

The Responses of Maize Roots to Nitrogen Supply

By

Kasra Sabermanesh

Thesis submitted in fulfilment of the requirements for the degree of

Doctorate of Philosophy in the Faculty of Sciences at

The University of Adelaide

Australian Centre for Plant Functional Genomics, Adelaide August 2014

The Responses of Maize Roots to Nitrogen Supply

By

Kasra Sabermanesh

Supervised by:

Associate Prof Sigrid Heuer Senior Research Scientist Australian Centre for Plant Functional Genomics The University of Adelaide

Dr Trevor Garnett Research Scientist The Plant Accelerator The University of Adelaide

Dr Darren Plett Research Scientist Australian Centre for Plant Functional Genomics The University of Adelaide

Prof Mark Tester Centre for Desert Agriculture Division of Biological and Environmental Sciences and Engineering King Abdullah University of Science and Technology Kingdom of Saudi Arabia Thesis submitted in fulfilment of the requirements for the degree of

Doctor of Philosophy

School of Agriculture, Food and Wine Faculty of Science, The University of Adelaide Waite Research Institute, Glen Osmond, SA 5064 Email: kasra.sabermanesh@acpfg.com.au

Declaration

I certify that this work contains no material which has been accepted for the award of any other degree or diploma in my name, in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text. In addition, I certify that no part of this work will, in the future, be used in a submission in my name, for any other degree or diploma in any university or other tertiary institution without the prior approval of the University of Adelaide and where applicable, any partner institution responsible for the joint-award of this degree.

I give consent to this copy of my thesis, when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1986.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library Search and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Kasra Sabermanesh

August, 2014

Acknowledgements

The successful completion of this dissertation was made possible by the invaluable contribution of numerous people. Firstly, I would like to offer my deepest gratitude to my supervisors Dr Trevor Garnett and Dr Darren Plett for their continuous contribution towards my training, development, and support over the course of this project. You two have truly made this PhD an experience I have absolutely enjoyed. Assoc Prof Sigrid Heuer and Prof Mark Tester, your expertise along the way have been very insightful and have driven some stimulating discussions. Dr Julian Taylor, your assistance along the way has been absolutely tremendous, and I cannot give enough thanks. I would like to thank the entire Nitrogen Use Efficiency group for their technical assistance and support along the way. I must thank you Hanne Thompson for helping me with my harvests, particularly for those falling on the weekend. To Dr Malcolm Hawkesford, Astrid Grün and Rothamsted Research Institute, thank you all for the support for my research project in the United Kingdom; I look forward to crossing paths with you all again soon.

I gratefully thank the University of Adelaide, the Australian Centre for Plant Functional Genomics (ACPFG), and the Grains Research and Development Corporation (GRDC) for their financial support over the candidature.

I would like to thank my mother for her encouragement along the way. To my brother and my friends, I would like to acknowledge you all for the friendship you have given me along the way. To my partner Lucy, thank you for the endless love and support you have given me during this PhD, you have stuck by me through the best and worst of times. This will be returned when you are writing your dissertation, I promise.

Table of Contents

Declaration	ii i
Acknowledgements	iv
Table of contents	V
Abstract	vii
List of abbreviations	ix
Chapter 1: Literature review	
1.1 Cereal crop production	
1.2 Plant nitrogen (N) nutrition	
1.3 N in soils	
1.4 N fertiliser use	2
1.5 Cereal N use efficiency	
1.5.1 Improving NUE	6
1.5.1.1 Optimising agronomic practises	6
1.5.1.2 Improving N uptake efficiency	7
1.5.1.2.1 Improving the N uptake system	7
1.5.1.2.2 Optimising root morphology	7
1.6 Root N uptake	9
1.6.1 Root NO_3^- uptake	9
1.6.2 Regulation of NO ₃ ⁻ uptake	13
1.6.3 NO_3^- storage and assimilation	15
1.7 Adapting to N limitation	16
1.7.1 Increasing NO ₃ ⁻ uptake capacity	17
1.7.2 N remobilisation	17
1.7.3 Modifying root morphology	
1.7.4 Changing biomass allocation	18
1.8 What is required to improve cereal root N uptake?	20
1.9 Aims and objectives	21
Chapter 2: The transition from maternal to external nitrogen so	urces in
maize seedlings	
o	

Chapter 3: Responses of maize seedling root morphology to nitrate	
supply	61
Chapter 4: Mapping quantitative trait loci for morphological seedling	g root
traits using the intermated B73 × Mo17 mapping population, relative	to
nitrate supply	90
Chapter 5: General discussion	118
5.1 Advances in knowledge from this study	118
5.2 Future directions	122
Chapter 6: Literature cited (Literature review & general discussion)	126

NOTE: Statements of authorship appear in the print copy of the thesis held in the University of Adelaide Library.

