tolerances. However, when such species enter higherlatitude ecosystems, they engage in novel interactions with local species, such as altered predator-prey dynamics and competition for food. Here we evaluate the trophic overlap between range-extending and local fish species along the east coast of temperate Australia, a hotspot for ocean warming and species range extensions. Stable isotope ratios ( $\delta^{15}$ N and  $\delta^{13}$ C) of muscle tissue and stomach content analysis were used to quantify overlap of trophic niche space between vagrant tropical and local temperate fish communities along a 730 km (6°) latitudinal gradient. Our study shows that in recipient temperate ecosystems, sympatric tropical and temperate species do not overlap significantly in their diet – even though they forage on broadly similar prey groups – and are therefore unlikely to compete for trophic niche space. The tropical and temperate species we studied, which are commonly found in shallow-water coastal environments, exhibited moderately-broad niche breadths and local-scale dietary 

plasticity, indicating trophic generalism. We posit that because these species are generalists,
 they can co-exist under current climate change, facilitating the existence of novel community
 structures.

37

#### 38 KEYWORDS

#### 39 diet, niche use, ocean warming, range shifts, stable isotopes

40

#### 41 Introduction

42 To persevere in a changing environment, species must acclimate (Donelson, Munday, McCormick, & Pitcher, 2011; Stillman, 2003), adapt, or relocate (Booth, Bond, & Macreadie, 43 2011; Pecl et al., 2017; Perry, Low, Ellis, & Reynolds, 2005). The balance between persistence 44 45 and extinction is jeopardized by climate change, causing shifts in range edges (Cheung et al., 2012; Poloczanska et al., 2013; Sinervo et al., 2010), greatly impacting the structure and 46 function of ecosystems (Dawson, Jackson, House, Prentice, & Mace, 2011). Marine species 47 shift their range edges under global warming at an average rate of 19 km year<sup>-1</sup> (Sorte, 48 Williams, & Carlton, 2010), an order of magnitude faster than the average terrestrial rate of 0.6 49 km year<sup>-1</sup> (Maggini et al., 2011; Poloczanska et al., 2013; Sorte et al., 2010). Range shifts may 50 be facilitated by the strengthening of boundary currents, such as Australia's East Australian 51 52 Current (EAC), as well as a drought-related reduction in freshwater flow (Booth et al., 2011; 53 Johnson et al., 2011; Last et al., 2011). To date, 75% of marine range shifts occur in a poleward direction (Sorte et al., 2010). 54

55 Whilst there are ample insights into the mechanisms and implications of species 56 invasions within terrestrial systems (Chen, Hill, Ohlemüller, Roy, & Thomas, 2011; Estrada, 57 Morales-Castilla, Caplat, & Early, 2016; Peers, Wehtje, Thornton, & Murray, 2014), different 58 traits and processes (e.g. ocean currents) can regulate invasions in marine ecosystems, and 59 therefore the same principles may not be relevant to both biomes (Burrows et al., 2011; Dawson et al., 2011; McKnight, García-Berthou, Srean, & Rius, 2017). Even among marine 60 61 ecosystems, range edge movement cannot always be generalized, in part due to the largely indiscriminate nature of water movement, e.g. non-linear eddies (Bates et al., 2014; McKnight 62 et al., 2017). When range shifts occur near islands and continent edges, where habitat is limited 63 (Cahill et al., 2012; Wernberg et al., 2016), or where water movement is caused by non-linear 64 65 currents and eddies (Garciá Molinos, Burrows, & Poloczanska, 2017), range shifts become even more difficult to predict, and sometimes recede rather than extend. 66

67 Many studies have focussed on range-extensions of warm-affiliated species to higher latitudes, but we still lack a detailed understanding of how recipient ecosystems and 68 communities are affected by these native invasions (Bates et al., 2014; Feary et al., 2014). 69 Several scenarios exist in which range-extending species might exert positive effects or 70 71 ecosystem services on recipient communities. For example, range-extending species can serve as functional substitutes for extinct local species and provide food and habitat to rare local 72 species, which may be vital in achieving future conservation goals (Dudgeon & Smith, 2006; 73 74 Gozlan, 2008; Schlaepfer, Sax, & Olden, 2011). Research on the range extensions of boreal 75 arctic generalist fish showed an increase in connectivity between pelagic and benthic communities and a subsequent reduction in modularity (Kortsch, Primicerio, Fossheim, 76 Dolgov, & Aschan, 2015). Additionally, range-extending species may be more adaptable than 77 78 native species, and as the climates continue to change, outlast local species and ultimately evolve into new endemic species (Schlaepfer et al., 2011). 79

Negative effects of range-extensions include novel predators causing local extinction of native species (Bampfylde & Lewis, 2007; Wiens, 2016), hybridization altering the integrity of local species (Gozlan, 2008), threating ecosystem functions, such as the overgrazing of macrophytes (Gallardo, Clavero, Sanchez, & Vila, 2016) and the creation of urchin barrens

84 (Ling et al., 2015), and carrying infectious diseases that can harm native species (Schlaepfer et al., 2011). If a non-native species has high rates of consumption and growth, they have the 85 potential to damage local communities by outcompeting, and potentially replacing, native 86 87 consumer populations (McKnight et al., 2017; Schlaepfer et al., 2011). This replacement could intensify the local extinction rates of native species by increasing their susceptibility to 88 anthropogenic stressors, having both ecological and commercial repercussions (McKnight et 89 90 al., 2017). On the other hand, tropical species in temperate environments may experience 91 greater predation by local predators, as they are a novel species within the community (Almany 92 & Webster, 2004). The exact effect of range-extending species on recipient ecosystems, 93 however, depends on a variety of factors including a species' geographic range, abundance, 94 and per capita effects. Species are thought to have a high per capita effect if they are 95 functionally distinctive in their food web position, their response to environmental factors, their acquisition of resources, or their effects on disturbance regimes (Chapin et al., 1997; Vitousek, 96 1990; Wardle, Bardgett, Callaway, & Van der Putten, 2011). In general, it is difficult to 97 98 estimate the per capita impact that a species will have until the invasion begins to happen 99 (Strayer, 2012).

