# EFFECTS OF WATER STRESS AND PARTIAL SOIL-DRYING OF SENESCENCE OF SUNFLOWER PLANTS

by

# Sathaporn Wongareonwanakij B.Sc.(Ag.) (CMU, Thailand)

Department of Horticulture, Viticulture & Oenology, Waite Agricultural Research Institute, The University of Adelaide, South Australia

Thesis submitted for the degree of Master of Science (Agriculture)

i n

The University of Adelaide (Faculty of Agricultural and Natural Resource Sciences)

August, 1995

## LIST OF CONTENTS

## Page

SUMMARY		v
STATEMENTvi		
ACKNOWLEDGM	ENTS v	111
LIST OF FIGUE	RES	İX
CHAPTER I:	GENERAL INTRODUCTION	1
CHAPTER II:	LITERATURE REVIEW	4
1 Leaf senes	cence	5
1.1 Intro	duction	5
1.2 The g	eneral metabolic changes in leaf senescence	6
1.2.1	Decline in photosynthetic rate	7
1.2.2	Loss of chlorophyll	8
1.2.3	Loss of protein content	12
1.3 The re	egulation of senescence	16
1.3.1	Nutrient redistribution	16
1.3.2	Hormonal control	17
	1.3.2.1 Senescence promoter hormones	17
	1.3.2.2 Senescence retardant hormones	18
1.3.3	Stomatal closure	19
2 Water stre	ss and its effect on leaf senescence	20
2.1 Intro	duction	20
2.1.1	Water status parameters	20
2.1.2	Possible mechanisms underlying plant responses to water stress	22
2.2 Metal	polic changes in response to water stress	24
2.2.1	Photosynthesis	24
2.2.2	Chlorophyll catabolism	25

			2.2.3	Protein metabolism	26
			2.2.4	Proline accumulation	27
		2.3	Compa	arison of water stress-induced senescence and natural senescence	ə 30
			2.3.1	Senescence induced by water stress	30
			2.3.2	Comparison of the symptoms of senescence in water stressed and	ł
				naturally senescing leaves	31
				2.3.2.1 Photosynthesis	31
				2.3.2.2 Chlorophyll catabolism	32
				2.3.2.3 Protein metabolism	33
	3	No	n-hydra	aulic root-signals regulating shoot responses	34
		3.1	Introd	duction	34
		3.2	Evide	nce of non-hydraulic root-signals regulating shoot responses	34
			3.2.1	Indirect evidence for non-hydraulic root signals	34
			3.2.2	Direct evidence of non-hydraulic root-signals	36
		3.3	Nature	e of chemical signals passing from roots to shoot	37
			3.3.1	Negative signals	37
			3.3.2	Positive signals	38
			3.3.3	Accumulative signals	40
CHAPTER		R	111:	MATERIALS AND METHODS	41
	1	Pla	Int grow	wth, growth environment and treatment application	42
	2	Pla	nt mea	surements	42
		2.1	Leaf	sampling and water potential determination	42
		2.2	Solub	ble protein content determination	43
		2.3	Total	amino acid content determination	44
		2.4	Prolir	ne content determination	45
		2.5	Chlor	rophyll content determination	46
	3 Soil measurements			46	
	4	Da	ita anal	lysis	47

ii

CHAPTER IV:	RESULTS AND DISCUSSION	48
1 Effects	of leaf water stress on mature leaf senescence	49
1.1 In	troduction	49
1.2 Ma	aterials and methods	49
1.2	2.1 Plant growth and growth environment	49
1.:	2.2 Imposition and relief of water stress	50
1.2	2.3 Plant and soil measurements	50
1.3 Re	esults	51
1.:	3.1 Soil water potential	51
1.:	3.2 Leaf water potential	51
1.:	3.3 Total soluble protein content	54
1.:	3.4 Total amino acid content	56
1.:	3.5 Total chlorophyll content	58
1.	3.6 Chlorophyll a/b ratio	60
1.4 D	Discussion	62
1.	4.1 Water potential responses	62
1	4.2 Senescent responses	63
2 Effects	s of partial soil-drying on mature leaf senescence	67
2.1 lr	ntroduction	67
2.2 M	aterials and methods	68
2.	2.1 Plant growth and growth environment	68
2.	2.2 Root division and treatment application	68
2.	2.3 Plant and soil measurements	69
2.3 R	esults	69
2.	3.1 Soil water potential	69
2.	3.2 Leaf water potential	71
2.	3.3 Total soluble protein content	71
2.	3.4 Total amino acid and proline contents	74
2.	3.5 Total chlorophyll content	76