Abstract

Substantial quantities of costly nitrogen (N) fertilisers are applied to cereal crops each year to maximise yields, but only approximately half of the N is captured by cereals, providing scope to increase root N uptake. However, our understanding of how the nitrate (NO_3^-) uptake system is regulated and how it could be improved is limited. Furthermore, the changes to root morphology in response to NO_3^- supply are not well understood, in this case due to the difficulties associated with phenotyping roots in soil.

To investigate how the NO₃⁻ uptake system is up-regulated, maize (*Zea mays* var. B73 and Mo17) was grown hydroponically with low or sufficient NO₃⁻ supply, and a range of physiological parameters associated with NO₃⁻ uptake were measured across the transition from seed N use, to external N capture. This transition provides an ideal system to clarify how the NO₃⁻ uptake system up-regulates as this is when plants first rely on increasing root N capture to meet demand. Across both lines and treatments, concentrations of shoot N and free amino acids in roots and shoots rapidly decrease as seed N reserves exhaust. Once free amino acid concentrations decrease to a critical level, root NO₃⁻ uptake capacity rapidly increased, corresponding with a rise in transcript levels of putative NO₃⁻ transporter genes *ZmNRT2.1* and *ZmNRT2.2*. As NO₃⁻ uptake capacity reached maximum levels, shoot N concentrations stabilised. Despite shoot N concentrations stabilising, B73 was unable to maintain net N uptake and shoot growth in low NO₃⁻, relative to sufficient NO₃⁻. Conversely, Mo17 maintained shoot growth and net N uptake, and increased root mass in low NO₃⁻ relative to sufficient NO₃⁻. The effects of NO₃⁻ limitation on growth were reflected by an increased root:shoot, which emerged just prior to shoot N concentrations stabilising.

In order to understand how root morphology may reflect the NO_3^- treatments differences observed in growth and net N uptake, morphological root traits were quantified

across seedling development. Analysis showed that although B73 achieved greater absorption area per unit root mass than Mo17, its morphology was unresponsive to NO_3^- supply. Conversely, Mo17 responded to NO_3^- limitation by increasing lateral and axial root length before increasing root mass or volume. Subsequently, 11 putative quantitative trait loci (QTL) associated with morphological root traits corresponding with shoot growth or N uptake were detected across low and sufficient NO_3^- , with one major QTL for lateral root length and surface area being identified in low NO_3^- on chromosome 5.

These results provide insight into the processes involved in up-regulating root NO_3^- uptake capacity and how root morphology can adapt to NO_3^- supply. These findings identify potential control points in the regulation of NO_3^- uptake capacity and root morphology, which may be investigated further via global transcriptional analysis or fine-mapping of identified QTL respectively. Ultimately, this work may lead to identification of candidate regulatory genes that could be either manipulated to generate new lines with enhanced N uptake efficiencies, or allow the identification of germplasm with this trait.

List of Abbreviations

analysis of variance
average root diameter
average lateral root length
average lateral root surface area
average lateral root volume
average seed mass
axial root
boron
best linear unbiased predictors
carbon
calcium
chloride channel
copper
day
days after imbibition
dry-weight
ethylenediamine-N,N'-bis(2-hydroxyphenylacetic acid)
Ethylenediaminetetraacetic acid
iron
figure
fresh-weight
glutamine
glutamate
glutamate synthase
glutamine synthetase
hour
hectares
high-affinity transport system
intermated B73 x Mo17
IBM centimorgans
intermated recombinant inbred line

Κ	potassium
kg	kilograms
kg	kilogram
LATS	low-affinity transport system
LOD	logarithm of odds
LR	lateral root
Mg	magnesium
min	minute
Mn	manganese
Мо	molybdenum
MQ	milli-Q
Mt	megatonne
Ν	nitrogen
NAOH	sodium hydroxide
NAR	nitrate assimilation related
$\mathrm{NH_4}^+$	ammonium
NiR	nitrite reductase
NO ₂ ⁻	nitrite
NO ₃ ⁻	nitrate
NPF	nitrate transporter 1/peptide transporter family
NR	nitrate reductase
NRT	nitrate transporter
NRT	nitrate transporter
NUE	nitrogen use efficiency
NUpE	nitrogen uptake efficiency
NUtE	nitrogen utilisation efficiency
Р	phosphorous
PTR	peptide transporter
Q-PCR	quantitative polymerase chain reaction