100 Here we test how range-extending tropical reef fish, thought to be the most at-risk group under future climate change (Comte & Olden, 2017), create novel trophic niches in temperate 101 102 coastal ecosystems. Novel interactions such as changes in predator-prey relationships and 103 trophic competition are likely to arise between newly sympatric groups (Alexander, Diez, Hart, 104 & Levine, 2016; Pörtner & Farrell, 2008; Wernberg et al., 2011). Using stable isotopes and 105 stomach contents, we determine how trophic niche partitioning might facilitate co-existence 106 among coastal vagrant tropical and local temperate species by exploring several scenarios of 107 niche modification: segregation, overlap, displacement, and expansion or contraction (Fig. 1). 108 Our study focuses on the temperate east coast of Australia, where coastal waters are warming at a rate 3-4 times the global average (Holbrook & Bindoff, 1997; Johnson et al., 2011;
Ridgway, 2007).

**Fig. 1**: Conceptual diagram of trophic niche space of co-existing species based on existing theory. Niches are displayed as convex hull polygons in isotope ( $\delta^{15}$ N and  $\delta^{13}$ C) space. Possible arrangements include niche segregation (no overlap) (**a**), niche overlap (**b**), niche displacement (**c**), and niche expansion and/or contraction (**d**). The different coloured niches represent different affinities (tropical vs temperate) or species. In the case of niche displacement, the dashed blue line represents where the blue niche was before it was displaced.

117

#### 118 Materials and Methods

119 *Study species* 

120 The fish species studied were selected based on their ubiquity and ease of capture at known 121 locations, and include tropical pomacentrid damselfishes (Abudefduf bengalensis\*, A. sexfasciatus<sup>\*</sup>, A. vaigiensis<sup>\*</sup>, Chrysiptera brownriggii, Dascyllus aruanus, Pomacentrus 122 bankanensis, P. coelestis, P. moluccensis), tropical butterflyfishes (Chaetodon auriga\*, C. 123 citrinellus, C. flavirostris, C. kleinii, C. rainfordi, C. tricinctus, and C. vagabundus), two 124 temperate sea chubs (Atypichthys strigatus\* and Microcanthus strigatus\*), a temperate 125 pomacentrid damselfish (Parma microlepis\*), and the temperate Port Jackson glassfish 126 (Ambassis jacksoniensis) (See Table S1 for fish lengths, weights, sample sizes, and in situ 127 128 densities). Some of these common tropical species, namely A. sexfasciatus and A. vaigiensis, are often observed to school with the temperate A. strigatus and M. strigatus (Figueira, Curley, 129 & Booth, 2019; Smith, Fox, Booth, & Donelson, 2018). All above species were included in the 130 131 stable isotope analysis, but due to their small sample sizes in some regions, only species indicated with '\*' could be used for stomach contents analysis, and consequently for 132 calculation of diet breadth and niche breadth. 133

The tropical species are commonly found in warm Indo-Pacific waters, including the 134 Great Barrier Reef (Getlekha et al., 2016; Masuda, Amaoka, Araga, Uyeno, & Yoshino, 1984). 135 The temperate species *M. strigatus* is found from Japan down into tropical and temperate 136 Australia and east towards the Hawaiian Islands, while temperate species A. jacksoniensis, A. 137 strigatus, and P. microlepis are found in south-eastern Australia only. The tropical species used 138 in this study that were observed at Sydney and further south (Fig. 2; A. bengalensis, A. 139 140 sexfasciatus, A. vaigiensis, C. auriga, C. citrinellus, C. flavirostris, C. vagabundus, P. bankanensis, and P. coelestis) are considered vagrants as they only occur at these locations on 141 142 a seasonal-basis. Their presence drops quickly as water temperature decreases throughout the seasons, and their presences are also much lower at the cold-water Merimbula site than the 143 144 warmer Sydney site (Table S2). The remainder of the tropical species were not observed as far 145 south as Sydney. Furthermore, the peak temperatures (below which abundances start to decline) of all the selected tropical species (Booth, Beretta, Brown, & Figueira, 2018) are lower 146 than the mean winter temperatures at the two study sites (Table S2). All species studied are 147 omnivorous (Lieske & Myers, 1994; Myers, 1991; Randall, 1985). The East Australian Current 148 149 (EAC) transports larvae of coral reef fishes from tropical regions to southern sub-tropical and temperate regions (Booth, Figueira, Gregson, Brown, & Beretta, 2007; Smith, Fox, Donelson, 150 151 Head, & Booth, 2016).

152

#### 153 *Field collection*

Field sampling was conducted in February and March of 2017 and 2018 in four different regions (Fig. 2; true coral reefs, and North, Middle, and South regions of range extensions). Sample locations include South West Rocks (two sites) and Port Stephens (two sites) ("North"); Sydney (two sites) ("Middle"); Bass Point (one site), Narooma (one site), and Merimbula (two sites) ("South"); and One Tree Island (two sites) and Lord Howe Island (three 159 sites) ("coral reef") (Fig. 2). One Tree Island and Lord Howe Island were sampled in 2017 only, and only tropical species were present and captured there. These coral reef locations were 160 chosen based on their latitude and geographical spacing, as well as *a priori* knowledge that our 161 species of interest reside there (Booth et al., 2007). Sample sizes of each species at each site 162 are shown in Table S1 and species' seasonal ranges are shown in Table S2. Fish were caught 163 using hand nets at depths of 1-3 m using ethanol and clove oil, to anesthetize the fish. The fish 164 165 were immediately killed using the *iki jime* method, and placed on ice before being frozen. All samples were frozen within four hours of collection and remained frozen at -30 °C before 166 167 processing.