iii

	2.3.6	Chlorophyll a/b ratio	78
2.4	Discu	ission	78
3 All	eviation	of the effects of a root-signal on leaf senescence by root-excision	82
3.1	Introd	duction	82
3.2	Metho	ds	82
3.3	Resul	ts	83
	3.3.1	Leaf water potential and proline content	83
	3.3.2	Soluble protein content	85
	3.3.3	Total chlorophyll content and chlorophyll a/b ratio	87
3.4	Discu	ssion	90
CHAPTER	IV:	GENERAL DISCUSSION	92

BIBLIOGRAPHY

ĩν

98

#### SUMMARY

The present thesis aimed to investigate the symptoms of leaf senescence in response to plant shoot water stress and explore the possibility of the involvement of a non-hydraulic root signal in the senescence response of mature leaves of sunflower.

The effect of plant leaf water status on mature leaf senescence of sunflower was evaluated from changes in the leaf water potential and soluble protein, total free amino acid and chlorophyll contents, and chlorophyll a/b ratio, following a regime of a single water stress cycle (15 days) and resumption of adequate water supply. The fall in leaf water potential accelerated the rate of loss of leaf soluble protein content compared to that occurring in non-stressed senescing leaves. In corresponding to the breakdown of soluble protein there was an accumulation of free amino acid,5 including a presumed proline accumulation. This phenomenon is different from that which occurred in attached naturally senescing leaves, in which the total amino acid content was found to decrease in company with the fall in leaf soluble protein content.

The rate of loss of chlorophyll content was also accelerated by water stress. Chlorophyll degradation in response to water stress, however, was slower than the protein response. A similar pattern of response to that in total chlorophyll content was also achieved in the chlorophyll a/b ratio.

Upon re-watering, there was a recovery in the protein content to the level found in control leaves coupled with a rapid disappearance of the total free amino acid accumulated. Total chlorophyll content and chlorophyll a/b ratio, on the other hand, did not recover to the control levels after watering was resumed. However, there was a change in the rate of decline of both total chlorophyll content and chlorophyll a/b ratio to the normal senescence rate, subsequent to the recovery in leaf water potential.

V

To examine the possibility of an effect of a root-sourced signal on leaf senescence, plants were grown with the root system divided equally between two containers. One half of the root system was exposed to drying soil by withholding water from this half of the soil, whereas the other half of the soil was well-watered to maintain the leaf water status. Metabolic changes in mature leaves of these plants following six weeks of the partially soil-drying imposition were determined in comparison to those in well-watered plants. The loss of leaf soluble protein content showed a significant response to the effect of treatment from week 4 of exposing the plant to soil-drying in part of the root system. However, there was no significant response in leaf total chlorophyll, free amino acid and proline contents, but a slight response, when summed across time and leaf position, in chlorophyll a/b ratio compared to that in the control leaf.

To investigate whether this increase in the leaf protein loss rate in response to soil-drying could be attributed to reductions in nutrient supply or undetectable changes in leaf water status as a result of the loss of half the nutrient or water gathering capacity, the half of the root system in drying soil was excised after four weeks of exposure to drying soil. This treatment was followed by an increase in leaf protein content, indicating relief from the effects of roots growing in dry soil.

The results are interpreted to demonstrate an effect of a non-hydraulic root signal on senescence of the leaves. The alleviation of the leaf soluble protein loss rate by the excision of the root system in drying soil also indicates that this signal originates in roots in dry soil and acts to promote protein loss.

vi

### STATEMENT

I hereby declare that the thesis here presented contains no material which has been accepted for the award of any other degree or diploma in any University and that, to the best of my knowledge and belief, this thesis contains no material previously published or written by another person, except where due reference is made in the text of the thesis.