168

Fig. 2: Sample locations along the southeast coast of Australia, including South West Rocks 169 170 (two sites; 30°52'34"S, 153°04'02"E and 30°53'00"S, 153°02'17"E) and Port Stephens (two sites; 32°42'56"S, 152°10'58"E and 32°44'55"S, 152°10'19"E) (included in region "North", red 171 marker); Sydney (two sites; 33°42'07"S, 151°18'28"E and 33°47'44"S, 151°17'25"E) (included 172 in region "Middle", purple marker); Bass Point (34°35'54"S, 150°53'18"E), Narooma 173 (36°12'54"S, 150°07'51"E), and Merimbula (two sites; 36°44'13"S, 149°58'58"E and 174 36°53'40"S, 149°55'25"E) (included in region "South", blue marker); and One Tree Island 175 (23°30'30"S, 152°05'30"E) and Lord Howe Island (31°33'15"S, 159°05'06"E) ("Coral reef", 176 177 black marker). Tropical species were collected from study sites spanning 1,500 km (13° 178 latitude), while temperate species were collected along 730 km (6° latitude) of coastline. Temperate species were not present at One Tree Island or Lord Howe Island. 179

180

181 Sample preparation

182 The scales and skin were removed from the right lateral side of each fish. After a thorough 183 cleaning of the scalpel blade, a pea-sized piece of white muscle tissue, free of bone, organs, and scales, was removed and placed into a labelled tube, and immediately re-frozen. Stomachs
and guts were removed and frozen separately for subsequent stomach content analysis, with
the exception of stomachs collected in 2017, which were stored in 70% ethanol.

187 The fish tissue was freeze-dried for at least 36 hours to remove moisture. A small metal 188 ball was then placed into each individual tube, and the tissue was ground into a fine powder 189 using a ball mill. This ensured that the sample was homogeneous, and therefore representative 190 of the organism's true  $\delta^{15}$ N and  $\delta^{13}$ C content (Eurich, Matley, Baker, McCormick, & Jones, 191 2019). Powdered samples were then weighed into tin capsules and analysed for stable isotope 192 content using a Nu Instruments Nu Horizon Continuous Flow IRMA (CF-IRMA).

193

194 Stable Isotope Analysis (SIA)

195 Stable isotope analysis (SIA) can be used to estimate an individual's position in trophic space 196 (Layman, Arrington, Montana, & Post, 2007), track migration routes (Hobson, 1999), and assess shifts in diet (Phillips & Eldridge, 2006). These questions can be answered by analysing 197 198 the type and abundance of prey and associated carbon and nitrogen stable isotopes that have been incorporated into muscle tissue over the past few weeks or months (Fitzgerald, 199 200 Winemiller, Perez, & Sousa, 2017; Fry, 2006; Newsome, del Rio, Bearhop, & Phillips, 2007). In this way, the accumulation and transformation of organic matter can be traced within a 201 community over time (Fanelli, Azzurro, Bariche, Cartes, & Maynou, 2015). SIA utilises  $\delta^{15}$ N, 202 the ratio of <sup>15</sup>N:<sup>14</sup>N, as an indicator of trophic position and  $\delta^{13}$ C, the ratio of <sup>13</sup>C:<sup>12</sup>C, as an 203 indicator of what the individual's source of prey has been over the past few weeks or months 204 (Cabana & Rasmussen, 1994; Campbell, Wandera, Thacker, Dixon, & Hecky, 2005; Newsome 205 et al., 2007).  $\delta^{15}$ N increases ~3.4‰ on average (Post, 2002) at each trophic transfer due to 206 nitrogen fractionation, with different prey items resulting in different fish tissue signatures 207 208 (Checkley & Entzeroth, 1985; DeNiro & Epstein, 1978; McCutchan, Lewis, Kendall, & 209 McGrath, 2003).  $\delta^{13}$ C varies little between trophic levels (0-1‰), as carbon fractionation is primarily influenced by prey type (DeNiro & Epstein, 1978; Newsome et al., 2007; Post, 2002). 210 Here, stable isotope analysis was used to display the trophic niche of 1) temperature 211 affinity groups, and 2) individual species in isotopic ( $\delta^{15}$ N and  $\delta^{13}$ C) space. At the affinity level, 212 species were grouped into 'tropical' or 'temperate' affinities. Trophic niche areas of the two 213 affinities and of individual species were calculated for each region (North, Middle, and South) 214 215 using convex hulls in the rKIN package available in R (Albeke, 2017). Individual fish were plotted in isotopic space (i.e. isospace) and a convex hull (100% of data points used) was 216 217 created to encapsulate the data points for each species or affinity, depending on the analysis (Layman, C. A. et al., 2007; Syväranta, Lensu, Marjomäki, Oksanen, & Jones, 2013). In 218 219 addition to convex hulls, standard Bayesian ellipses (95% confidence interval of the bivariate

means) were used to calculate the niche area (SEAc) of individual species using the *R* package

SIBER (Jackson, Inger, Parnell, & Bearhop, 2011; Jackson & Parnell, 2019; Jackson et al.,

2012). Standard ellipses can correct for small sample size, such is the case for some of the

species used in this study, making them a more suitable tool for individual species niche area

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analysis than convex hulls alone.

In addition to niche area, we also calculated the degree of overlap between niches. For 225 overlap between affinities, percentages were calculated using convex hulls in the program rKIN 226 227 (Albeke, 2017), with overlap expressed as the percent that each species area is being 228 overlapped. Affinity overlap percentages are from the perspective of the temperate species, as it is the temperate recipient community that is being invaded and is under investigation. For 229 individual species, niche overlap was calculated using ellipses in the *R* package nicheROVER 230 231 (Lysy, Stasko, & Swanson, 2014). Ten random elliptical projections were created for each species at each region based on posterior distribution. Overlap was then determined based on 232 the probability of one species falling into the niche of another (Swanson et al., 2015). Because 233

niches are not symmetrical, there are two possible values for each species pair, one for the 234 percent probability that hypothetical group "X" is being overlapped by group "Y", and the 235 other for the percent probability that group "Y" is being overlapped by group "X". For example, 236 if a large ellipse was directly on top of a smaller ellipse in isospace, one could calculate the 237 probability of overlap from the perspective of the large ellipse, e.g., 15% probability of overlap, 238 whereas the overlap probability from the perspective of the small ellipse would be 100%. Niche 239 240 areas which overlap by > 60% are thought to be sharing a significant amount of niche space (Guzzo, Haffner, Legler, Rush, & Fisk, 2013; Schoener, 1968; Wallace, 1981). The niche 241 range, or the maximum/minimum difference of both  $\delta^{15}N$  and  $\delta^{13}C$  values, was also calculated 242 for each species, and then averaged across affinity for each region, to identify whether the fish 243 are occupying a similar isotopic breadth across the regions. 244

245

#### 246 Stomach Content Analysis

Stomach content analysis is used to provide direct evidence of prey items consumed in the 247 hours leading up to the fish's collection (Rybczynski, Walters, Fritz, & Johnson, 2008). Stable 248 isotope signatures provide a measure of where in  $\delta^{15}N$  and  $\delta^{13}C$  isospace fishes are situated, 249 but stomach content analysis reveals on what taxa, and in what quantities, tropical and 250 temperate fish are feeding (Svensson et al., 2017). Only stomachs collected in 2018 were used 251 252 for stomach content analysis, due to disintegration of the 2017 stomachs that were stored in 253 ethanol. As such, some species did not have a large enough regional sample size (e.g. A. *bengalensis* in the Middle and *C. auriga* in the North) to be used in diet analyses (stomach 254 contents, Levins' index of niche breadth, and Pianka's index of diet overlap) in all regions. 255 256 Only species with at least 3 replicate stomachs were included for the diet analyses. However, all species were still used in the isotopic niche analyses, as stable isotope data from both 2017 257 and 2018 were included. 258

The stomachs of all fish were emptied of their contents. Contents that were too digested 259 to be distinguished, as well as gastric juices, were excluded from analysis. Food items were 260 divided into broad categories, including algae, seagrass, crustaceans, worms, gastropods, eggs, 261 zooplankton, detritus, hydroids, fish larvae, parasites, nudibranchs, and terrestrial insects and 262 seeds (Layman, Quattrochi, Peyer, & Allgeier, 2007). For example, both fleshy green algae 263 and branching red algae were grouped into the broad category 'algae'. If a stomach was 264 265 composed of 10% zooplankton, 10% algae, 30% gastric juice, and 50% unknown matter, it would be classified as containing 50% zooplankton and 50% algae to standardise diets solely 266 267 based on identifiable prey items consumed. The prey categories in the stomachs of each species were averaged for each region to calculate diet overlap among species and across regions using 268 Pianka's index (Pianka, 1973). Pianka's diet overlap is calculated using the following equation: 269 270

271 (Equation 1)

272 
$$O_{jk} = \frac{\sum_{i}^{n}}{\sqrt{1-x^{2}}}$$

$$O_{jk} = \frac{\sum_{i}^{n} p_{ij} p_{ik}}{\sqrt{\sum_{i}^{n} p_{ij}^2 \sum_{i}^{n} p_{ik}^2}}$$

273 where:274

275  $O_{ik}$  = Pianka's index of diet overlap between species *j* and species *k* 

276  $p_{ij}$  = proportion resource *i* is of the total resources used by species *j* 

277  $p_{ik}$  = proportion resource *i* is of the total resources used by species *k* 

278 n = total number of resource states

279

Pianka's values range from 0-1, with 0 indicating total segregation, and 1 indicating total diet
overlap (Vieira & Port, 2007). Similar to isotope niche overlap, Pianka's values > 0.6 are
considered to represent a significant diet overlap (Wathne, Haug, & Lydersen, 2000; Zaret &

Rand, 1971). Statistical analyses were performed by grouping Pianka's values into either intraaffinity overlap (tropical-tropical or temperate-temperate), or inter-affinity overlap (tropicaltemperate). ANOVA was then performed to test if there was a significant difference in overlap
between each of these three groups. As such, ANOVA only had region (North, Middle, South)
and affinity group (tropical-tropical, temperate-temperate, and tropical-temperate) as factors.

Niche breadth represents the range of resources that a species uses (Slatyer, Hirst, & Sexton, 2013) and can be used to determine feeding strategies, and separate species into specialists or generalists. Niche breadth was calculated using Levins' method (Levins, 1968) and standardized using Hurlbert's method (Hurlbert, 1978). Levins' standardized niche breadth equation is as follows:

293

294 *(Equation 2)* 

295

$$B_A = \frac{\frac{1}{\sum p_j^2} - 1}{n - 1}$$

where:

297

298  $B_A$  = Levins' standardized niche breadth

299  $p_i$  = proportion of individuals using resource state j

 $300 \quad j = \text{resource state}$ 

301 n = number of possible resource states

302

Levin's standardized index produces niche breadths ranging in value from 0-1, with species exhibiting a value < 0.3 considered to be specialists and values of 0.3-1 considered to be generalists (Nagelkerken, Van Der Velde, Wartenbergh, Nugues, & Pratchett, 2009; Novakowski, Hahn, & Fugi, 2008; Sa-Oliveira, Angelini, & Isaac-Nahum, 2014). A food 307 specialist consumes a narrow range of food sources, while a generalist consumes a wide variety
308 of food sources (Amundsen, Gabler, & Staldvik, 1996). Note that Levins' index does not take
309 into account the possibility of variation in resource abundance and availability.