I give consent to this copy of my thesis, when deposited in the University Library, being available for loan and photocopying.

(Sathaporn Wongareonwanakij)

### ACKNOWLEDGMENTS

I would like to express special gratitude to Dr. D. Aspinall and Professor M. Sedgley, my supervisors for their valuable suggestions and encouragement. In particular, my thanks are chiefly due to Dr. D. Aspinall for his great patience in discussing my work and his courteous help to shape and improve the manuscript during the various stages of the writing. I also wish to thank Ms. L. Giles and Ms. R. Middelberg for their advice in statistical analysis. My thanks are also extended to all staff and members of the Department of HVO for being very nice, friendly, and helpful.

I would also like to acknowledge AusAID and the Thai Government for awarding me a scholarship for this study.

### LIST OF FIGURES

Fig. 1	The relationship between soil water potential and soil water	
	content (%).	47
Fig. 1.1	Changes with time in water potential of the soil in which	
	well-watered and water-stressed sunflower plants were	
	rooted.	52
Fig. 1.2	Changes with time in leaf water potential of well-watered and	
	water- stressed sunflower plants at 3 different leaf positions:	
	(a) leaf 3; (b) leaf 5; and (c) leaf 7 from the base.	53
Fig. 1.3	Changes with time in leaf soluble protein content of well-	
	watered and water-stressed sunflower plants at 3 different	
	leaf positions: (a) leaf 3; (b) leaf 5; and (c) leaf 7 from the	
	base.	55
Fig. 1.4	Changes with time in leaf total amino acid content of well-	
	watered and water-stressed sunflower plants at 3 different	
	leaf positions: (a) leaf 3; (b) leaf 5; and (c) leaf 7 from the	
	base.	57
Fig. 1.5	Changes with time in leaf total chlorophyll content of	
	sunflower: (a) in well-watered and water-stressed plants and	
	(b) at leaves 3, 5 and 7 from the base.	59
Fig. 1.6	(a) Changes with time in leaf chlorophyll a/b ratio of well	
	watered and water-stressed sunflower plants. (b) Average	
	chlorophyll a/b ratio of leaves 3, 5, and 7 from the base.	61
Fig. 2.1	Changes with time in water potential of well-watered and	
	drying halves of the soil in which sunflower plants were	
	grown with divided rooted systems.	70

Page

ix

- Fig. 2.2 Comparison of water potential of leaves 3, 5 and 7 (from the base) at different ages of well-watered and partial soildrying sunflower plants growing with divided root systems.
- Fig. 2.3 Comparison between the soluble protein contents of leaves 3, 5 and 7 (from the base) at different ages of well-watered and partial soil-drying sunflower plants growing with divided root systems.
- Fig. 2.4 Comparison on total amino acid and proline contents in leaves 3, 5 and 7 (from the base) of different ages of well-watered and partial soil-drying sunflower plants growing with divided root systems.
- Fig. 2.5 Comparison on total chlorophyll content in leaves 3, 5 and 7 (from the base) of different ages of well-watered and partial soil-drying sunflower plants growing with divided root systems.
- Fig. 2.6 Comparison on chlorophyll a/b ratio in leaves 3, 5 and 7 (from the base) of different ages of well-watered and partial soil-drying sunflower plants growing with divided root systems.
- Fig. 3.1 Water potential of leaves 5, 7 and 9 (from the base) at different ages of well-watered, partial soil-drying and partial root-excised sunflower plants growing with a divided root system.
- Fig. 3.2 Changes in soluble protein content in leaves 5, 7 and 9 (from the base) at different ages of well-watered, partial soildrying and partial root-excised sunflower plants growing with a divided root system.

Х

72

73

75

77

79

86

84

- Fig. 3.3 Changes in total chlorophyll content in leaves 5, 7 and 9 (from the base) at different ages of well-watered, partial soil-drying and partial root-excised sunflower plants growing with a divided root system.
- Fig. 3.4 Changes in chlorophyll a/b ratio in leaves 5, 7 and 9 (from the base) at different ages of well-watered, partial soildrying and partial root-excised sunflower plants growing with a divided root system.

89

88