310

311 Statistical analyses

Statistical analyses were performed using the program Primer-e, except the isospace niche 312 313 areas and overlap analyses which were calculated using the packages rKIN and nicheROVER available in *R* (Albeke, 2017; Lysy et al., 2014). Analysis of variance (ANOVA – type III error) 314 315 was used to test the effects of region ("Re"; North, Middle, South; fixed factor), affinity ("Af"; tropical, temperate; fixed factor), and species ("Sp"; 18 species for isotope statistics, and 7 316 species for all other analyses; random factor) on niche space. All analyses were performed on 317 318 fourth root transformed data, and used either Bray-Curtis or Euclidian resemblance matrices. 319 All residuals were permutated under a reduced model. Where significant differences were found, pairwise tests were used to determine which treatments differed. Because the factor 320 'species' is nested within factor 'affinity', *post-hoc* tests of any significant  $\text{Re} \times \text{Sp}$  (Af) 321 322 interactions yielded only intra-affinity comparisons, e.g., how a particular tropical species differed across regions, or how a temperate species in the North differed from other temperate 323 species in the North. As such,  $\text{Re} \times \text{Sp}$  (Af) *post-hoc* results are not useful in investigating 324 dynamics between tropical and temperate affinities, only changes within each affinity. 325

326

#### 327 Results

328 Niche segregation

Trophic niches of tropical and temperate species were largely segregated (despite their coschooling *in situ*), irrespective of latitude (Figs. 1a, 3). The niche convex hulls showed that overlap between tropical and temperate niche space was minimal (28.4% in the North, 8.8% in 332 the Middle, 15.5% in the South). The negligible overlap between tropical and temperate species was driven primarily by tropical *Abudefduf* species overlapping with temperate sea chubs 333 (Microcanthus strigatus and/or Atypichthys strigatus) in the North and Middle, with Chaetodon 334 species also overlapping temperature niche space in the South, respectively, and a combination 335 of all temperate species in the Middle (Fig. S1). In total, only four tropical-temperate species 336 pairs out of a possible 37 combinations exhibited overlap greater than 60%, above which 337 338 overlap is considered to be significant (Table S3). In the North, the trophic niche of tropical species A. bengalensis was overlapped by M. strigatus 67.5%, yet none of the trophic niches 339 340 of temperate species were overlapped significantly by tropical species. In the Middle, temperate species Parma microlepis was overlapped by tropical species Chaetodon auriga 341 342 (69.7%), but no tropical species was overlapped significantly by a temperate species. In the South, temperate species A. strigatus overlapped tropical species A. sexfasciatus (76.9%) and 343 344 C. flavirostris (98.9%), but similar to the North, no temperate species was significantly overlapped by a tropical species. The considerable overlap between A. sexfasciatus and A. 345 strigatus in the South is particularly notable because they are known to co-school. 346 347 Unsurprisingly, species exhibited considerable overlap with other species of the same affinity (i.e. tropical species overlapping with other tropical species), regardless of region. The niche 348 segregation between affinity groups was caused by temperate species having consistently 349 higher  $\delta^{15}$ N values (indicative of feeding at a higher trophic level; Fig. 3) than tropical species 350 (3-way ANOVA, p < 0.001), but  $\delta^{13}$ C values did not differ (p = 0.077; Table S4a). 351

Pianka's index of diet overlap between tropical and temperate species affinities did not exceed 0.6 in any region, the value above which overlap is considered significant (Fig. S3). Diet overlap between species with tropical and temperate affinities was similar to the intratropical and intra-temperate dietary overlap (Pianka's index, 2-way ANOVA, p = 0.620; Table S4a; Fig. S3), indicating that affinity did not affect diet overlap. In contrast to isotope niche area and Pianka's diet overlap, tropical and temperate species affinities had similar diet compositions (3-way ANOVA, p = 0.538; Table S4a; Fig. 4).

Niche range (i.e. maximum minus minimum values) of trophic levels ( $\delta^{15}$ N; 3-way ANOVA, p = 0.762) and of prey origin ( $\delta^{13}$ C; p = 0.098) did not differ between tropical and temperate fishes, indicating that they feed within similar isotopic boundaries (Table S4a; Fig. S4). Species with tropical and temperate affinities had similarly moderately-broad niche breadths (Levins' index, 3-way ANOVA, p = 0.455; Table S4a; Fig. 5) with most specieslocation combinations showing an index of > 0.3 (Fig. S5), indicative of trophic generalism.

365

**Fig. 3**: Trophic niche of tropical and temperate affinity groups, displayed in  $\delta^{15}$ N and  $\delta^{13}$ C 366 space for North, Middle, and South regions (see Fig. 2) based on convex hulls. Tropical fish 367 368 are indicated with triangles, and temperate fish are indicated with circles. Each marker represents a single fish, and polygons represent convex hulls that encapsulate 100% of data 369 points. The percentage of the temperate niche that is being overlapped by the tropical niche is 370 371 shown for each region. Percent overlap was calculated using the rKIN package available in R. See Fig. S6 for the total niche area size of these convex hulls, and Fig. S1 for standard ellipses 372 of individual species. 373

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Fig. 4: Proportional stomach contents averaged for fish species within tropical and temperate affinity groups in North, Middle, and South regions from fish collected in 2018. Stomach gastric juice was excluded, as was partially-digested, unidentifiable food. The remaining food items were scaled up to 100%. Affinity means were calculated based on the species means in each region. Algae, seagrass, crustaceans, worms, and gastropods are broad groups and may contain multiple families within the taxa. See Fig. S7 for the stomach contents of individual species. **Fig. 5**: Mean ( $\pm$  SE) Levins' index of niche breadth (ranging from 0 to 1) for tropical and temperate species across regions. A low value indicates that a species has a very small niche breadth and is a specialist, and higher values (> 0.3) indicate broad niche breadth, or generalism. Affinity values were calculated using species means at each region. See Fig. S5 for the niche breadths of individual species.

387

#### 388 Niche expansion/contraction

When grouped into affinities, tropical species had the largest niche in the tropics (One Tree 389 390 Island, southern Great Barrier Reef; Figs. 1d, S2, S6a); likewise, the largest temperate species 391 niche area was found at the coldest location (i.e. highest latitude sampled). In the regions where 392 tropical and temperate species geographically overlap (North, Middle, and South), tropical 393 species had the largest niche area where temperate species had their lowest (Middle), with 394 tropical species exhibiting a niche area nearly three times the size of the temperate niche area (10.5 vs 3.8, Fig. S6a). Similarly, temperate species exhibited their largest niche area where 395 396 tropical species exhibited their smallest (South), with the temperate niche almost double the size of the tropical niche (10.9 vs 5.8). When examining species individually (using either 397 398 convex hulls or standard ellipses), affinity-level patterns were less discernible, although temperate species continued to have a large niche area in the South and several tropical species 399 400 a large area in the Middle (Figs. S6b, c). The considerable dichotomy in mean tropical and 401 temperate affinity niche areas in the Middle is primarily due to the large niche areas of tropical 402 species C. auriga and A. vaigiensis (and A. sexfasciatus when using standard ellipses) and the small niche areas of all temperate species, particularly A. strigatus. Both trophic level ( $\delta^{15}N$ ; 403 3-way ANOVA, p < 0.001) and prey origin ( $\delta^{13}$ C; 3-way ANOVA, p < 0.001) signatures 404 differed regionally, with signatures in the tropics being lower in  $\delta^{15}$ N and higher in  $\delta^{13}$ C than 405 all other regions, suggesting that coral reef fishes were consuming isotopically-different prey 406

items from the far north to the South (Tables S4a, b; Fig. S2a). However, the niche range of  $\delta^{15}N$  (3-way ANOVA, p = 0.624) and  $\delta^{13}C$  (p = 0.645) did not differ between regions for either tropical or temperate species (Table S4a; Fig. S4).

410 Tropical and temperate fish consumed primarily algae, zooplankton, and crustaceans, 411 and overall decreased their consumption of algae and increased their consumption of zooplankton and crustaceans as a function of increasing latitude, i.e. from North to South (Figs. 412 413 4, S7; Table S4a). All tropical species had different diet compositions between the latitude extremes (North and South), with one species (A. vaigiensis) also differing between North and 414 415 Middle regions. Only one temperate species (*M. strigatus*) had a different diet composition between the latitude extremes, yet all differed between the Middle and South regions (Fig. S7; 416 Table S4b). Additionally, temperate species decreased the number of food items consumed 417 418 from the highest latitude (South: 13 items) to the two lower latitudes (North/Middle: 9 and 10, respectively), but tropical species ate a similar number of food items across all three latitudes 419 (9, 8, 9, respectively) (Fig. 4). Neither diet overlap (Pianka's index, 2-way ANOVA, p = 0.702; 420 Fig. S3) nor niche breadth (Levins' index, 3-way ANOVA, p = 0.917; Fig. 5) varied among 421 regions for temperate or tropical species affinity groups. On the individual species level, the 422 tropical species A. vaigiensis and the temperate species M. strigatus varied in niche breadth 423 between regions (Fig. S5; Table S4b). 424

425

#### 426 Discussion

Here we show that the novel co-existence of range-extending tropical and local temperate species in temperate ecosystems under climate change is facilitated by trophic niche segregation (see conceptual diagram Fig. 1a). On average, temperate species had consistently higher  $\delta^{15}$ N values than tropical species in all regions, indicating that temperate species are consuming food sources that are higher up the food chain than the tropical species they co-

occur with. Some individual species exhibited some degree of isotopic niche overlap, although 432 only four out of 37 tropical-temperate species combinations exhibited an overlap in niche space 433 434 of > 60%, above which overlap is considered to be significant. Similarly, there was a low degree of overlap in stomach contents between tropical and temperate species (i.e., based on 435 Pianka's index < 0.6) in all regions. In contrast, stomach content composition revealed that 436 species of different affinities were foraging on broadly similar prey groups. This similarity in 437 438 prey groups is likely due to the low resolution of prey identification from stomachs (e.g. broad categories like 'crustaceans' and 'algae'), which may not reflect taxonomic differences in prey 439 440 species within broader groups consumed. Previous studies have demonstrated that minor prey items (which might not be significantly reflected within bulk stable isotope signatures) can be 441 of great importance in facilitating species co-existence (Koussoroplis et al., 2010; Nagelkerken 442 443 et al., 2009), and that co-existence can occur between species even when there is no apparent 444 partitioning of food resources (Pratchett, 2005). Nevertheless, niche segregation (based on isotopic niche space and Pianka's dietary overlap) was observed across all latitudes studied and 445 indicates that tropical and temperate fishes are therefore less likely to compete for the same 446 prey resources at the initial phases of range-extensions in a warming ocean. 447

The fishes studied here consumed a wide range of prey groups (8-13 per region), 448 suggesting that they are generalists (Roper, 1994). Additionally, affinities exhibited 449 moderately-broad niche breadths (based on Levins' index of > 0.3) across regions, which 450 451 further indicates a certain degree of trophic generalism (Devictor et al., 2010; Hurlbert, 1978; Sa-Oliveira et al., 2014). Generalistic feeding strategies are known to play an important role in 452 spatial partitioning and co-existence among species by minimizing competition for trophic 453 454 space (e.g. pomacentrid fishes) (Eurich et al., 2019). Generalists are thought to have an advantage over specialists, as they are able to consume a wider array of food sources and are 455 therefore better equipped to deal with food shortages (Clavel, Julliard, & Devictor, 2011; 456

Wilson et al., 2008). Additionally, generalists are thought to be less vulnerable to climate 457 change as they have higher trophic niche plasticity and may be better able to accommodate 458 changes in resource availability and habitat (Afonso Silva et al., 2017; Slatyer et al., 2013; 459 Sunday et al., 2015; Travis, 2003). As such, trophic generalists, such as the tropical vagrants 460 and temperate locals that were examined in this study and are known to frequently co-occur or 461 school together under increasing ocean warming (Smith et al. 2018), will likely be the most 462 463 successful feeding strategists, or "winners", under future climate change (Ho, Fu, Sun, Kao, & Jan, 2009; Warren et al., 2001; Wilson et al., 2008). Our findings therefore provide strong 464 465 support that trophic generalism is mediating the co-existence of tropical fishes with various local species in temperate waters under ongoing global warming. 466

Across broad spatial scales (from tropical coral reefs to temperate kelp systems), 467 tropical fishes exhibited a niche contraction with increasing latitude (as per Fig. 1d), whilst at 468 smaller spatial scales (i.e. within their novel temperate ranges) they showed niche expansion 469 where local species experienced niche contraction. In the regions where species of both 470 471 affinities co-occurred, tropical species exhibited their largest niche area in the region where the 472 temperate niche area was the smallest, and vice versa. This suggests that tropical and temperate fish may be unable to maintain large trophic niches simultaneously, although this may simply 473 474 be an artefact of small and unequal sample sizes. Previous studies have suggested that an expanding niche area may be a result of ecological release, or a sudden population increase due 475 476 to the disappearance of limiting factors (Bolnick et al., 2010; Svanbäck & Bolnick, 2007), while others posit that an expansion of niche area may be due to competition (Namgail, Mishra, 477 De Jong, Van Wieren, & Prins, 2009; Schulter, Price, & Grant, 1985). In the case of 478 479 competition, a narrow niche may prove harmful for the local temperate fish, as a previous study has shown that a small niche area and high overlap with non-native species can lead to a decline 480 in native fish populations in a freshwater lake (Córdova-Tapia, Contreras, & Zambrano, 2015). 481

A study of marine snails living along their range edge showed that thermal stress, as is possibly 482 experienced by our tropical species in the South (cool water) and temperate species in the North 483 (warm water), led to increased diet variation and therefore trophic niche expansion (Reddin, 484 O'Connor, & Harrod, 2016), highlighting the effect that range edges may have on feeding 485 plasticity. Our findings highlight that species that extend their ranges from coral reefs with high 486 prey diversity to less-diverse temperate areas may need to adjust to novel environments by 487 488 contracting their trophic niche. However, on smaller spatial scales local (a)biotic conditions may alter trophic dynamics and lead to opposite outcomes in terms of niche expansion versus 489 490 niche contraction. These initially variable and plastic dynamics at the leading edges of range extensions are typical of species invading new environments (irrespective of climate change) 491 492 (Broennimann et al., 2007; Fernández & Hamilton, 2015; Michel & Knouft, 2012), but as oceans continue to warm such spatial refuges where local species can resist invasion are likely 493 494 to slowly disappear with more definite outcomes in terms of which species will become winners versus losers under climate change (Byers, 2002; Hellmann, Byers, Bierwagen, & 495 496 Dukes, 2008; Moyle & Light, 1996).

497 It is important to note that stable isotope signatures may be confounded by environmental stressors unrelated to changes in diet, potentially leading to inaccurate 498 depictions of trophic niche space (Karlson, Reutgard, Garbaras, & Gorokhova, 2018). A study 499 500 of freshwater snails showed that high temperatures, such as those experienced by temperate species along their trailing range edge, may enrich  $\delta^{15}$ N and  $\delta^{13}$ C values (Ek, Karlson, Hansson, 501 502 Garbaras, & Gorokhova, 2015). Other factors that may contribute to differential incorporation of stable isotopes include body size, growth rate, and protein turnover (Martínez del Rio, Wolf, 503 504 Carleton, & Gannes, 2009). Because tropical species are not yet established in the southern 505 study sites, fish were caught opportunistically, making it difficult to control for body size. 506 Temperature extremes may affect metabolic rate (Clarke & Johnston, 1999; Gillooly, Brown, West, Savage, & Charnov, 2001), causing altered stable isotope tissue incorporation to increase for reasons unrelated to food intake (Carleton & Rio, 2005). For these reasons, other factors might have also partially contributed to some of the variability in trophic niche segregation as displayed by our study species.

511

#### 512 Conclusions

513 How tropical and temperate species fill and share trophic niches within novel mixed communities resulting from global change is a key determinant of future species persistence 514 515 and biodiversity. We demonstrate that range-extending tropical and local temperate fish species exhibit segregated trophic niches, mediated by trophic generalism and local-scale dietary 516 plasticity. We conclude that range-expanding species adjust their trophic niches at their leading 517 edges and occupy open niche space thereby avoiding significant dietary overlap with local 518 species, despite operating under suboptimal environmental and biotic conditions that are 519 characteristic of novel ranges at the initial stages of climate change. 520

521

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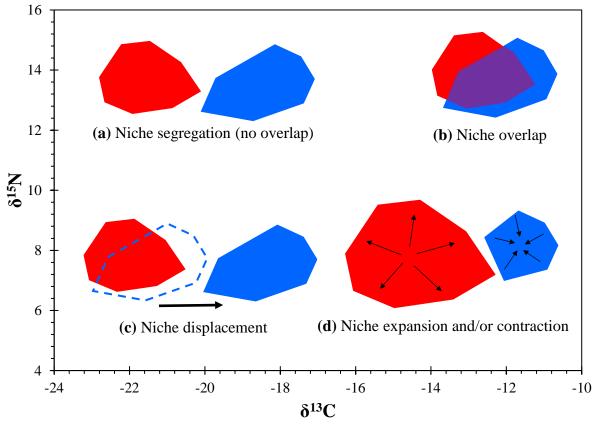


Fig. 1

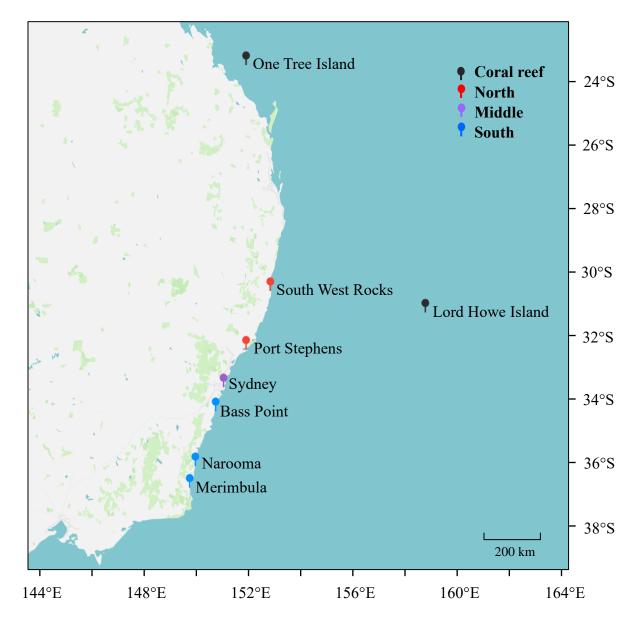


Fig. 2

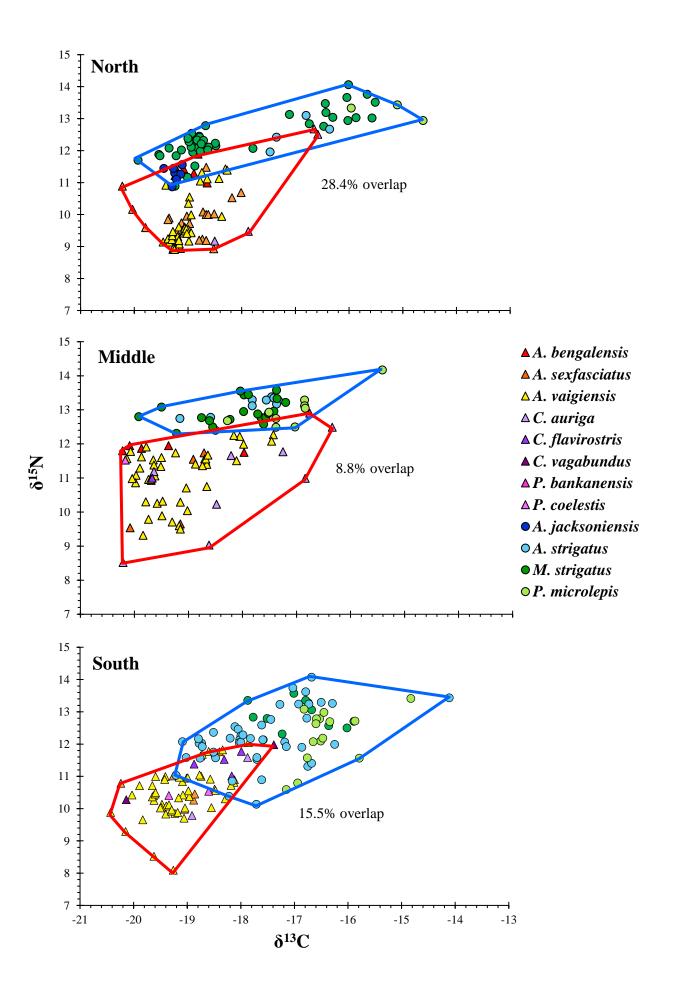


Fig. 3

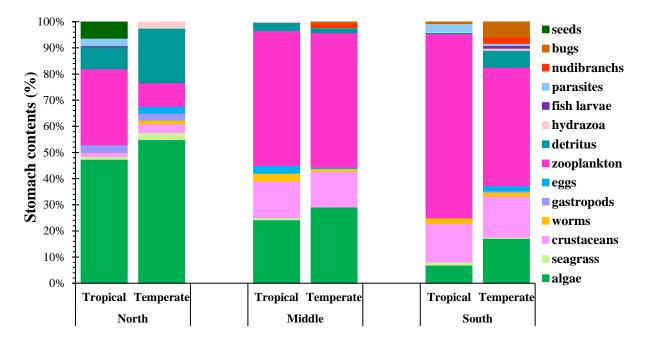


Fig. 4

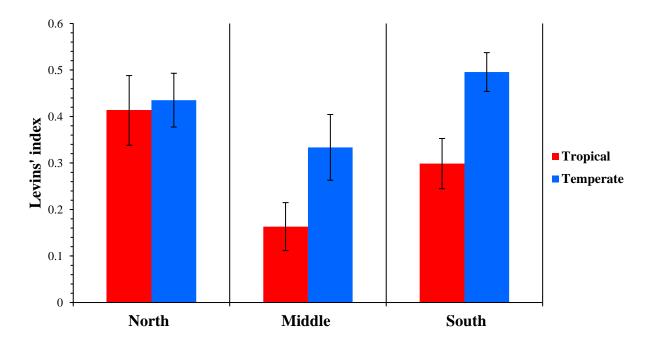


Fig. 5